RESEARCH PAPER

Pharma



Phytochemical and *In-Vitro* Antioxidant Activities of *Melia azedarach* Linn, Catharanthus *Rosea* and Brassica oleracea L.var.capitata

KEYWORDS	Melia azedarach Linn, Catharanthus Rosea and Brassica oleracea L.var.capitata, DPPH, Ethanolic extracts and Antioxidant activity.			
Mohammed Fazil Ahmed		A. Srinivasa Rao		
Nizam Institute of Pharmacy & Research Center, Deshmukhi, Pochampally (M), Near Ramoji Film City, Nalgonda,(AP), INDIA-508284		Bhaskar Pharmacy College,Yeknapally, Moinabad(Mandal), R.R(Dist), Hyderabad-500075		
ABSTRACT The present study was to analysis the antioxidant activities of ethanolic leaves extracts of Melia azedarach				

Linn, Catharanthus Rosea and Brassica oleracea L.var.capitata by DPPH scavenging assay. The phytochemical screening was also carried on the leaves extracts of Melia azedarach, Catharanthus Rosea and Brassica oleracea L.var. capitata revealed the presence of some active ingredients such as Alkaloids, Tannins, Saponins, Phenols, glycosides and flavonoids. The results revealed that the ethanolic extracts of the leaves of Brassica oleracea L.var.capitata had exhibited more antioxidant activity than Catharanthus Rosea and Melia azedarach Linn. While the ethanolic leaves extract of Catharanthus Rosea had exhibited moderate antioxidant activity than other extracts. The high scavenging property of extracts may be due to hydroxyl groups existing in the phenolic compounds.

INTRODUCTION

Free radicals are the compounds generated from normal body processes and also from environmental pollutions. Within the human body, millions of chemical reactions are occurring constantly .These processes require oxygen. Reactive oxygen spices (ROS), sometimes called active oxygen species, are various from of activated oxygen, which include free radicals such as superoxide ions (O2 -) and hydroxyl radicals (OH-), as well as non-free-radicals species such as hydrogen peroxide (H2O2)^{1,2}. They tend to attack the healthy cells DNA as well as proteins and fats, causing them to deteriorate. Anti-oxidants are compounds that protect cells against the damaging effects of reactive oxygen specious, such as singlet oxygen, super oxide, peroxyl radicals, hydroxyl radicals and peroxynitrite. An imbalance between antioxidants and reactive oxygen species results in oxidative stress, leading to cellular damage. Oxidative stress has linked to cancer, ageing, atherosclerosis, and ischemia injury, inflammation and neurodegenerative diseases (Parkinson's and Alzheimer's)³.

Catharanthus Rosea, which is commonly known as 'periwinkle' or 'Vinca rosea' belongs to family Apocynaceae and is an important source of indole alkaloids, which are present in all plant parts. The physiologically important and antineoplastic alkaloids namely Vincristine and Vinblastine are mainly present in the leaves whereas antihypertensive alkaloids such as ajmalicine, serpentine, and reserpine are reported to be present in the roots⁴. Vincristine and Vinblastine alkaloids are used in the treatment of various types of lymphoma and leukemia ^{5, 6}.

Melia azedarach, the Persian Lilac is popularly known as Maha neem tree and cultivated in all stations. It is a large evergreen tree found throughout India and very similar to Neem. Leaves have been shown to contain nimbinene, meliacin, quercetrin, quercetin-3-0-b-rutinoside, kaempferol- 3-0b rutinoside, rutin and kaempferol-3-L-rhamno-Dglucoside ⁷, ⁸. Hot methanolic etract of *Melia azedarach* leaves contain dipentadecyl ketone, glycerol 1,3-bis-undec-9- enoate 2-dodec-9-enoate and glycerol tris-tridec-9-enoate ⁹. The plant is traditionally used for the treatment of leprosy, inflammations, and cardiac disorders. Its fruits extracts possess ovicidal¹⁰ and larvicidal activity¹¹. The leaf extracts also possess antiviral ¹² and antifertility activity¹³.

Brassica oleracea var. capitata (Cabbage) (Family Brassi-

caceae) is an excellent source of vitamin C. It also contains significant amounts of glutamine, an amino acid that has antiinflammatory properties. Cabbage can also be included in dieting programs, as it is a low calorie food. The present study was directed to investigate the hepatoprotective activities of Brassica oleracea L. var. capitata against simvastatin induced hepatotoxicity. There are increasing evidences that increased consumption of fruits and vegetables and intake of certain non-nutrients that are present in foods reduce the risk of various pathological events such as cancer^{14, 15} and cardio- and cerebro-vascular diseases ¹⁶. The vegetables are rich sources of many nutrients and antioxidant vitamins. Therefore, the objective of the present study was to determine the antioxidant activity phytochemical and in-vitro antioxidant activities f Melia azedarach Linn, Catharanthus Rosea(Vinca rosea) and Brassica oleracea L.var.capitata (Cabbage)

MATERIALS and METHOD Chemicals and Reagents

Folin-Ciocalteu reagent (Merck Pvt. Ltd, India), Sodium chloride (S.D. Fine Chem, India), Sodium carbonet (Merck Pvt. Ltd, India), Catechol (Himedia Lab., India), 2, 2-Diphenyle-2-picryl hydrazyl (DPPH) and Vitamin C are obtained from (Himedia Lab., India).All solutions, including freshly prepared doubled distilled water. Stock solutions of the test extracts were prepared in ethanol. Appropriate blanks were used for individual assays.

Plant collection and identification

The plant material of *Brassica oleracea var. capitata* (Cabbage) ware btain from local market, while *Melia azedarach Linn* and *Vinca rosea* L obtained from Mount Opera Garden, Near Ramoji Film City, Nalgonda Dist. The plant can be identified authenticated by Department of Botany, research office (Botanist), Anwar-ul- loom College of Pharmacy, Hyderabad.

Extraction

The leaves were dried under shade and powdered in a mechanical grinder. The powdered material (250gms) was extracted successively in Ethanol using Soxhlet apparatus at $55^{\circ}C$ for 18 h. The extracts was concentrated in vacuo and kept in a vacuum dessicator for complete removal of solvent and weighed.

Phytochemical investigation

The phytochemical studies of leaves of Brassica oleracea var.

capitata (Cabbage), Melia azedarach Linn and Vinca rosea L were performed for testing the different chemical groups such as alkaloids, tannins, glycosides and saponins etc present in ethanol, petroleum ether and aqueous extracts ^{17,18}.

Antioxidant activity of Melia azedarach, Catharanthus Rosea (Vinca rosea) and Brassica oleracea L. var capitata by using DPPH method

The antioxidant activitties of the Melia azedarach, Vinca rosea and Brassica oleracea L. var capitata on the basis of the scavenging activity of the stable 2, 2- diphenyl-2-picrylhydrazyl (DPPH) free radical was determined according to the method described by Brand-Williams et al.(1995)¹⁹ with slight modification. The following concentrations of ethanol extracts were prepared 20µg/mL, 40µg/mL, 60µmg/mL, 80µmg/mL and 10µg/mL. All the solutions were prepared with methanol. 5 ml of each prepared concentration was mixed with 0.5mL of 1mM DPPH solution in methanol. Experiment was done in triplicate. The test tubes were incubated for 30 min at room temperature and the absorbance measured at 517nm. Lower the absorbance of the reaction mixture indicates higher free radical scavenging activity. Vitamin C (0.1 mg/ml) was used as a standard and the same concentrations were prepared as the test solutions. The different in absorbance between the test and the control (DPPH in methanol) was calculated and expressed as % scavenging of DPPH radical. The capability to scavenge the DPPH radical was calculated by using the following equation.

Scavenging effect (%) = $(1-As/Ac) \times 100$

As is the absorbance of the sample at t =0 min.

Ac is the absorbance of the control at t=30 min.

RESULTS

Preliminary phytochemical screening

The ethanolic leaf extracts of Melia azedarach Linn, Vinca rosea and Brassica oleracea L. var capitata, Linn, showed the presence of Alkaloids, Tannins, Phenols, Flavonoids, Glycosides and Saponins.

Steroids and Terpenes are absent in Vinca rosea and while saponins are absent in Brassica oleracea L. var capitata (Table 1).

Yields of ethanolic extracts of Melia azedarach, Vinca rosea and Brassica oleracea L. var capitata

The variation in yields of ethanolic leaf extracts of Melia azedarach, Vinca rosea and Brassica oleracea L. var capitata were 22.13gm, 23.80gm and 27.98gm respectively, shown in Table .2

DPPH free radical scavenging activity of Melia azedarach, Vinca rosea and Brassica oleracea L. var capitata

The result shows that the ethanol extract of Melia azedarach Linn and Brassica oleracea L. var capitata exhibited the highest radical scavenging activity with 71.42±0.04 and 75.65±0.06 at 100 (µg/ml) concentrations of extracts. While Vinca rosea exhibited the moderate radical scavenging activity with 56.43±0.05 at 100($\mu g/ml)$ concentration of extract. Fig. 1 and Table.3 Shows the antioxidant activities of different plants extracts.

IC₅₀ value IC50 value is defined as the concentration of substrate that causes 50% loss of the DPPH. The $\rm IC_{50}$ value of Melia azedarach Linn , Vinca rosea and Brassica oleracea L. var capitata was calculated as 0.0054µg/ml, 0.0074µg/ml 0.0042 µg/ml respectively. The extracts of Melia azedarach and Brassica oleracea L. var capitata showed highest free radical scavenging activity among with the lowest $\rm IC_{50}$ While Vinca rosea exhibited the moderate radical scavenging activity with the highest IC_{50} as shown in Table 2 and Fig 2.

Table	1. Phytochemical analysis of Ethanolic extracts of
Melia	azedarach Linn, Vinca rosea and Brassica oleracea
L. var	capitata, Linn.

SI. No.	Phytochemical Tests	Melia Azedarach	Vinca rosea	Brassica oleracea
1	Alkaloids	+	+	+
2	Steroids	+	-	+
3	Tannins	+	+	+
4	Phenols	+	+	+
5	Flavonoids	+	+	+
6	Glycosides	+	+	+
7	Saponins	+	+	+
8	Terpenes	+	-	-

+= indicates Present of compounds

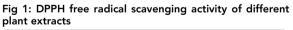
-=indicates absence of compounds

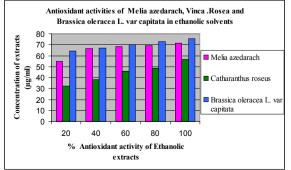
Table-2: Crude extract and IC₅₀ Value in Melia azedarach, Catharanthus roseus and Brassica oleracea L. var capitata.

	Ethanolic Solvent		
Extracts	Crude Extracts (gm)	IC 50Value (µg/ml)	
Melia azedarach	22.13	0.0054	
Catharanthus roseus	23.80	0.0074	
Brassica oleracea L. var capitata	27.78	0.0042	

Table-3: Antioxidant activities of Melia azedarach, Catharanthus roseus and Brassica oleracea L. var capitata in ethanolic solvents.

Concentration	% Antioxidant activity of Ethanolic extracts			
of extracts (µg/ml)	Melia azedarach	Vinca rosea	Brassica oleracea L. var capitata	
20	54.98±0.05	32.23±0.03	64.31±0.01	
40	66.65±0.07	38.26±0.06	67.13±0.02	
60	68.38±0.06	45.89±0.03	69.99±0.05	
80	69.11±0.03	48.76±0.07	72.76±0.06	
100	71.42±0.04	56.43±0.05	75.65±0.06	

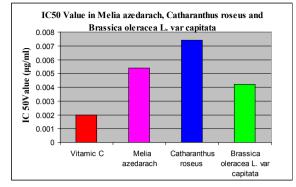




RESEARCH PAPER

Volume : 3 | Issue : 7 | July 2013 | ISSN - 2249-555X

Fig 2: IC_{50} Value of different plant extracts compared with Vitamin C.



DISCUSSION

The result of the present study showed that the ethanolic leaf extracts of Melia azedarach Linn and Brassica oleracea L. var capitata exhibited the greatest anti-oxidant activity, while Vinca rosea exhibited moderate anti-oxidant activity. The high scavenging property may be due to hydroxyl groups existing in the phenolic compounds. It is reported that phenols are responsible for the variation in the antioxidant activity of

the plant^[20]. They exhibit antioxidant activity by inactivating lipid free radicals or preventing decomposition of hydroperoxides into free radicals ^{21, 22}. It is reported that -OH groups in phenolic compounds are thought have a significant role in antioxidant activity 23. It is also reported that antioxidant activity of phenolic compounds is mainly due to their redox properties²⁴. Methanol and ethanol has been proven as effective solvent to extract phenolic compounds^[25]. Ethanol is preferred for the extraction of antioxidant compounds mainly because its lowers toxicity ²⁶. Vegetables contain a wide variety of biologically active, non-nutritive compounds known as phytochemicals. This is often attributed to the antioxidants such as vitamin C, E, carotenoids, lycopenes and flavonoids that prevent free radical damages²⁷⁻²⁹. These phytochemicals impart health benefits beyond basic nutrition³⁰.

CONCLUSION

The result of the present study showed that the extracts of Melia azedarach Linn and Brassica oleracea L. var capitata exhibited the greatest anti-oxidant activity in comparison to Vinca rosea. The high scavenging property of Melia azedarach Linn and Brassica oleracea L. var capitata may be due to hydroxyl groups existing in the phenolic compounds chemical structure that can provide the necessary components as a radical scavenger.



REFERENCE 1. Halliwell B. How to characterize anantioxidant: an update. Biochem Soc Symp 1995; 61: 73-101. | 2. Squadriato GL, Pelor WA. Oxidative chemistry of nitric oxide: The roles of superoxide, peroxynitrite, andcarbon dioxide. Biol Med 1998; 25: 392-403. | 3. Donald RB, Cristobal M. Antioxidant Activities of Flavonoids. PhD thesis submitted to Department of Environmental and Molecular Toxicology, Oregon State University; 1987. | 4. Mishra P., Uniyal G.C., and Sharma S., Pattern of diversity for morphological and alkaloid yield related trades among the perivinkle Catharanthus rose accessions collected from in and around Indian Subcontinent, Genetic Res Crop Evol., 2001, 48, 273-286. [5. Farnsworth N.R., Svoboda G.H., Blomster R.N., Antiviral activity of selected Catharanthus alkaloids, J Pharm. Sci., 1968, 57, 2174-2175. [6. Svoboda G.H., Blake D.A., The phyto-chemistry and pharmacology of Catharanthus roseus (L) G. Don. Inc. In: Taylor, W.J., Farnsworth, N.R. (eds.): The Catharanthusalkaloids. Marcel Deckr, New York, 1975, 45-84. [7. P. C. Sharma, M.B. Yelne, T. J. Dennis, Data on Medicinal plants used in Ayurveda, (Documentation and Publication Division, Central Council for Research in Ayurveda and Siddha, New Delhi, 2001) pp. 389-406. [8. C.P. Khare, Encyclopedia of Indian Medicinal Plants, (Springer, Germany) pp.305-306. [9. P. Suhag, Merra and S.B. Kalidhar. Phytochemical investigation of Melia azedarach leaves. J. Med. Aromatic Plant Sci. 25(2): 397-399 (2003). [10. Corpinella MC, Miranda M, Almiron WR, Ferrayoli CG, Almedia FL, Palacios SM. In vitro pediculicidal and ovicidal activity of an extract and oil from fruit of Melia azedarach. J Am Acad Dermatol 2007; 56: 250-6.] 11. Wandscheer CB, Duque JE, da silva MA, et al. Larvicidal action of ethanolic extracts from fruits endocarps of Melia azedarach and Azadirachta indica against the dengue mosquito Aedes Aegypti. Toxicol 2004; 44: 829-35. | 12. Descalzo AM, Coto C. Inhibition of the pseudorabies virus (scis herpesvinyl) by an andvital agent isolated from the leaves of melia azedarach 2004; 44: 829-35. | 12. Descalzo AM, Coto C. Inhibition of the pseudorabies virus (scis herpesvinyl) by an andvital agent isolated from the leaves of melia azedarach Rev. Argent Microbial 1989; 21: 133-40. | 13. Choudhary DN, Singh JN, Verma SK, Singh BP. Antifertility effects of leaf extracts of some plants in male rats. Indian J Exp Biol 1990; 28: 714. | 14. Goodwin JS, Brodwick M. J Am Diet Assoc 1996; 96: 1027-1039. | 15. Rimm EB, Ascherio A, Giovannucci E, Spiegelman D, Stampfer MJ, Willett WC. JAMA 1996; 275:447-451. | 16. Trease GE, Evans WC, A Text book of Pharmacognosy, 11th edition, Bailliere Tidall, London, 1978, 530. | 17. Kokate, C.K., Purohith, A.P.&Gokhale, S.B. (1990). Pharmacognosy, Nirali Prakashan, Pune, 120. | 18. Khandelwal, K.R. (2006). Practical Pharmacognosy techniques and experiments, 16 Edition, Nirali Prakashan, 149-156. | 19. Brand-Williams, W., Cuvelier, M.E., and Berset, C. Use of free radical method to evaluate antioxidant activity. Lebensmittel Wissenschaft and Technologie 1995; 28(1):25-30. | 20. Cai Y, Luo Q, Sun M, Corke H Antioxidant activity and phenolic compounds of 112 traditional Chinese medicinal plants associated with anticancer. Life science 2004;74:2157-84. | 21. Pokorny J, Yanishlieva N, Gordon M. Antioxidants in food, Practical Applications, Cambridge Woodhead publishing limited 2001;72(5):145-71. | 22. Pitchaon M, Suttajit M, Pongsawatmani R. Assessment of phenolic content and free radical acevenging capacity of some Thai indigenous plants. Food Chem 2007;100:1409-18. | 23. Arumugam P, Ramamurthy P, Santhiya ST and Ramesh A (2006). Antioxidant activity measured in different solvent fractions obtained from Mentha spicata Linn. - An analysis by ABTS +. declorization assav. Asia Pac. J. Clin. Nutr. 119-124. | 24. Rahman K in different solvent fractions obtained from Mentha spicata Linn.: An analysis by ABTS.+ decolorization assay. Asia Pac. J. Clin. Nutr. 119-124. | 24. Rahman K (2007). Studies on free radicals, antioxidants, and cofactors. Clin Interv Aging. 2(2): 219–236. | 25. Siddhuraju P, Becker K.Antioxidant properties of various extracts of total phenolic constituents from three different agroclimatic origins of drumstick tree (Moringa oleifera Lam.) leaves. Journal of Agriculture and Food Chemistry 2003;51:2144-55. | 26. Karadeniz F, Burdurulu HS, Koca N, Soyer Y. Antioxidant activity of selected fruits and vegetables grown in Turkey. Journal of Agriculture and Food Chemistry 2005; 29, 297-303. | 27. Steinberg D. Circulation 1991;84:1420-1425. | 28. Willett WC. Am J Clin Nutr 1994;59: S265-S269. | 29. Oomah BD, Mazza G, Functional foods, In: Francis FJ (ed) The Wiley encyclopedia of science and technology, 2nd edition, Wiley, New York, 2000, 1176–1182. | 30. Sheetal Gupta, Jamuna Prakash. Plant Foods Hum Nutr 2009; 64:39-45.