



PERIODONTAL PERSPECTIVES OF AUTOLOGOUS BLOOD PREPARATIONS

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ABSTRACT

Periodontitis is an infectious disease of attachment apparatus. Untreated periodontitis leads to bone loss and attachment loss. Though several periodontal treatments are available, only some of them are regarded as truly regeneration. Regeneration is reconstitution of both hard and soft tissues in structure and function. Various modalities i.e. bone grafts and substitutes, guided tissue regeneration (GTR) membranes and polypeptide growth factors (PGFs) are used for periodontal regeneration. Platelet concentrates are richest source for polypeptide growth factors. This review highlights various platelet concentrates, and their clinical applications in the treatment of periodontal diseases.

KEYWORDS : Periodontitis, Growth factors, Platelet rich plasma, Fibrin tissue adhesive

Introduction: Periodontitis is an infectious disease causing destruction to periodontal tissues¹. The goal of periodontal therapy is reconstitution of periodontium in structure and function. Periodontal regeneration requires series of biologic events i.e. migration, proliferation and differentiation of cells in the process of wound healing [2]. Platelets play a crucial role in hemostasis and wound healing. The α granules of platelets release platelet-derived growth factor (PDGF), transforming growth factor (TGFβ), and insulin-like growth factor (IGF-I). During activation, α granules fuse with platelet cell membrane releases growth factors. These growth factors bind to transmembrane receptors of target cells (Osteoblasts, fibroblasts, endothelial cells, and epithelial cells) leads to expression of various genes resulting cell proliferation, collagen synthesis and osteoid formation which results formation of soft and hard tissues of periodontium²

Platelet rich plasma (PRP): Marx first used PRP in the mandibular reconstruction defects³. It is a first generation platelet concentrates. It is a by-product of blood that is rich in platelets⁴. It contains platelets, coagulation factors and plasma proteins (Fig.1). A natural human blood clot contains 95% red blood cells (RBCs), 5% platelets, less than 1% white blood cells (WBCs), whereas a PRP blood clot contains 4% RBCs, 95% platelets, and 1% WBCs [4]. It contains the maximum amount of platelets that can release desired growth factors (platelet derived growth factor (PDGF), transforming growth factors-β1 and -β2 (TGF-β1 and -β2) and insulin-like growth factor-1⁵. The increased amount of growth factors will enhance soft and hard tissue healing process⁵. PRP contains Growth Factors (PDGF-AA, PDGF-BB, PDGF-AB & TGF), high concentration of platelets, phagocytes and fibrinogen. [6]. The maturation rate during bone regenerative procedure is increased up to 2.16-times by PRP⁶. The PRP production requires blood collection with anticoagulant, 2 steps of centrifugation, and using of calcium chloride and bovine thrombin for polymerization (Fig.2)⁷

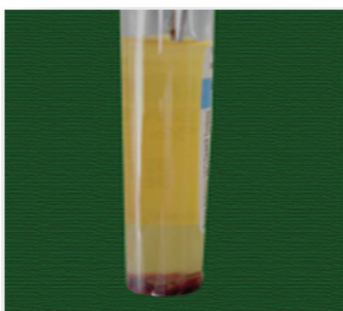


Fig.1: Platelet rich plasma

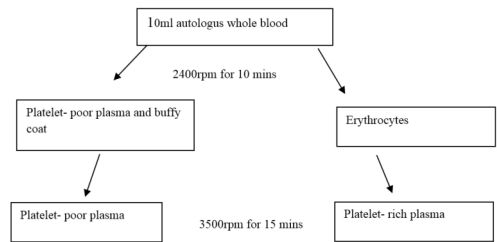


Fig.2: PRP protocol

Clinical implications, advantages, limitations and contraindications of PRP⁸(Fig.3):

Clinical implications	Advantages	Limitations	Contraindications
1.Osseous defects 2. Sinus lift surgeries 3. Augmentation techniques 4. Peri-implant defects 5. Ridge preservation 6. Gingival recession 7.Healing of extraction wound	1. Safe autogenous preparation 2. Blood is collected at the time of preoperational	1. Presence of bovine thrombin which initiates allergic reaction 2. Lack of uniformity in PRP preparation protocols	1.Platelet disorders 2. Local infection at the site 3. Unwilling patient

Studies regard PRP application in different periodontal and implant surgical procedures: (Fig.4):

Treatment of	Positive Studies	Negative Studies
Infrabony defects(RCT)	Piemontese et al.2008 ⁸ ,Kaushick BT et al. 2011 ¹⁰ ,A.R. Pradeep et al.2012 ¹³ ,Menzes et al.2012 ¹⁴],Hassan S et al. 2012 ¹⁵ Kukreja BJ et al.2014 ¹⁶ ,Agarwal .A et al.2014 ¹⁷	Dori et al.2007 ⁷ , Camargo et al.2009 ⁹ , Harnack et al.2009 ¹⁰ , Ozdemir B et al.2012 ¹² , Pinpe J et al. 2014 ¹⁸
Gingival recession(RCT)	Jovovic et al.2013 ²¹	Keceli. H et al.2008 ¹⁹ , LeLafzi A et al. 2010 ²⁰
Sinus augmentation	Aiemetti et al.2008(RCT) ²² Torres et al.2009(RCT) ²⁴ , Khairy M et al. 2013(RCT) ²⁵	Schaff et al.2008(CS) ²⁶

RCT= Randomized clinical trial, CS= Case series

Positive Studies = Statistical significant difference between clinical parameters (PPD, CAL, Bone fill, Increase in keratinized width, root coverage, gingival thickness, bone formation around implant, no mobility of implant) in test and control groups.; **Negative Studies** = Statistical no significant difference between clinical parameters in test and control groups.

Platelet rich fibrin (PRF): It is a second generation platelet concentrate²⁶ It was developed by Choukroun. It is devoid of bovine thrombin which is seen in PRP preparation.^{26,27} The interaction between leukocytic cytokines and fibrin complex play a vital role in the regeneration.

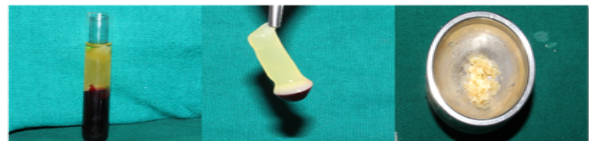
A physiological enriched fibrin complex matrix (PRF) releases growth factors in a controlled manner for longtime when compared to fibrin glue enriched with cytokines.

The preparation of PRF is simple. A blood sample is taken in 10 ml tube without anticoagulant and centrifuged at 3,000 rpm for 10 mins. The coagulation process is natural in test tube as it is devoid of thrombin²⁷. It consists of 3 layers, upper layer-platelet poor plasma, middle layer- Fibrin clot with platelet, bottom layer-RBC (**Fig.5a**).

It contains cytokines such as IL-1, -4, -6, and growth factors such as Transforming Growth Factor beta 1 (TGF-β1), Platelet Derived Growth Factor (PDGF), and Vascular Endothelial Growth Factor (VEGF) [28,29] PRF acts as a powerful scaffold with an integrated reservoir of growth factors for tissue regeneration. The fibrin matrix in PRF acts as natural guide for angiogenesis, natural support to immunity and guides the coverage of wounds (**Fig.5b**).²⁷

Clinical implications, advantages, limitations and contraindications of PRF (Fig.6)²⁶⁻²⁸

Clinical implications	Advantages	Limitations
1. In sinus lift procedures	1. It is completely safe.	1. As it is produced in limited quantities, which limits the utilization in general surgery or extensive surgical procedures.
2. Socket preservations	2. Standard protocol for preparation.	2. PRF membranes are totally specific to the donor and cannot constitute an allogenic graft tissue. So PRF tissue banks are un feasible
3. Intra-bony defects with or without bone grafts (Fig. 5c).		
4. PRF membrane has been used for gingival recession coverage with coronally advanced or lateral pedicle flap for multiple and single recession respectively		
5. Endo perio lesions		
6. Furcation defects		



(**Fig.5a**):Platelet –rich fibrin in test tube (**Fig.5b**): Platelet-rich fibrin (**Fig.5c**):PRF mixed with bone graft

Difference between first and second generation platelet concentrate (Fig.7)²⁷:

Platelet rich plasma (PRP)	Platelet rich Fibrin (PRF)
First generation platelet concentrate	Second generation platelet concentrate
Use of anticoagulants	No anticoagulants used
Fibrin polymerization is depends on the thrombin and calcium chloride and polymerization process is rapid.	Polymerization starts on contact with glass particles of the test tube which results in physiologic thrombin formation polymerization process is slow.
3-D organization of a fibrin network-condensed to tetra molecular structure which leads to a rigid network, not very favorable to cytokine enmeshment and cellular migration	3-D network-connected tri molecular allows the establishment of a fine and flexible fibrin network and able to support cytokines enmeshment and cellular migration
It can firmly seal biologic tissues because of gel in consistency	It can act as membrane because of it's elasticity and flexibility

Studies regard PRF application in different periodontal and implant surgical procedures (Fig.8):

Treatment of	Positive Studies	Negative Studies
Infrabony defects(RCT)	M Thorat et al. 2011 ³⁰ .V. Rosamma Joseph et al.2012 ³¹ A.R .Pradeep et al.2012 ¹³ Chhya Bansal et al.2013 ³² Ajwani H et al.2015 ³⁵ ,Agarwal .A et al. 2015 ³⁶ ,A.R .Pradeep et al.2015 ³⁷ Gupta SJ et al.2015 ³⁸	Mathur A et al.2015 ³³ ,Shah M et al.2015 ³⁴
Gingival recession(RCT)	Del corso M et al.2009 ³⁹ Sofia Aroca et al.2009 ⁴⁰ Padma R et al.2013 ⁴¹ , Tunali M et al.2015 ⁴³	Jankovic et al.2012 ⁴¹ , Eren G et al.2014 ⁴³ , Thamaraiselvin et al.2015 ⁴⁵
Sinus augmentation	Toffler M et al.20109(Early report of 110 pts) ⁴⁶	Zhang et al.2012(CS) ⁴⁷
Grade II furcation defects(RCT)	Sharma et al.2011 ⁴⁸ Bajaj P et al. 2013 ⁴⁹	-
Post extraction socket filling(RCT)	Hauser F et al.2013 ⁵⁰	-
Ridge preservation(RCT)	Barone A et al.2014 ⁵¹	-
Peri implant bone defects(RCT)	Hamzacebi.B et al.2015 ⁵²	-

RCT= Randomized clinical trial, CS= Case series

Positive results = Statistical significant difference between clinical parameters (PPD, CAL, Bone fill, Increase in keratinized width, root coverage, gingival thickness, bone formation around implant, no mobility of implant) in test and control groups.; **Negative results**= Statistical no significant difference between clinical parameters in test and control groups.

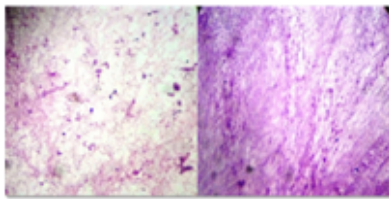
Titanium-prepared platelet-rich fibrin (T-PRF):

Some authors are worried about glass-evacuated blood collection tubes with silica particles as these particles may cause health hazards⁵³. Only small fraction of these silica particles are sedimenting with red blood cells. Majority of silica particles suspends in a buffy coat so that these particles reach to patient when these product is used for treatment⁵⁴. Although this is not practically concluded (the architecture of L-PRF change with type of material used for its preparation), some of the authors used more biocompatible material titanium for PRF preparation (**Fig.9**).⁵⁵ Although basic histological structure similar between T-PRF and L-

PRF, there is some difference in fibrin structure in T-PRF. The fibrin of T-PRF is more woven and thicker when compare with L-PRF. The difference may be due to the biocompatibility and hemocompatibility of titanium, which led to the formation of a more polymerized fibrin (Fig.10, 11). [55]. More research is required on T-PRF in terms of absorption time in body and clinical advantage over L-PRF.

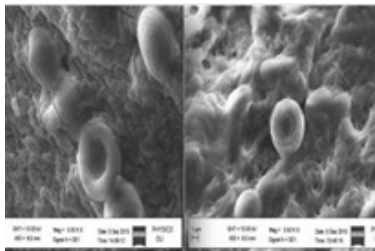


(Fig.9): Titanium test tubes for T-PRF preparation



L-PRF T-PRF

(Fig.10): Histological analysis shows more number of fibroblasts and thicker fibrin structure in T-PRF when compares to L-PRF



L-PRF T-PRF

(Fig.11): SEM analysis shows more woven and thicker fibrin structure in T-PRF when compares to L-PRF

Advanced Platelet rich fibrin (A- PRF):

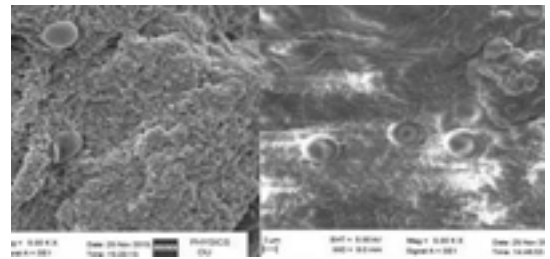
The centrifugation protocol is 1500 rpm 14 mins. Later it was modified to 1300 rpm 14 mins. It is based on the lower centrifugation protocol. Besides Platelets, Macrophages also produce growth factors. With the lower centrifugation protocol, it was proved that presence of macrophages in Advanced platelet rich fibrin. PRF clots formed with A-PRF centrifugation protocol showed a loose structure with more interfibrinous space, and more cells in distal part of fibrin clot (Fig-12). More research is needed to find the effect of APRF on Periodontal Regeneration.⁵⁶

Advanced Platelet rich fibrin (A- PRF) +:

The centrifugation protocol is 1300 rpm 8 mins. It is also based on lower centrifugation protocol. More research is needed to find the effect of APRF+ on Periodontal Regeneration.⁵⁷

Injectable-PRF:

The centrifugation protocol is 700 rpm 3 mins. This liquid PRF mix with bone grafts results steaky bone which uses for augmentation procedures. I PRF provides longer release of growth factors. I PRF releases growth factors even after 10 days. I-PRF demonstrated the ability to release higher concentrations of various growth factors and induced higher fibroblast migration and expression of PDGF, TGF-β, and collagen1 molecules which helps in regeneration.⁵⁷



L-PRF A-PRF

Fig.12: SEM analysis shows more a loose structure with more interfibrinous space in A-PRF when compares to L-PRF

CONCENTRATED GROWTH FACTORS (CGF): PRF uses constant centrifugation (2700 rpm 12 mins) speed, while CGF (Concentrated growth factors) utilizes altered centrifugation speed (2400-2700 rpm 12 mins) which leads to production of much larger, denser and richer fibrin matrix containing higher amount of growth factors⁵⁸

STICKY BONE: The centrifugation protocol of autologous fibrin glue (AFG) is 2400-2700 rpm 2 mins. Less centrifugation time leads to availability of more growth factors. Sohn et al. 2010 fabricated-growth factors enriched bone graft matrix and called it as Sticky bone. Mixing of AFG (Autologous fibrin glue) to allo graft or to mixture of allo graft and xenograft produce Yellow sticky bone. Addition of exudates from CGF (Concentrated growth factors) is added to the above mixture leads to red color sticky bone formation. Uncoated tube uses for preparation of AFG⁵⁸

BRIEF SUMMARY ON CENTRIFUGATION PROTOCOL OF VARIOUS PRF:

Various PRF	Centrifuge protocols
L-PRF	2700 rpm 12 mins
T-PRF	2700 rpm 12 mins
A-PRF	1500 rpm 14 mins
A-PRF(Modified)	1300 rpm 14 mins
A-PRF+	1300 rpm 8 mins
I-PRF	700 rpm 3-4 mins
CGF	2400 -2700rpm 12 mins
AFG	2400-2700 rpm 2 mins

Conclusion:

The preparation of platelet concentrates and clinical usage is simple and cost effective when compares to other regenerative materials i.e. GTR (guided tissue regeneration) membranes, EMD (enamel matrix derivatives), bone grafts and substitutes. Most of the studies are showed that platelet concentrates are positive edge (either additive or alone) over other regenerative treatments i.e. GTR (guided tissue regeneration) membranes, EMD(enamel matrix derivatives), bone grafts and substitutes in periodontal regeneration .Despite the evidence of clinical advantage of these preparations, evidence of their beneficial effects is still lacking. Hence large and long term follow up randomized clinical trials are required for the determining the full effect of these preparations. They have been used for surgical procedures as they provide consistent benefits for the patient

REFERENCES

- Polimeni G, Xiropaidis AV, Wikesjo UM. Biology and principles of periodontal wound healing/regeneration. Periodontol 2000 2006;41:30-47.
- Pradeep AR, Shetty SK, Garg G, Pai S. Clinical effectiveness of autologous platelet-rich plasma and peptide-enhanced bone graft in the treatment of intrabony defects. J Periodontol 2009; 80:62-71.
- Marx RE, Carlson ER, Eichstaedt RM, Schimmele SR, and Strauss JE, Georgeff KR. Platelet-rich plasma: Growth factor enhancement for bone grafts. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 1998; 85:638-646.
- Marx RE. Platelet-rich plasma: Evidence to support its use. J Oral Maxillofac Surg 2004; 62:489-496.
- Okuda K, Kawase T, Momose M, Murata M, Saito Y, Suzuki H et al. Platelet-rich plasma contains high levels of platelet derived growth factor and transforming growth

- factor- and modulates the proliferation of periodontally related cells in vitro J Periodontol 2003;74:849-857.
6. Arora NS, Ramanayke T, Ren YF, Romano GF. Platelet-rich plasma: a literature review. *Implant Dent* 2009; 18(4):303-308.
 7. Dori F, Nikolidakis D, Huszar T, Arweller NB, Gera I, Sculean A. Effect of platelet-rich plasma on the healing of intrabony defects treated with natural bone mineral and a collagen membrane. *J Clin Periodontol* 2007; 34:254-261.
 8. Piemontese M, Aspriello SD and Rubini C, Ferrante L, Procaccini M. Treatment of periodontal intrabony defects with de-mineralized freeze-dried bone allograft in combination with platelet-rich plasma: A comparative clinical trial. *J Periodontol* 2008; 79:802-810.
 9. Camargo PM, Lekovic V, and Weinlaender, Divnic-Resnik T, Pavlovic M, Kenny EB. A surgical re entry study on the influence of platelet-rich plasma in enhancing the regenerative effects of bovine porous mineral and guided tissue regeneration in the treatment of intrabony defects in humans. *J Periodontol* 2009; 80:915-923.
 10. Haranack L, Boedeker RH, Kurtulus I, Boehm S, Gonzales J, Meyle J et al. Use of platelet-rich plasma in periodontal surgery – A prospective randomized double blind clinical trial. *Clin Oral Invest* 2009; 13:179-183.
 11. Kaushick BT, Jayakumar ND, Padmalatha O, Varghese S. Treatment of human periodontal infrabony defects with hydroxyapatite + β tricalcium phosphate bone graft alone and in combination with platelet rich plasma: a randomized clinical trial. *Indian J Dent Res*. 2011; 22(4):505-10.
 12. Ozdemir B, Okte E. Treatment of intrabony defects with beta-tri calcium phosphate alone and in combination with platelet-rich plasma. *J Biomed Mater Res B Appl Biomater*. 2012; 100(4):976-83.
 13. Pradeep A.R, Rao N S, Agarwal E, Bajaj P, Kumari M, Naik SB. Comparative evaluation of autologous platelet-rich fibrin and platelet-rich plasma in the treatment of 3-wall intrabony defects in chronic periodontitis randomized controlled clinical trial. *J Periodontol* 2012; 83:1449-1457.
 14. Menzes LM, Rao J. Long term clinical evaluation of platelet-rich plasma in the treatment of human periodontal intraseous defects: A comparative clinical trial. *Quintessence Int* 2012; 43:571-582.
 15. Hassan KS, Alag AS, Abdel-Hady A. Torus mandibularis bone chips combined with platelet rich plasma gel for treatment of intrabony osseous defects: clinical and radiographic evaluation. *Int J Oral Maxillofac Surg*. 2012; 41(12):1519-26
 16. Kukreja BJ, Dodwad V, Kukreja P, Ahuja S, Mehra P. A comparative evaluation of platelet-rich plasma in combination with demineralized freeze-dried bone allograft and DFDBA alone in the treatment of periodontal intrabony defects: A clinic radiographic study. *J Indian soc periodontol* 2014; 18(5):618-623.
 17. Agarwal A, Gupta ND. Platelet-rich plasma combined with decalcified freeze-dried bone allograft for the treatment of non contained human intrabony periodontal defects: a randomized controlled split-mouth study. *Int J Periodontics Restorative Dent*. 2014; 34(5):705-11.
 18. Pinipe J, Mandalapu NB and Manchala SR, Mannem S, Gottumukkala S, Koneru S. Comparative evaluation of clinical efficacy of β -tri calcium phosphate (Septodont-RTB) alone and in combination with platelet rich plasma for treatment of intrabony defects in chronic periodontitis. *J Indian Soc Periodontol*. 2014; 18(3):346-51.
 19. Keceli HG, Sengun D, Berberoglu A, Karabulut E. Use of platelet gel with connective tissue grafts for root coverage: A randomized-controlled trial. *J Clin Periodontol* 2008; 35:255-262.
 20. Lafzi A, Chitsazi MT and Farahani RM, Faramarzi M. Comparative clinical study of coronally advanced flap with and without use of plasma rich in growth factors in the treatment of gingival recession. *Am J Dent*. 2011; 24(3):143-7.
 21. Jovovic B, Lazic Z and Nedic M, Matijevic S, Gostovic-Spadijer A. Therapeutic efficacy of connective tissue auto transplants with periosteum and platelet rich plasma in the management of gingival recession. *Vojnosanit Pregl*. 2013; 70(7):664-9.
 22. Aimetti M, Romano F, and Deliaiva C et al. Sinus grafting using autogenous bone and platelet-rich plasma: Histological outcomes in humans. *Int J Periodontics Restorative Dent* 2008; 28: 585-591.
 23. Schaff H, Streckbien P, and Lendeckel S, Heidinger K S, Rehmann P, Bodeker RH et al. Sinus lift augmentation using autogenous bone grafts and platelet-rich plasma. Radiographic results. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2008; 106:673-678.
 24. Torres J, Tamimi F, Martinez PP, Alkhrasat MH, Linares R, Hernandez G et al. Effect of platelet-rich plasma on Sinus lifting: A randomized clinical trial. *J Clin Periodontol* 2009; 36: 677-687.
 25. Khairy NM, Shendy EE, Askar NA, El-Rouby DH. Effect of platelet rich plasma on bone regeneration in maxillary sinus augmentation (randomized clinical trial). *Int J Oral Maxillofac Surg*. 2013; 42(2):249-55.
 26. Sunitha Raja V, Munirathnam Naidu E. Platelet rich fibrin: evolution of a second generation platelet concentrate. *Indian J Dent Res* 2008; 19:42-46.
 27. Dohan DM, Choukroun J, Diss A, Dohan SL, Dohan AJ, Mouhyi J et al. Platelet-rich fibrin (PRF): a second-generation platelet concentrate. Part I: technological concepts and evolution. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2006; 101: e37-e44.
 28. Dohan DM, Choukroun J, Diss A, Dohan SL, Dohan AJ, Mouhyi J et al. Platelet-rich fibrin (PRF): a second-generation platelet concentrate. Part III: leucocyte activation: a new feature for platelet concentrates? *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2006; 101: e51-e55.
 29. Choukroun J, Diss A, Simonpieri A, Girard MO, Schoeffler C, Dohan SL et al. Platelet-rich fibrin (PRF): a second-generation platelet concentrate. Part IV: clinical effects on tissue healing. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2006; 101: e56-e60.
 30. Thorat M, Pradeep AR, Pallavi B. Clinical effect of autologous platelet-rich fibrin in the treatment of intra-bony defects: a controlled clinical trial. *J Clin Periodontol* 2011; 38(10):925-32.
 31. Rosamma Joseph V, Raghunath A, Sharma N. Clinical effectiveness of autologous platelet rich fibrin in the management of infrabony periodontal defects. *Singapore Dent J* 2012; 33(1):5-12.
 32. Bansal C, Bharti V. Evaluation of efficacy of autologous platelet-rich fibrin with demineralized-freeze dried bone allograft in the treatment of periodontal intrabony defects. *J Indian Soc Periodontol*. 2013; 17(3):361-6.
 33. Mathur A, Bains VK, and Gupta V, Jhingran R, Singh G.P. Evaluation of intrabony defects treated with platelet-rich fibrin or autogenous bone graft: A comparative analysis. *Eur J Dent*. 2015; 9(1):100-8.
 34. Shah M, Patel J, and Dave D, Shah S. Comparative evaluation of platelet-rich fibrin with demineralized freeze-dried bone allograft in periodontal infrabony defects: A randomized controlled clinical study. *J Indian Soc Periodontol*. 2015; 19(1):56-60.
 35. Ajwani H, Shetty S, Gopalakrishnan D. Comparative evaluation of platelet-rich fibrin biomaterial and open flap debridement in the treatment of two and three wall intrabony defects. *Int Oral Health*. 2015; 7(4):32-7.
 36. Agarwal A, Gupta ND, Jain A. Platelet rich fibrin combined with decalcified freeze-dried bone allograft for the treatment of human intrabony periodontal defects: a randomized split mouth clinical trial. *Acta Odontol Scand*. 2015; 14:1-8.
 37. Pradeep A.R., Nagpal K, Karvekar S, Patanaik K, Naik SB, Guru Prasad CN. Platelet rich fibrin with 1% Metformin for the treatment of intrabony defects in chronic periodontitis: A randomized clinical trial. *J Periodontol* 2015; 86(6):729-737.
 38. Gupta SJ, Jhingran R, Gupta V, Bains VK, Madan R, Rizvi I. Efficacy of platelet-rich fibrin vs. enamel matrix derivative in the treatment of periodontal intrabony defects: a clinical and cone beam computed tomography study. *J Int Acad Periodontol*. 2014; 16(3):86-96.
 39. Del Corso M, Sammartino G, Dohan Ehrenfest DM. Clinical evaluation of a modified coronally advanced flap alone or in combination with a platelet-rich fibrin membrane for the treatment of adjacent multiple gingival recessions: a 6-month study. *J Periodontol*. 2009; 80(11):1694-7
 40. Sofia Aroca, Tibor Keglevich, Bruno Barbieri et al. Clinical evaluation of modified coronally advanced flap alone or in combination with a platelet-rich fibrin membrane for the treatment of adjacent multiple gingival recessions: A 6-month study. *J Periodontol* 2009; 80(2):244-252.
 41. Jankovic S, Aleksic Z, Klokkevold P, Lekovic V, Dimitrijevic B, Kenny EB, Camargo P. Use of platelet-rich fibrin membrane following treatment of gingival recession: A randomized clinical trial. *Int J Periodontics Restorative Dent* 2012; 32:e41-e50.
 42. Padma R, Shilpa A, Kumar PA, Naga sri M, Kumar C, Sreedhar A. A split mouth randomized controlled study to evaluate the adjunctive effect of platelet-rich fibrin to coronally advanced flap in Miller's class-I and II recession defects. *J Indian Soc Periodontol*. 2013; 17(5):631-637
 43. Eren G, Atilla G. Platelet-rich fibrin in the treatment of localized gingival recessions: a split-mouth randomized clinical trial. *Clin Oral Invest*. 2014; 18(8):1941-8.
 44. Tunali M, Ozdemir H, Arabaci T, Gurbuzer B, Pikkoken L, Firatli E. Clinical evaluation of autologous platelet-rich fibrin in the treatment of multiple adjacent gingival recession defects: a 12-month study. *Int J Periodontics Restorative Dent*. 2015; 35(1):105-14
 45. Thamaraiselvin, Elavarasu S, Thangakumaran S, Gadgi JS, Arthie T. Comparative clinical evaluation of coronally advanced flap with or without platelet rich fibrin membrane in the treatment of isolated gingival recession. *J Indian Soc Periodontol*. 2015; 19(1):66-71.
 46. Toffler M, Toscano N, Holtzclaw D. Osteotome-mediated sinus floor elevation using only platelet-rich fibrin: an early report on 110 patients. *Implant Dent*. 2010; 19(5):447-56.
 47. Zhang Y, Tangl S, Huber CD, Lin Y, Qiu L, Rausch-Fan X. Effect of Choukroun's platelet rich fibrin on bone regeneration in combination with deproteinized bovine bone material in maxillary sinus augmentation: A histological and histomorphometric study. *J Craniomaxillofac Surg* 2012; 40(4):321-8.
 48. Sharma A, Pradeep A.R. Autologous Platelet-rich fibrin in the treatment of mandibular degree II furcation defects: A randomized clinical trial. *J Periodontol* 2011; 82:1396-1403.
 49. Bajaj P, Pradeep AR and Agarwal E, Rao NS, Naik SB, Priyanka N et al. Comparative evaluation of autologous platelet-rich fibrin and platelet-rich plasma in the treatment of mandibular degree II furcation defects: a randomized controlled clinical trial. *J Periodontol Res*. 2013; 48(5):573-81.
 50. Hauser F, Gaydarov Nand Badoud I et al. Clinical and histological evaluation of post extraction platelet-rich fibrin socket filling: a prospective randomized controlled study. *Implant Dent*. 2013; 22(3):295-303.
 51. Barone A, Ricci M and Romanos GE et al. Buccal bone deficiency in fresh extraction sockets: a prospective single cohort study. *Clin Oral Implants Res*. 2014; 31:45-49
 52. Hamzacebi B, Oduncuoglu B, Alaaddinoglu EE. Treatment of Peri-implant Bone Defects with Platelet-Rich Fibrin. *Int J Periodontics Restorative Dent*. 2015; 35(3):415-22.
 53. M. Tunali, H. Ozdemir, Z. Kuçukodaci, S. Akman, E. Yaprak, E. Firatli et al. "In vivo evaluation of titanium-prepared platelet-rich fibrin (T-PRF): a new platelet concentrate". *Br J Oral Maxillofac Surg* 2012; 51(5):438-443.
 54. S. Takemoto, T. Yamamoto, K. Tsuru, S. Hayakawa, A. Osaka, S. Takashima et al. "Platelet adhesion on titanium oxide gels: effect of surface oxidation". *Biomaterials* 2004; 25(17):3485-3492.
 55. M. Tunali, H. Ozdemir, Z. Kuçukodaci, S. Akman, E. Yaprak, E. Firatli et al. A Novel Platelet Concentrate: Titanium-Prepared Platelet-Rich Fibrin. *Biomed research international* 2014; 1; 1-7.
 56. Ghanaati S, Booms P, Orłowska A, Kubesch A, Lorenz J, Rutkowski J et al. Advanced Platelet-rich fibrin: A new concept for cell based tissue engineering by means of inflammatory cells. *J Oral Implant* 2014; 40; 680-689
 57. Reduction of relative centrifugation force within injectable platelet-rich-fibrin (PRF) concentrates advances patients' own inflammatory cells, platelets and growth factors: the first introduction to the low speed centrifugation concept. *Eur J Trauma Emerg Surg*. 2018 Feb; 44(1):87-95.
 58. Sohn DK, Huang B, Kim J, Park W.E, Park CC. Utilization of Autologous concentrated growth factors (GGF) enriched bone graft matrix (Sticky bone) and CGF-Enriched membrane in implant dentistry. *The J Implant Adv Clin Dent* 2015; 7:11-27.