



Isolation and Characterization of Phosphate Solubilizing Pseudomonas Sp. from Rhizospheric Soil of Rice

KEYWORDS

Phosphate solubilization, Rhizobacteria, Fluorescent pseudomonas, Macronutrients

Manidipa Roy

Department of Life science & Bioinformatics, Assam University, Silchar

G.D.Sharma

Vice Chancellor, Bilaspur University, Chattisgarh

Ch.V. Ramana

Department of Plant sciences, University of Hyderabad, Hyderabad

ABSTRACT Phosphorus (P) is an essential macronutrient for the growth of plants. However, in most soils a large portion of phosphorus becomes insoluble and therefore, unavailable to plants. Thirty one Pseudomonas sp. were isolated from rhizospheric soil of rice. Out of thirty one nine isolates were able to solubilize $\text{Ca}_3(\text{PO}_4)_2$ on Pikovskya's agar medium. A total six sp. of phosphate solubilizing fluorescent pseudomonads which showed fluorescence under UV radiation ($\lambda=365\text{nm}$) viz. *Pseudomonas aeruginosa*, *P. fluorescens*, *P. putida* and *P. plecoglossicida* and three species of phosphate solubilizing non-fluorescent pseudomonas were isolated. Isolate MGR38 was tentatively identified as *Pseudomonas fluorescens* which showed 83.33% efficiency in phosphate solubilization and the other isolate MGR31, MGR37, MGR39, MGR40, MGR41, MGR42, MGR43 and MGR44 were showed 33.33, 57.14, 50, 50, 16.16, 40, 60 and 80 % efficiency of phosphate solubilization respectively.

INTRODUCTION

Phosphorus is one of the most essential macronutrients of plants and its deficiency is a severe constraint to crop production. Plants absorb only inorganic form of phosphorus and the level of inorganic phosphorus in soil is very low normally 1ppm or less (Goldstein,1994). Most of the phosphorus in soil is present in insoluble mineral forms such as hydroxyapatites, oxyapatites and appetites. Rice field soils possess considerable accumulation of phosphorus due to regular application of chemical phosphate fertilizers and a large proportion of the applied fertilizer is converted in to insoluble form and become unavailable to plants (Rodriguez & Fraga, 1999). Soil microorganisms have the ability to solubilize insoluble phosphates and to improve soil health and fertility Jaharamma, Badri, and Sakthivel (2009). Phosphate solubilizing microbes have been reported for plant growth promoting and enhancing yield Kapoor, Mishra, and Kuhreja (1989); Rodriguez and Fraga (1999). Microorganisms play an important role in the availability of soil phosphate to the root system and enhance the mobilization of phosphate in soil (Richardson, 2001). Production of organic acids and the enzymes phosphatase and phytase by fluorescent pseudomonads seems to be the main cause of phosphate solubilization Jaharamma et al. (2009). Considering the plant health and crop productivity it is important to study the diversity of phosphate solubilizing Pseudomonas sp. as these native strains may be used as bioinoculants for sustainable agriculture which will be cost effective and ecofriendly.

MATERIALS AND METHODS**Sample collection**

Rhizospheric soil samples of rice (*Oryza sativa* L.) were collected in pre sterilized polythene bags from Sone beel wet land, Assam,India. Samples were stored at 40C within 24h of collection before being processed.

Isolation of Pseudomonas species

Root samples were shaken vigorously to remove loosely adhering soil. 10g of tightly attached soil with roots were taken in a conical flask containing 90ml of 0.1M $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ buffer and kept for 10min on a rotary shaker at 150 rpm. Resulting suspensions were tenfold diluted, 0.1 ml aliquots of 10^{-6} dilution was inoculated on sterilized King B agar medium King,Ward,and Pandey (1954) and incubated for 2-3days at 28°C. After incubation, fluorescent Pseudomonads were identified under UV light for few seconds at 365 nm and colonies exhibited bluish green fluorescence were picked up and streaked on sterilized King B agar plate to obtain

pure cultures. Stock cultures were made in Luria Bertani (LB) broth containing 50% (w/v) glycerol and stored at -86°C.

Qualitative estimation of phosphate solubilization

To detect phosphate solubilizing bacteria, strains were spot inoculated on to Pikovskaya's agar medium (Pikovskaya,1948) containing (per liter): 0.5g yeast extract, 10g dextrose, 5g $\text{Ca}_3(\text{PO}_4)_2$, 0.5g $(\text{NH}_4)_2\text{SO}_4$, 0.2g KCl, 0.1g $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$, 0.0001g $\text{MnSO}_4 \cdot \text{H}_2\text{O}$, 0.0001g $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$ and 15g agar and incubated for 3- 4 days at 28°C. Formation of clear zone around the colonies indicated phosphate solubilization by the bacteria. Diameter of clear zone around the colonies was recorded. Solubilization efficiency of bacterial isolates were calculated based on the following formula Nguyen, Yan, LeTacon, and Lapayrie (1992).

Identification of bacterial isolates

Cultural and biochemical characterization was done by the methods of Cappuccino and Sherman (2004) and were tentatively identified by following Bergey's Manual of determinative Bacteriology Holt, Krieg ,Sneath, Staley, and Williams (1994).

RESULTS AND DISCUSSION

A total of thirty one Pseudomonas sp. were isolated from rhizospheric soil of rice. Out of thirty one nine isolates were phosphate solubilizer (Fig 1). Among them four were fluorescent pseudomonads which showed fluorescence under UV radiation ($\lambda=365\text{nm}$) and five were Pseudomonas sp. All these phosphate solubilizing isolate MGR31, MGR37, MGR38, MGR39, MGR40, MGR41, MGR42, MGR43 and MGR44 were tentatively identified as Pseudomonas sp. , *P. aeruginosa*, *P. fluorescens*, *P. putida*, *P. sp.*, *P. sp.*, *P. plecoglossicida*, *P. sp.*, and *P. sp.* respectively on the basis of morphological (Table 1) and biochemical characteristics (Table 2). All the isolates were gram negative, rod shaped and were produced either yellowish green or light green or green pigment on Kings B agar medium. Among all the phosphate solubilizer the isolates MGR38 showed highest (83.33%) and MGR41 showed least (16.66%) efficiency in phosphate solubilization and the other MGR31, MGR37, MGR39, MGR40, MGR42, MGR43 and MGR44 was found 33.33, 57.14, 50, 50, 40, 60 and 80 % respectively (Fig 2). The isolate MGR38 produce enzyme like amylase, cellulase, protease and gillatinase. It has been reported that strains of Pseudomonas which produce protease, cellulase, pectinase, amylase and gillatinase solubilize inorganic phosphate and 18% of the

fluorescent pseudomonads are efficient phosphate solubilizer Naik, Raman,Badri Narayan, and Sakhthivel (2008).

Table 1.Morphological characteristics of the bacterial isolates

SL. NO	STRAIN NAME	SIZE	COLOUR	FORM	OPACITY	MARGIN	ELEVATION	FLURESCENCE
1	MGR31	Small	Light green	Circular	Translucent	Entire	Flat	+
2	MGR37	Small	Greenish yellow	Irregular	Translucent	Entire	Convex	+++
3	MGR38	Small	Green	Circular	Translucent	Undulate	Flat	+++
4	MGR39	Moderate	Light green	Circular	Translucent	Entire	Flat	-
5	MGR40	Small	Light green	Circular	Opaque	Entire	Convex	-
6	MGR41	Small	Yellowish green	Circular	Opaque	Entire	Convex	++
7	MGR42	Small	Yellowish green	Circular	Translucent	Undulate	Convex	+++
8	MGR43	Moderate	Light green	Circular	Opaque	Entire	Raised	+++
9	MGR44	Moderate	Yellowish green	Circular	Opaque	Entire	Convex	+

Note: +++ = high fluorescence, ++ = medium fluorescence, + = weak fluorescence, - = no fluorescence

Table 2.Biochemical characteristics of the bacterial isolates

PARAMETERS	MGR31	MGR37	MGR38	MGR39	MGR40	MGR41	MGR42	MGR43	MGR44
Gram stain	-, Rod	-, Rod	-, Rod	-, Rod	-, Rod	-, Rod	-, Rod	-, Rod	-, Rod
Motility	+	+	+	+	+	+	+	+	+
FERMEN-TATION	Lactose	-	-	-	-	-	-	-	-
	Dex-trose	-	+	+	+	-	-	-	-
	Fructose	-	-	-	+	-	-	-	-
ENZYME PRODUCTION	Catalase	+	+	+	+	+	+	-	-
	Amylase	-	-	-	-	-	+	-	-
	Protease	+	+	-	-	+	-	+	+
	Gillati-nase	+	+	-	-	+	+	-	+
	Cellu-lase	-	+	-	-	-	-	-	-
	Pecti-nase	-	-	+	+	-	-	-	-
	Urase	-	+	-	-	-	-	-	-
	Oxidase	+	+	+	+	+	+	+	-
H ₂ S Production	+	-	-	-	+	-	+	+	-
Indole Production	+	-	-	-	+	-	+	+	+
Citrate Utilization	+	+	+	+	+	+	+	+	+
NO ₃ Reduction	-	+	+	+	+	+	+	-	-
MR-Reaction	-	-	+	-	+	-	-	-	+
VP-Reaction	+	-	-	-	-	-	-	-	+
Arginine Hydrolysis	+	+	+	+	+	+	+	+	+
Lipid Hydrolysis	-	+	+	+	+	+	+	+	-
Casein Hydrolysis	-	+	+	+	+	+	+	+	-
GROWTH AT	4°C	+	-	-	+	+	-	+	+
	41°C	-	+	-	-	-	-	-	-
Tantative identifica-tion	P.sp.	P.aeruginosa	P.fluorescens	P.putida	P.sp.	P.sp.	P.plecoglossicida	P.sp.	P.sp.

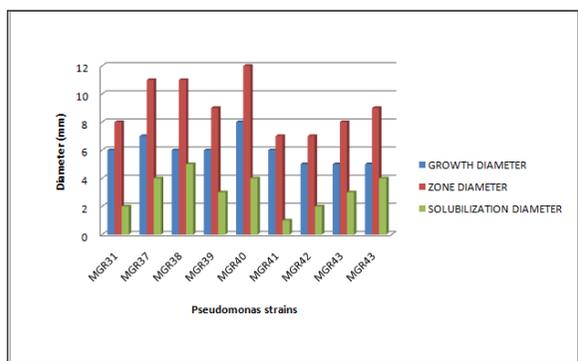


Fig.1: Qualitative estimation of Phosphate solubilization

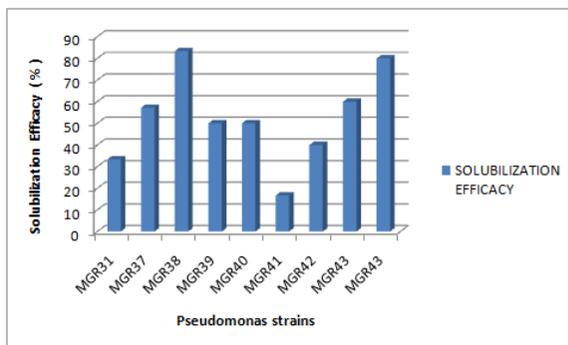


Fig.2: Phosphate solubilization efficiency of Pseudomonas strains

CONCLUSION

Soil fertility is one of the most important factor for crop production . The excess use of chemical pesticides in agricultural fields minimizes the solubility of chemical fertilizers and making it unavailable to plants. Plant growth and yield is often limited by insufficient phosphate availability. Our study revealed that genus *Pseudomonas* play an important role in phosphate solubilization . Considering the phosphate solubilizing efficiency it may be used as bioinoculants for sustainable agriculture.

Acknowledgement:

The authors are thankful to the Head, Department of Life science & Bioinformatics, Assam University, Silchar, India for providing laboratory facilities.

REFERENCE

1. Cappucino, J.G., & Sherman, N. (2004). *Microbiology: A Laboratory manual* (6th ed.). Singapore, Pearson Education publication. | 2. Goldstein, A.H. (1994). Involvement of the quinoprotein glucose dehydrogenase in the solubilization of exogenous phosphates by gram-negative bacteria. In: Torriani-Gorini, A., Yagil, E., Silver, S. (Ed.). *Phosphate in Microorganisms, Cellular and Molecular Biology*, (pp.197-203). Washington, D.C: ASM Press. | 3. Holt, J.G., Krieg, N.R., Sneath, P.H.A., Staley, J.T., Williams, S.T. (1994). *Bergey's Manual of Determinative Bacteriology* (9th ed.). Baltimore, The Williams & Wilkins Co. | 4. Jaharamma, M., Badri Narayan, K., & Sakthivel, N. (2009). Genetic and functional diversity of Phosphate Solubilizing fluorescent *Pseudomonas* and their simultaneous role in promotion of plant growth and soil health. In: Mahoney, C.L., & Springer, D.A. (Ed.). *Genetic diversity*, (pp.1-8). | 5. King, E.O., Ward, M.K., & Raney, D.E. (1954). Two simple media for demonstration of pyocyanin and fluorescein. *J. Lab. Clin. Med.*, 44, 301-307. | 6. Kapoor, K.K., Mishra, M.M., & Kuhreja, K. (1989). Phosphate solubilization by soil microorganism-A review. *Indian Journal of Microbiology*, 29, 119-127. | 7. Nguyen, C., Yan, W., Le Tacon, F., & Lapayrie, F. (1992). Genetic variability of phosphate solubilizing activity by monocaryotic and dicaryotic mycelia of the ectomycorrhizal fungus *Laccaria bicolor* (Maire) P.D. Orton. *Plant and Soil*, 143:193-199 doi:10.1007/BF00007873. | 8. Pikovskaya, R.I. (1948). Mobilization of phosphorus in soil in connection with vital activity of some microbial species. *Mikrobiologiya*, 17, 367-370. | 9. Rodriguez, H., & Fraga, R. (1999). Phosphate solubilizing bacteria and their role in plant growth promotion. *Biotechnol Adv.*, 17, 319-339. | 10. Richardson, A.E. (2001). Prospects for using soil microorganisms to improve the acquisition of phosphorus by plants. *Australian Journal of Plant Physiology*, 28, 8797-8906. | 11. Ravindra Naik, P., Raman, G., Badri Narayanan, K., & Sakthivel, N. (2008). Assessment of genetic and functional diversity of phosphate solubilizing fluorescent pseudomonads isolated from rhizospheric soil. *BMC Microbiology*, 8:230 doi:10.1186/1471-2180-8-230. |