

Original Research Paper

General Medicine

AN OBSERVATIONAL STUDY OF THE CLINICO-MICROBIOLOGICAL SENSITIVITY PATTERN IN A TERTIARY CARE TEACHING HOSPITAL OF THE RAMGANGA REGION OF U.P.(INDIA) DURING A PERIOD OF 16.9.2019-15.3.2020

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ABSTRACT

Background and rationale: Worldwide antibiotic resistance is a problem. Many complex factors are related to this. India is also seeing the rise in resistant strains of several bacteria on the antibiotic sensitivity pattern and prevalent microorganisms. This study also attempts to establish a way of presentation of the relevant findings which can be used in future to ensure easy comparability and contrasting of findings. In any tertiary care teaching hospital antimicrobials are a major class of drugs prescribed. Empirical use of antibiotics are needed many a times but it cause drug resistance which need study and need evaluation after the culture and sensitivity reports are available. This study was planned to assess the drug resistance and sensitivity patterns of the isolates from urine, bold, body fluids as well as pus of which 856 samples were studied within sixmonths' time and the sensitivity to antibiotics were studied.

Methods: An observational-prospective study was conducted in the Tertiary care teaching hospital over a period of six months to assess antibiotic sensitivity and resistance pattern. The specimens were collected from the adult patients (age >18 years) admitted in the Internal Medicine ward of Rohilkhand Medical College Hospital, Bareilly, over a period of 6 months. The sampling technique was consecutive sampling method. Specimens which were culture positive, were only included in the study for analysis. Multiple specimens were taken. A total of 856 consecutive patients were included in the study receiving antibiotics in the hospital as indoor patients having cultures with significant growth. Samples of urine, blood and body fluids were collected and culture growth was identified with antimicrobial sensitivity testing was done and resistance pattern was noted.

KEYWORDS: Antibiotics, sensitivity pattern, Resistance

METHODS:

An observational- prospective study was conducted in the Tertiary care teaching hospital over a period of six months to assess antibiotic sensitivity and resistancepattern. The specimens were collected from the adult patients (age >18 years) admitted in the Internal Medicine ward of Rohilkhand Medical College Hospital, Bareilly, over a period of 6 months. The sampling technique was consecutive sampling method. Specimens which were culture positive, were only included in the study for analysis. Multiple specimens were taken.

A total of 856 consecutive patients were included in the study receiving antibiotics in the hospital as indoor patients having cultures with significant growth. Samples of urine, blood and body fluids were collected and culture growth was identified with antimicrobial sensitivity testing was done and resistance pattern was noted.

RESULTS:

Overall Only 28.73%(246 out of 856 sample) had Staphyllococus aureus, a Gram positive organism and hence, a Gram negative preponderance was seen where E. coli was present in 37.5% of samples and Pseudomonas 15.5%, Klebsila 11.9% and Acinobacter 6.3%.of samples.

- S. aureus was sensitive to vancomycin (100 %), linesezolid(100 %), teicarrcillin (98%), amikacine (89%), levofloxacine (88%),
- E. colihad sensitivity to imipramin (85 %), amikacin (80%), chloramphenicol (60%), pipracillin + tazobacter (58%), gentamicin (60%)
- Pseudomonashad sensitivity toimipramin (83%), Pip+Tazo(76%),Amika(71%),Pipera(71%),Cipro(68), Oflox(61%)
- Klebsilahad sensitivity toimipramin (85%), Chlora(55%), pipracillin + tazobacter (65%), Amika (67%), Cefo+Sul (63%), levofloxacine (58%)
- Acinobacter had sensitivity topipracillin + tazobacter (42%), imipramin(50%), cefop+sul(55%), ofloxacine (50%), levofloxacine (66%), amikacine (48%)
- In uinary samplesE.coli had sensitivity to norfloxacine 7% and nitofurantoin 36%
- Staph aureus had sensitivity to norfloxacine nitofurantoin 3% and 16%, Pseudomonas had sensitivity to norfloxacine 3% and nitofurantoin 0.7%, Klebsila had sensitivity to norfloxacine 3% and nitofurantoin 2%
- Acinobacterhad sensitivity to norfloxacine 4% and nitofurantoin 0%

	Pathogens	(%)	Trend of sensitivity pattern (% sensitivity)	Norflox	Nitrofur
1	E .coli Tobra (54%)	Present (37.5 %)	Imip(85 %),Amika(80%),Chlora (60%),Pip+Tazo(58%), Genta (60%)		36%
		Past (37 %)	Imip(91%)Amika(79%), Chlora(68%),Genta(55 %)Tobra(61%),		88%
2	Staphylococcus aureus Clinda (55 %) Tetra (74%),Doxy (61)	Present (28.7 %)	Vanco(100 %),Linez(100 %),Teico(98%),Amika(8 9%),Levo(88 %),	3%	16%

		Past (27 %)	Vanco(100 %),Linez(100 %),Teico(91 %),Tetra (88%),Doxy(87 %),	17%	65%
3	Pseudomonas Tobra (52 %) Genta (63 %)	Present (15.5 %)	Imipen(83%),Pip+Tazo (76%),Amika(71%),Pip era(71%), Cipro(68), Oflox(61%)	3%	0 .7 %
		Past (19%)	Imipen(85%),Amika(75 %),Pip+Tazo(71%), Genta(68%),Tobra (67%)	0%	0%
4	Klebsiella Tobra (44 %)	Present (11.9%)	Imipen(85%),Chlora(5 5%),Pip.+Tazo(65%),A mika(67%),Cefo+Sul(6 3%),Levo(58%)		2%
		Past (10 %)	Imipen(86%),Pip.+Taz o(71%),Amika(67%),Ce fo+Sul (60%),Tobra (59%)		31%
5	Acinetobacter Mero(57%) Tobra(32 %)	Present (6.3 %)	Pip+Tazo(42%),Imipen (50%),Cefop+sul(55%) ,Oflox(50%),Levo(66%),Amika(48%)		0%
		Past (7 %)	Imip(77%),Merop(66%),Tobra (58%),Pip+Tazo(54%),Cefop+sul(48%)	0%	0%
Total Sample856			-		
Clinical Sample	321 E.coli 31	7 246 Sathaph 210	133 pseudom 155	102 Klebsiella 182	54 Acinetobacter 56
	Pre. Past.	Pre. Past.	Pre. Past.	Pre. Past.	Pre. Past.
Body fl+Resp+ Pus	166 158	173 147	103 115	76 60	43 48
Urine	145 156	48 44	19 23	23 19	4
Blood	10 3	25 19	11 17	3 3	7
MRSA / ESBL PRODUCER	ESBL 135	MRSA 116		ESBL 35	
_	42%	47%		34%	

DISCUSSION-

- From the sensitivity patternit is obvious that the vancomycin and lineszolid had the best sensitivity for the Gram Positive Staphylococcus but should be kept reserved as drugs to be used after failingof any of the levofloxacine, amikacine and teicarrcillin
- For Gram negatives for E.coli, imiprmine is the most sensitive but should be used after the failure amikacin chloramphenicol pipracillin + tazobacter, gentamicin
- Pseudomonas had sensitivity to imipramin (83%), Pip+Tazo(76%),Amika(71%),Pipera(71%),Cipro(68), Oflox(61%)
- Klebsila had sensitivity to imipramin (85%), Chlora(55%), pipracillin + tazobacter (65%), Amika(67%), Cefo+Sul(63%), levofloxacine (58%)
- Acinobacter had sensitivity to pipracillin + tazobacter (42%), imipramin(50%), cefop+sul (55%), ofloxacine (50%), levofloxacine (66%), amikacine (48%)

For urinary samples sensitivity is relatively low (with *E.coli* had sensitivity to norfloxacine 7% and nitofurantoin 36%.

 Staph aureus had sensitivity to norfloxacine nitofurantoin 3% and 16%, Pseudomonas had sensitivity to norfloxacine 3% and nitofurantoin 0.7%, Klebsila had sensitivity to norfloxacine 3% and nitofurantoin 2%

- Acinobacter had sensitivity to norfloxacine 4% and nitofurantoin 0%
- It will suggest clinician to choose according to the severity
 of the signs and symptoms to continue or change the
 antibiotics or uinary disinfectants in different clinical
 settings.

CONCLUSIONS:

Probably our sensitivity pattern is different than of the western world and we may use selected antibiotics for full course to avoid resistance.

A preponderance of gram negative bacteria over gram positive bacteria was noted with a higher degree of resistance to most of the first line antimicrobial agents. For urine samples.

Despite the widespread availability of antibiotics, UTI remains the most common bacterial infection in the human population. I Antibiotics are usually given empirically before the laboratory results of urine culture are available. To ensure appropriate therapy, current knowledge of the organisms that cause UTI and their antibiotic susceptibility is mandatory. 2

The development of antimicrobial resistance is a natural

process, which cannot be stopped. Resistance means that people cannot be effectively treated and they remain ill for longer period of time. It also means that epidemics are prolonged and thus that there is a greater risk of infection to others. The development of resistance is accelerated when antimicrobials are misused (http://www.emro.who.net). Despite the use of potent antibiotics still high mortality exists in case of P. aeruginosa infections. Nosocomial multidrug resistant P. aeruginosa is an important health care problem worldwide³ Antimicrobial resistance prolongs the duration of hospitalization, thereby, increasing the cost of patient care. There are multiple factors, which contribute to the global spread of resistance. Decreasing unnecessary antibiotic use, treating with narrow spectrum agents, improving compliance with therapy, decreasing use of antibiotic in animal and agriculture, and improving infection control all have a role in confronting this problem. In addition, immunization may diminish the impact of resistance by preventing infection and also the carriage of transmission.

The indiscriminate use of broad-spectrum antibiotics is much emphasized in various studies 4,6

Carbapenems and teicoplanin and as found least used and piperacillin-tazobactum was the most commonly used parentral antimic robial followed by amoxicillin-clavulanic acid which may be suggested as per our sensitivity pattern also 7

The role of fluoroquinolones is well established in CAP and all western guidelines where the baseline prevalence of TB is low endorse them. However, in India where there is a high burden of TB and where TB may present as CAP, use of empiric fluoroquinolone therapy may lead to masking and delayed diagnosis of TB and promotion of drug resistance. Therefore, fluoroquinolines are best avoided. Similarly, drugs with antitubercular activity including linezolid and aminoglycosides should not be used as per ICMR guidelines. §

REFERENCES:

- Sharma S. rrent understanding of Pathogenic mechanisms in UTIs. Ann Natl Acad Med Sci 1997; 33(1):31-8.
- Gruneberg GN. Antibiotic sensitivities of urinary pathogens: 1971-1982. J Antimicrob Chemother 1984; 14:17-23
- Susceptibility pattern of pseudomonas aeruginosa against various antibiotics Farida Anjum 1 and Asif Mir 2 * 1 Pakistan Council for Science and Technology (PCST), Islamabad. 2Department of Biosciences, COMSATS Institute of Information Technology, Islamabad.African Journal of Microbiology Research Vol. 4 (10), pp. 1005-1012, 18 May 2010 Available online http://www.academicjournals.org/ajmr
- Ochoa C, Eiros JM, Inglada L, Vallano A, Guerra L. Assessment of antibiotic prescription in acute respiratory infections in adults. J Infect. 2000;41(1):73–83. [PubMed] [Google Scholar]
- Piccirillo JF, Mager DE, Frisse ME, Brophy RH, Goggin A. Impact of first-line vs second-line antibiotics for the treatment of acute uncomplicated sinusitis. Jama-Journal Am Med Assoc [Internet] 2001;286(15):1849–56. [PubMed] [Google Scholar]
- Salvo F, Polimeni G, Moretti U, Conforti A, Leone R, Leoni O, et al. Adverse drug reactions related to amoxicillin alone and in association with clavulania acid: Data from spontaneous reporting in Italy. J Antimicrob Chemother. 2007;60(1):1
- Tiwari SA, Ghongane BB, Daswani BR, Dabhade SS. Restricted Parenteral Antibiotics Usage Policy in a Tertiary Care Teaching Hospital in India. J Clin Diagn Res. 2017;11(5):FC06–FC09. doi:10.7860/JCDR/2017/24048.977
- 8. Treatment Guidelines for Antimicrobial Use in Common Syndromes 2019, ICMR, New Delhi, India