

Research Paper

Botany

In Vitro Seed Treatment of Fungicides for the Control of Seed Borne Fungi of Soybean Variety Durga

L. R. Rathod

Dept. of Botany Mahatma Phule A. S. C. College, Panvel. Dist. Raigad

N. B. Pawar

Dept. of Botany Mahatma Phule A. S. C. College, Panvel. Dist. Raigad

Soybean (Glycine max L.) are associated by several seed borne pathogens. These fungi were detected by the methods **ABSTRACT** as recommended ISTA. Fungi such as Aspergillus flavus, Aspergillus niger, Aspergillus fumigatus, Alternaria tenuis, Fusarium oxysporum, Penicillium notatum, Sclerotium rolfsii, Mucor mucedo Curvularia lunata Cladosporium herbarum and Drechslera oryzae, and Rhizopus stolonifer were isolated from the Soybean variety Durga. Eeffect of different fungicides like Thiram, captan and Copper oxycholoride on the incidence of seed borne fungi and their effect on seed germination were evaluated. The seed treatment by the fungicides showed that Copper oxycholoride increased the germination percentage and reduced seed mycoflora. Thus the seeds are treated with fungicides to eliminate the seed borne mycoflora of Soybean.

KEYWORDS: Soybean seed, Seed borne fungi, fungicides, blotter paper test, Agar- Agar plate method Seed washates

Introduction -

In a world facing problem of malnutrition oil rich crops assume special significance. Obtaining maximum production through all available avenues and protecting adequately what is produced would certainly alleviate the problem. In India, soybean estimated yield is 9.81 mt from 9.21 m ha. In developing countries like India, most of the crops yield is reduced due to pathogenic fungi (Agrios 2005). Soybean is an important host of the Aspergillus flavus, Alternaria tenuis and Fusarium oxysporum which causes Seed rot, Seedling blight, Root rot and wilt diseases . in past , several fungicides have been employed in the control of fungal diseases of crops. Agrochemical like Thiram, captan and Copper oxychloride were used to control a number of diseases caused by fungal pathogens of Soybean. Ibiam et al (2000) and (2006) observed that seed dressing fungicides- Benlate, Apron plus 50 Ds Fernasan- D, Dithane M-45 and Bavistin controlling seed borne fungi of Rice Mane et al. (2010). evaluated various fungicidal Seed Treatment on Seed Mycoflora and Seed Germination During Storage of Sorghum. Even through effective and efficient control of seed borne fungi can be achieved by the use synthetic chemical fungicides, the same cannot be applied to grains for reasons of pesticide toxicity (Harris et al. 2001). It is now realized that chemical fungicides cause serious environmental problems and are toxic to non target organisms. (Anon, 2005). The toxic effect of synthetic chemicals can be overcome, only by persistent search for new and safer pesticides accompanied by wide use of pest control method, which are eco-friend by and effective (Mohana et al. 2011).

Materials and Methods -Collection of seed samples (Cultivars) -

The method described by Neergaard (1973) has been adopted for the collection of seed samples. Accordingly, three random samples of seeds (half Kg each) were collected from oil mills, market places, Oil Seeds Research Station, Latur and Pulses Research Center, Badnapur. (M. S.)

Detection of Seed Mycoflora -

The seed mycoflora was isolated by using different methods such as Standard blotter paper method, Agar plate method, and Seed washates as recommended by International Seed Testing Association ISTA (1966), De Tempe (1970), Neergaard (1997) and Agrawal (1976).

Agar plate method -

In Northern Ireland, Muskkett (1948) first used this method for seed health management. In this method, pre sterilized petriplates were poured with 15 mL of autoclaved Potato Dextrose Agar (PDA). On cooling the medium, ten seeds per plate of the sample to be studied were equidistantly placed aseptically. Pair of sterile white blotter papers of 8.5 cm diameter were soaked in sterile distilled water and were placed in pre-sterilized petriplates of 90 mm diameter. Ten seeds of test sample per petriplate were then placed at equal distance on moist blotter. 400 seeds were used in each experiment. The plates were incubated at $28^{\circ} \pm 2^{\circ}$ C under diurnal conditions. On seventh day of incubation, seeds were first examined under stereoscopic microscope for determining the various fungal growths. The identification and further confirmation of seed borne fungi was made by preparing slides of the fungi.

The frequency of the fungus was calculated by the following formula

No. of seeds containing a particular × 100 Total seeds used

Relative abundance of the fungi was calculated by the formula

No. of seeds colonies of a fungus on seed \times 100 Total no. of colonies of all fungus

Treatments -

Effect of seed treatment fungicide on seed mycoflora, seed germination and vigour index of Soybean seed was studied in - vitro condition. Different concentration of fungicides made were 0.1 to 0.5% and apllied to the seeds of Soybean. The effect on seed mycoflora, seed germination and vigour index was recorded. The vigour index was determined by multiplying the percentage germination with the sum of the root and shoot length. Percent inhibition was recorded bt using the formula given below by Vincent (1947).

Vgour index = (Root length in cm + shoot length in cm) \times germination (%).

Result and Discussion -

Seed borne pathogens of Soybean are responsible for causing various fungi and also reducing yield from 20-22% if infected are planted in the field. Seeds play a vital role in the transmission of plant pathogens causing plant diseases. The pathogen may be externally or internally seed – borne or associated with seeds contaminant.

In Agar plate method the fungi isolated identified were Aspergillus flavus, Aspergillus niger, Aspergillus fumigatus, Alternaria tenuis, Fusarium oxysporum, Penicillium notatum, Sclerotium rolfsii, Mucor mucedo Curvularia lunata Cladosporium herbarum and Drechslera oryzae, and Rhizopus stolonifer. These 11 fungi were isolated from the untreated seeds of variety (Table 1). Similarly from the PDA and seed washates 8-11 were isolated and identified. Similarly from the untreated seed of Blotter paper method, Agar plate method and Seed washates mathod highest frequency 46.00, 50.00 & 40.00 and relative abundance 25.00, 30.00 & 20.00 were recorded from Aspergillus flavus and Aspergillus niger..

Cultiver Durga shows the response to the Copper oxychloride treatment in relation to its seed mycoflora, seed germination and vigour index. The results are given in table 2. It was found that as concentration of Copper oxychloride increases, there was decrease in seed mycoflora where as increase in seed germination and vigour index. At 0.5% of

Copper oxychloride seed treatment it shows minimum seed mycoflora with 00% as compared with control with 70%. At the same concentration there was increase in seed germination which was 90% as compared with control with 55%. At the concentration vigour index was 800 as compared with control with 150.

Table 1. Seed mycoflora of Soybean (Glycine max L.) variety Durga on Blotter, Agar plate and seed washates

Isolated fungi	Blotter paper		Agar plate		Seed washate	Seed washates	
	Frequency	Relative abundance	Frequency	Relative abundance	Frequency	Relative abundance	
Aspergillus flavus	46	25	50	30	40	20	
Aspergillus niger	42	21	46	28	35	30	
Aspergillus fumigatus	21	14	25	17	25	15	
Fusarium oxysporum,	15	10	20	12	13	08	
Alternaria tenuis	09	05	10	07	08	03	
Rhizopus stolanifer	07	4.25	09	06	05	02	
Penicillium notatum	06	04	08	05	03	01	
Mucor mucedo	04	03	05	04	05	00	
Curvularia lunata	03	02	04	03	00	00	
Cladosporium herbarum	02	01	03	01	00	00	
Drechslera oryzae,	01	0.5	0.2	00	00	00	
S.E. <u>+</u>	4.68	2.40	16.40	9.83	13.84	9.70	
C. D at 5%	10.43	5.35	36.57	21.92	30.88	21.63	

00 = absence of fungi

Table No. 2: Effect of Copper oxycholoride seed treatment on seed mycoflora, seed germination and vigour index of Soybean (Cv. Durga)

Conc. (%)	Seed mycoflora (%)	Seed germination (%)	Vigour index
0.00 (Control)	70	55	150
0.1	67	65	180
0.15	65	67	198
0.2	50	70	245
0.25	40	78	290
0.3	25	80	300
0.35	15	83	510
0.4	10	85	650
0.45	05	88	700
0.5	00	90	800
S.E ±	8.15	3.42	72.32
C.D. at 5%	18.41	7.72	163.44

REFERENCES

 $Seed \, Res. \, 4: 24 - 31 \, | \, Agrios, \, G. \, N. \, (2005). \, Plant \, Pathology \, fifth \, edition, \, Elsevlier \, academic \, press. \, 922pp. \, | \, De \, Tempe \, J. \, (1948). \, The \, blotter \, method \, Plant \, Pathology \, fifth \, edition, \, Elsevlier \, academic \, press. \, 922pp. \, Plant \, Plant$ of seed health testing programme. ISTA 2. 133-15. | Harris, C.A. Renfrew, M.J. and Woolridge, M.W. (2001). Assessing the risk of pesticide residues to consumers: recent and future developments. Food Additive. Contam.18 (12): 1124-1129. | ISTA (1976). International Rules of Seed Testing; Aneexes 1976. Seed Sci. & Technol. 4:

3 - 49; pp 50 - 177. | Ibiam, O. F. A., Umechuruba, C. I. and Arinze, A. E. (2000). Field Evaluation of seed -dressing fungicides Bavistin, Benlate Fernasan - D and Apron Plus 50 DS associated with three rice varities Faro 12, Faro 15, Faro 29. Journal of Health and Visual Science. 2. 96 – 106. | Ibiam, O. F. A., Umechuruba, C. I. and Arinze, A. E. (2006). Evaulation of the Efficacy of seed Dressing fungicides (Bavistin, Benlate, Fernasna – D, Apron Plus 50 DS, AND Dithane – M45). In the control of Seed – Borne Fungi of Rice (Oryzae sativa L) Variety Faro 15 In Vitro. Sciencia Africana. 5 (1) 1-10. P. V. Mane, L. R. Rathod G. B. Honna, V. C. Patil & S. M. Muley (2011). Effect of Fungicidal Seed Treatment on Seed Mycoflora and Seed Germination during Storage of Sorghum. Bioscience Discovery, 02 (2):214-216,2011 | Mohana, D.C. Raveesha, K.A. and Lokanath, R.(2008). Herbal remedies for the management of seed-borne fungal pathogens by an edible plant Decalepis hamiltonii (Wight and Arn) Arch. Phytopathol. Plant Protect. 41(1): 36-49. Muskett, A. E. (1948). Technique for the examination of seed for the presence of seed borne fungi. Trans. Brit. Mycol. Soc. 30:74-83. | Muskett J.P., Malone A.E. (1964). Seed-borne fungi. Description of 77 fungus species. Proc. Int. Seed Test. Assoc. 29: 179–384. Neergard, P. (1997). Seed Pathology. Vol. 1. The Macmillan Press Limited, Danist Govt. Institute of seed pathology for developing countries. Copenhagen, Denmark. | Vincent, J.M., (1947). Distortion of fungal hyphae in the presence of certain inhibitors. Nature. 159: 850. |