



EXTENDED SPECTRUM BETA LACTAMASE PRODUCING UROPATHOGENIC ESCHERICHIA COLI AND THEIR ANTIMICROBIAL SUSCEPTIBILITY PATTERN IN A TERTIARY CARE HOSPITAL

Dr Shakshi Bansal

MBBS, PGT (MD), Department of Microbiology, Mahatma Gandhi Memorial Medical College and Maharaja Yashwantrao Hospital, Indore, MP, India

Dr Ila Srivastava Bajpai*

MBBS, MD (Assistant Professor) Department of Microbiology, Mahatma Gandhi Memorial Medical College and Maharaja Yashwantrao Hospital, Indore, MP *Corresponding Author

Dr Anita Mutha

MBBS, MD (Professor & HOD) Department of Microbiology, Mahatma Gandhi Memorial Medical College and Maharaja Yashwantrao Hospital, Indore, MP, India.

Dr Prachi Priya

MBBS, PGT (MD) Department of Microbiology, Mahatma Gandhi Memorial Medical College and Maharaja Yashwantrao Hospital, Indore, MP, India.

ABSTRACT

Urinary tract infection (UTI) due to extended spectrum beta-lactamase (ESBL)-producing bacteria including *Escherichia coli* has become widespread and also there is a changing trend in the antibiotic susceptibility pattern to the conventional drugs used in the treatment of urinary tract infections due to the production of extended-spectrum beta-lactamases (ESBLs). This study was done to find the percentage of ESBL producing *Escherichia coli* and to study the antibiotic resistance profile of the ESBL and non-ESBL producing *Escherichia coli*. Urine samples were processed and identification of bacterial growth was confirmed by standard microbiological procedures. Antimicrobial susceptibility and ESBL detection were performed using Clinical and Laboratory Standards Institute (CLSI M100 2022) guidelines, ESBL detection was done by combined disc method using cefotaxime (30 ug) versus cefotaxime plus clavulanate (30+10 ug). Out of total 1238 urine samples 549 were found to be culture positive. *Escherichia coli* (36.2%) was the most frequently isolated uropathogen followed by *Klebsiella pneumoniae* (23.6%). Among the isolated *Escherichia coli* 44.2% were ESBL producers which were found highly resistant in comparison to non-ESBL producers.

KEYWORDS : Uropathogen, *Escherichia coli*, Antimicrobial susceptibility, Extended spectrum beta lactamases (ESBL)

INTRODUCTION:

Urinary Tract Infections (UTI) is one of the most common community-acquired as well as nosocomial infections ⁽¹⁾, *Escherichia coli* being the common causative organism of urinary tract infections (UTI) ⁽²⁾. Other organisms responsible are *Proteus*, *Pseudomonas*, *Salmonella*, *Staphylococcus saprophyticus*, *Enterococcus*, *Staphylococcus aureus* ⁽³⁾. The introduction of antimicrobial therapy has contributed significantly to the management of UTIs. However the main problem with current antibiotic therapies is the rapid emergence of antimicrobial resistance in hospitals and the community ⁽⁴⁾.

Extended spectrum beta-lactamases (ESBL's) are enzymes that confer resistance to most common beta-lactam antibiotics such as Penicillins, Cephalosporins and Monobactams but not for Cephamecins and Carbapenems. Detection of ESBL producers from sample is important because it represents an epidemiologic marker of colonization and therefore there is potential for transfer of such organisms to other patients. The rapid increase of resistance to broad spectrum beta lactams among uropathogens has recently become a major problem globally. It leads to antibiotic ineffectiveness, increased severity of illness and cost of treatment. The serious increase in the prevalence of ESBL's worldwide creates a need for effective and easy to perform screening methods for detection ⁽⁵⁾.

MATERIAL AND METHODS:

The present prospective study was carried out from October 2022 to December 2022 in the Department of Microbiology of a tertiary care hospital. A total of 1238 urine samples were collected in sterile containers. The samples were processed on chromogenic (urochrome) media and the culture plates were incubated at 37°C for 18-24 hrs under aerobic conditions. Identification of bacterial growth was confirmed by standard microbiological and biochemical techniques.

Confirmed *Escherichia coli* identified phenotypically were further tested for antimicrobial susceptibility which was performed using Clinical and Laboratory Standards Institute (CLSI M100 2022) guidelines

Extended spectrum beta-lactamase confirmatory tests as per CLSI M100 2022, was done by using cefotaxime and cefotaxime plus clavulanic acid (30/10 mcg) discs on Muller-Hinton agar using double disc diffusion method. Organism was considered as ESBL producer if there was an increase of ≥ 5 mm increase in the zone diameter to increase of ≥ 5 mm in the zone diameter of cefotaxime/ clavulanic acid disc with respect to that of cefotaxime disc alone.

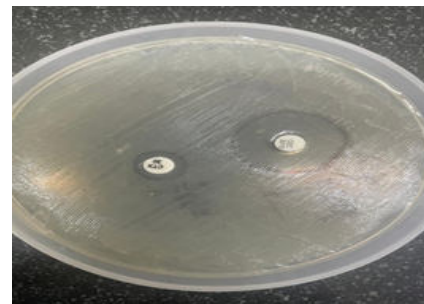


Figure 1: Phenotypic expression of extended spectrum beta-lactamase production (zone sizes of cefotaxime (CTX)/clavulanic acid(CEC) ≥ 5 mm. than the zone sizes of cefotaxime)

RESULT AND DISCUSSION:

A total of 1238 urine samples were collected from patients visiting the outpatient departments (OPD) and inpatient departments (IPD) of a tertiary care hospital out of them 638 were sterile and only 549 sample revealed uropathogens. Among them 376 (69.0%) isolates were Gram-negative, 128

(23%) were Gram-positive while 45 (9%) were identified as *Candida* species. Out of the total 549 uropathogenic isolates *Escherichia coli* (199,36.2%) was the predominant uropathogen, followed by *Klebsiella spp.* (130,23.6%), *Enterococcus spp.* (120,21.8%), *P. aeruginosa* (20,3.6%) and other pathogens like *Proteus spp.*, *Acinetobacter baumannii*, *Enterobacter spp.* (26,4.84%) were also isolated (Table 1).

Among 199 strains of *Escherichia coli* isolate, 88 were shown to be ESBL producers.

In the current study, out of total 199 *Escherichia coli* 88 (44.2%) were ESBL producers (Table 2) which is similar to the studies done by Bajpai *et al.*, Kumar *et al.*, Taneja *et al.* and Tankhiwala *et al.* (49.8%), (41.1%,39.0% and 40.2%)^(5,6,7,8) but much less than the studies done by Ramesh *et al.* (60.7%), Singhal *et al.* (62%)^(9,10).

Table 1: Distribution Of Organisms Isolated

ORGANISM ISOLATED	NUMBER	PERCENTAGE(%)	ORGANISM	PERCENTAGE
GRAM NGATIVE BACILLI	357	65.0(%)	<i>Escherichia coli</i>	36.2%
			<i>Klebsiella spp</i>	23.6%
			<i>Pseudomonas aeruginosa</i>	3.6%
			<i>Acinetobacter baumannii</i>	3.4%
			<i>Proteus spp</i>	0.9%
			<i>Enterobacter spp</i>	0.54%
			<i>Enterococcus spp</i>	21.8%
GRAM POSITIVE COCCI	128	23.0	<i>Staphylococcus aureus</i>	1.45%
BUDDING YEAST CELL	45	9.8%	<i>Candida spp</i>	9.8%

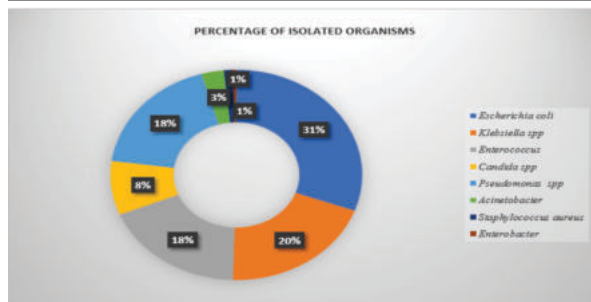


Figure 2: Distribution Of Organisms Isolated

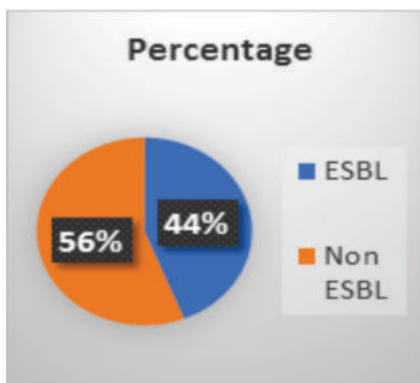


Figure 3: Distribution Of Escherichia Coli As Esbl And Non Esbl Producers

The antibiogram (Figure 3) of the *Escherichia coli* isolates shows that the ESBL producers have high degree of resistance not only towards penicillin and cephalosporin group of antibiotics but also towards other group of antimicrobials

which are routinely prescribed for UTI's as compared to non-ESBL producers, These findings are similar to those reported by other authors like Bajpai *et al.*, Sasirekha, Ndugulile *et al.* and Mehrgan *et al.*^(5,11,12,13), this can be explained by the fact that ESBL are plasmid-coded enzymes, which are transferable between one bacterium to another and such transferable plasmids also code for resistance determinants to antimicrobial agents other than beta-lactams⁽¹⁴⁾.

Table 2: Comparison Of Resistance Pattern Of Esbl And Non Esbl Producers

DRUGS	RESISTANCE PATTERN (%)		
	ESCHERICHIA COLI	ESBL PRODUCERS	NON ESBL PRODUCERS
Gentamicin	35.3%	29.4%	41.6%
Ciprofloxacin	83.1%	81.3%	68.7%
Norfloxacin	74.2%	84.8%	75.3%
Cotrimoxazole	53%	59.1%	43.0%
Imipenam	48.5%	39%	33.3%
Nitrofurantoin	6%	8%	3%
Cefipime	47.4%	58.9%	49.0%
Meropenem	38.4%	42.3%	24%
Ampicillin	90%	100%	85%

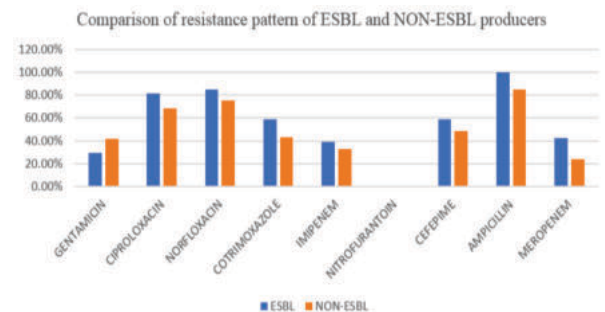


Figure 4: Comparison Of Resistance Pattern Of Esbl And Non Esbl Producers

Highest resistance in our study is seen with ampicillin 100% and 85% among ESBL and non ESBL producers it may be due their widespread use^(5,15), high level of resistance is also seen with Norfloxacin (84%,75%) and Ciprofloxacin (81.3%,68.7) which may be due to an increased Quinolone resistance in our institute. This finding is consistent with previous studies^(5,15,16). These drugs are ubiquitously prescribed which may be the reason for the emergence of resistance against them⁽⁵⁾.

In case of Gentamycin (29.4% and 41.6%) resistance was observed for both ESBL and non-ESBL producers respectively. In our study resistance to Imipenam and Meropenem is in range of 25%-45% but it is 100% sensitive in most of the previous studies^(1,17,18,19) and 6.3% and 11% resistance is observed in the study of Bajpai *et al.* and Rajeshwari *et al.* respectively^(5,3). The resistance exhibited in our case is due to existence of carbapenemase producing isolates in our setting. The increasing resistance for Carbapenems is alarming and gives rise to an increasing concern over the judicious use of carbapenems in our health facilities^(20,21).

In our study lowest degree of resistance is with Nitrofurantoin (8% and 3.7%) it can be attributed to its localized action on urinary tract and not being exposed outside urinary tract⁽¹⁵⁾, and thus Nitrofurantoin is recommended as an appropriate agent for first line treatment of community-acquired UTI⁽¹⁴⁾.

CONCLUSION:

Resistance to commonly used antibiotics is increasing day by day and it also vary over time and geographical distribution. Therefore, continuous monitoring of the antibiotic resistance pattern guides the clinician to initiate the empirical treatment

of UTI. Antimicrobial stewardship knowledge helps in avoiding the treatment failure and rapid dissemination of the antibiotic resistance and its mechanism can be prevented.

REFERENCES:

- 1) Aruna K, Mobashshera T. Prevalence of extended spectrum beta-lactamase production among uropathogens in south Mumbai and its antibiogram pattern. EXCLIJ. 2012 Jul 6;11:363-72. PMID: 27418912; PMCID: PMC4942789.
- 2) Arif Hussain, Christa Ewers, Nishant Nandanwar, Sebastian Guenther, Savita Jadhav, Lothar H. Wieler, Niyaz Ahmed; Multiresistant Uropathogenic *Escherichia coli* from a Region in India Where Urinary Tract Infections Are Endemic: Genotypic and Phenotypic Characteristics of Sequence Type 131 Isolates of the CTX-M-15 Extended-Spectrum- β -Lactamase-Producing Lineage, ASM journal:volume56 no.12
- 3) Rajeswari Pilli, Vamsi Chakradhar Kapaganty, Study of extended spectrum beta lactamase producing uropathogens and their antibiotic susceptibility pattern Indian Journal of Microbiology Research: 2018;5(2):280-283
- 4) Sood S, Gupta R. Antibiotic resistance pattern of community acquired uropathogens at a tertiary care hospital in jaipur, rajasthan. Indian J Community Med. 2012 Jan;37(1):39-44. doi: 10.4103/0970-0218.94023. PMID: 22529539; PMCID: PMC3326806.
- 5) Bajpai T, Pandey M, Varma M, Bhatambare GS. Prevalence of extended spectrum beta-lactamase producing uropathogens and their antibiotic resistance profile in patients visiting a tertiary care hospital in central India: Implications on empiric therapy. Indian J Pathol Microbiol 2014;57:407-126) Kumar MS, Lakshmi V, Rajagopalan R. Occurrence of extended spectrum beta-lactamases among *Enterobacteriaceae* spp. isolated at a tertiary care institute. Indian J Med Microbiol 2006;24:208-11.
- 7) Taneja N, Rao P, Arora J, Dogra A. Occurrence of ESBL & Amp-C beta-lactamases & susceptibility to newer antimicrobial agents in complicated UTI. Indian J Med Res 2008;127:85-8.
- 8) Tankhiwale SS, Jalgaonkar SV, AhamadS, HassaniU. Evaluation of extended spectrum beta lactamase in urinary isolate. Indian J Med Res 2004;120:553-6
- 9) Ramesh N, Sumathi CS, Balasubramanian V, Palaniappan KR, Kannan VR. Urinary tract infections and antimicrobial susceptibility pattern of extended spectrum of beta lactamase producing clinical isolates. Adv Biol Res 2008;2:78-82.
- 10) Singhal S, Mathur T, Khan S, Upadhyay DJ, Chugh S, Gaiind R, et al. Evaluation of methods for AmpC beta-lactamase in gram negative clinical isolates from tertiary care hospitals. Indian J Med Microbiol 2005;23:120-4
- 11) Sasirekha B. Prevalence of ESBL, AMPC B-lactamases and MRSA among uropathogens and its antibiogram. EXCLIJ 2013;12:81-
- 12) Ndugulile F, Jureen R, Harthug S, Urassa W, Langeland N. Extended spectrum beta-lactamases among Gram-negative bacteria of nosocomial origin from an intensive care unit of a tertiary health facility in Tanzania. BMC Infect Dis 2005;5:86.
- 13) Mehrgan H, Rahbar M, Arab-Halvahi Z. High prevalence of extended-spectrum beta-lactamase-producing *Klebsiella pneumoniae* in a tertiary care hospital in Tehran, Iran. J Infect Dev Ctries 2010;4:132-8.
- 14) Paterson DL. Resistance in gram-negative bacteria: Enterobacteriaceae. Am J Infect Control 2006;34:20-8.
- 15) Manjunath GN, Prakash R, Vamseedhar A, Shetty K. Changing trends in the spectrum of antimicrobial drug resistance pattern of uropathogens isolated from hospitals and community patients with urinary tract infections in Tumkur and Bangalore. Int J Biol Med Res 2011;2:504-7
- 16) Akram M, Shahid M, Khan AU. Etiology and antibiotic resistance patterns of community-acquired urinary tract infections in J N M C Hospital Aligarh, India. Ann Clin Microbiol Antimicrob 2007;6:4
- 17) Babypadmini S, Appalaraju B. Extended spectrum -lactamases in urinary isolates of *Escherichia coli* and *Klebsiella pneumoniae* - prevalence and susceptibility pattern in a tertiary care hospital. Indian J Med Microbiol. 2004 Jul Sep;22(3):172-4. PMID: 17642726
- 18) Ponnusamy P, Natarajan V, Sevanan M. In vitro biofilm formation by uropathogenic *Escherichia coli* and their antimicrobial susceptibility pattern. Asian Pac J Trop Med. 2012 Mar;5(3):210-3. doi: 10.1016/S1995-7645(12)60026-1. PMID: 22305786.
- 19) Nachimuthu Ramesh. Advances in Biological Research 2 (5-6): 78-82, 2008 ISSN 1992-0067 © IDOSI Publications, 2008 Urinary Tract Infection and Antimicrobial Susceptibility Pattern of Extended Spectrum of Beta Lactamase Producing Clinical Isolates 1
- 20) Kumarasamy KK, Toleman MA, Walsh TR, Bagaria J, Butt F, Balakrishnan R, et al. Emergence of a new antibiotic resistance mechanism in India, Pakistan, and the UK: A molecular, biological, and epidemiological study. Lancet Infect Dis 2010;10:597-602.
- 21) Yong D, Toleman MA, Giske CG, Cho HS, Sundman K, Lee K, et al. Characterization of a new metallo- β -lactamase gene, bla, and a novel erythromycin esterase gene carried on a unique genetic structure in *Klebsiella pneumoniae* sequence type 14 from India. Antimicrobial Agents Chemother 2009;53:5046-54