

Comparison of Haematology in Various Physiological States in Sahiwal Cattle

KEYWORDS	Haematology, Lactation, Pregnancy, Sahiwal cows		
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ABSTRACT Twenty seven Sahiwal cows were used in the experimentation and were divided into 3 groups (n=9). Animals were divided into three groups as Pregnant dry cows, Non Pregnant Lactating cows and Non Pregnant Dry cows. The highest Hb concentration (10.42 ± 0.42 gm/dl) was recorded in group-1, while the lowest (9.26 ± 0.34 gm/dl) was observed in group-2. The highest and lowest RBCs count and PCV ($5.83 \pm 0.39 \times 106/\mu$ l, $32.67 \pm 1.36\%$ and $4.57 \pm 0.27 \times 106/\mu$ l and $29.26 \pm 1.11\%$) were recorded in group-3 and group-1 respectively. The highest MCV, MCH and MCHC (65.10 ± 2.27 fl, 23.36 ± 1.51 pg and 35.58 ± 1.04 gm/dl) were noted in group-3. The highest and lowest ESR (7.58 ± 1.15 mm/24 hours and 5.12 ± 0.67 mm/24 hours) was recorded in group-3 and group 2 respectively. Highest and lowest ($8.72 \pm 1.32 \times 103/\mu$ l) and $7.61 \pm 0.87 \times 103/\mu$ l) was observed in group-3 and group 2 respectively. Highest lymphocyte count ($65.24 \pm 3.00\%$) was observed in group-2, while lowest count ($64.29 \pm 2.91\%$) was in group-1.

INTRODUCTION

Physiological equilibrium is maintained mainly by the blood in the body (Geneser, 1986) but this equilibrium is altered in various physiological conditions changing the homeostasis of animals. Hence, the haematological values during different physiological situations should be known for the diagnosis of various pathological and metabolic disorders, which can adversely affect the productive and reproductive performance of cows, leading to heavy economic losses (Dutta et *al.*, 1988).

MATERIALS AND METHODS

Twenty seven Sahiwal cows of different age groups were used in the experimentation. These animals were maintained and housed under similar conditions of feeding and management. The animals were fed daily 40-50 Kg of green fodder and 2-3 Kg of concentrate mixture; containing 15% crude protein and 65% total digestible nutrients. These animals were divided into 3 groups, having 9 animals in each group. The grouping was done depending upon their physiological conditions as follows: group1- Pregnant dry cows, group 2- Non Pregnant Lactating cows and group 3-Non Pregnant Dry cows.

Fifteen ml of venous blood from the Jugular vein of each animal was collected, using one mg of disodium salt of ethylenediaminetetraacetic acid per ml of blood as an anticoagulant (Sastri, 1985). Haematological parameters were studied according to the methods described by (Sastri 1985). The detail of the parameters studied is as follows: haemoglobin (Hb) concentration, red blood cells (RBC) count, packed cell volume (PCV), mean corpuscular volume (MCV), mean corpuscular haemoglobin (MCH), mean corpuscular haemoglobin concentration (MCHC), erythrocyte sedimentation rate (ESR), white blood cells (WBC) count and differential leucocytic count (DLC).

Statistical analysis

The arithmetic means (\pm SE) of haematological parameters in different groups were calculated. Further analysis was done using analysis of variance (ANOVA) technique (Steel and Tor-

rie, 1984) and significant results were subjected to Duncan's multiple range (DMR) test (Duncan, 1955). The differences were considered statistically significant at P<0.05.

RESULTS

The results for various haematological parameters of three physiological states of Sahiwal cattle are given in Table 1. The highest Hb concentration was recorded in Pregnant dry cows (Group-1), while the lowest values were observed in non-pregnant lactating cows (Group-2), the difference was statistically significant (P<0.05). Similarly, the highest RBCs count and PCV were recorded in Non Pregnant Dry cows (Group-3), while the lowest values were observed in pregnant dry cows (Group-1), the difference being statistically significant (P<0.05). In the current study, the highest MCV, MCH and MCHC were noted in Pregnant dry cows (Group-1) and the lowest values were observed in Non Pregnant Dry cows (Group-3), the differences were statistically significant (P<0.05). In the present study, the highest ESR was recorded in pregnant dry cows (Group- 1) and the lowest values were observed in Non Pregnant Lactating cows (Group-2), the difference was statistically significant (P<0.05). The highest WBCs count was recorded in Non Pregnant Dry cows (Group-3) and the lowest values were observed in Non Pregnant Lactating cows (Group-2), the difference was significant (P<0.05). Significantly (P<0.05) higher lymphocyte count was observed in Non Pregnant Lactating cows (Group-2) compared to Pregnant dry cows (Group-1).

DISCUSSION

Ahmad (1995) reported PCV 28.4 \pm 0.61 to 31.4 \pm 0.50%, Hb 9.7 \pm 0.30 to 11.1 \pm 0.30 gm/dl and RBC count 4.7 \pm 0.41 to 7.0 \pm 0.42×10⁶/µl in Sahiwal cows during last trimester of pregnancy (pregnant dry cows). These values are closely related to the values of the present study. Unlike our study, Steinhardt et al. (1994) reported decrease in Hb with advancing lactation and pregnancy, which increased at parturient stage. Kumar and Pachauri (2000) reported highest MCV and MCH, and lowest MCHC in non-pregnant dry cows compared to other groups which are closely related to our study. Ahmad (1995) reported WBC count of 6.8 \pm 0.28 to 8.3 \pm

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0.29 10³/µl, lymphocytes 55.5 ± 1.96 to 65.3 ± 2.49%, monocytes 3.50 ± 0.87 to $3.90 \pm 0.81\%$, neutrophils 21.1 ± 1.12 to $30.0 \pm 3.69\%$, eosinophils 5.50 ± 1.66 to $8.00 \pm 0.71\%$ and basophils 0.20 ± 0.25 to 0.40 ± 0.18% and ESR 5.9 ± 0.40 to 17.1 ± 1.89 mm/24hr in Sahiwal cows during last trimester of pregnancy (pregnant dry cows) on the other hand, the differences of monocytes, neutrophils, eosinophils and basophils between all the groups were statistically non-significant. Most of these values are closely related to the present study. Pereira et al. (1987) also reported that there was an increase in the number of leukocytes during gestation. Discrepancies in values for various haematological parameters between our findings and previous studies may be explained by differences in sampling interval, methods used, numbers of cows sampled, and/or degree of metabolic disturbances. Moreover, genetic differences between cows (Mallard et al., 1998) and environmental conditions of the present study might have played a role for the differences with other studies. In the present study, the reasons for immune suppression in pregnant cows are not fully known, but several factors such as management, feeding and changes in hormonal levels may be involved (Meglia et al., 2005). According to Meglia et al. (2005), suppression of leukocyte functions in dairy cows has been associated with negative energy balance around calving and in early lactation. Sometimes blood leukocyte numbers and their functions change considerably around parturition, resulting in suppression of the immune response from a few weeks before to a few weeks after calving (Mallard et al., 1998). Lymphocytes decrease around parturition mainly due to reduced lymphocyte proliferation (Saad et al., 1989).

CONCLUSION

The responses of the studied haematology in various physiological states in sahiwal cattle show that the dairy cows are in a particular quantum of stress during different physiological stages ranging from pregnancy to lactation. These stressors although inevitable during pregnancy and lactation, can be abated by good nutrition and proper management of dairy and pregnant cows.

Table 1: Haematological	parameters (mean ± SE) in sahiwal cattle at different physiological	stages

Table 1. The match of gital parameters (mean ± 52) in samwar cattle at unrerent physiological					
Pregnant	Non Pregnant	Non Pregnant			
dry cows	Lactating cows	dry cows			
(Group1)	(Group2)	(Group3)			
$10.42 \pm 0.42^{\text{abc}}$	9.26 ± 0.34°	10.31 ± 0.41 ^{bc}			
4.57 ± 0.27°	$5.36 \pm 0.57^{\rm abc}$	5.83 ± 0.39^{ab}			
29.26 ± 1.11 ^b	29.81 ± 1.79 ^b	32.67 ± 1.36^{ab}			
65.10 ± 2.27^{ab}	58.27 ± 2.46^{bc}	57.87 ± 2.15°			
23.36 ± 1.51 ^{ab}	18.75 ± 1.64 ^{bc}	18.59 ± 1.43°			
35.58 ± 1.04 ^a	31.72 ± 1.56 ^{bc}	31.76 ± 1.22°			
7.58± 1.15ª	5.12 ± 0.67^{ab}	5.22 ± 0.36^{ab}			
8.10 ± 0.76^{ab}	7.61 ± 0.87^{ab}	8.72 ± 1.32^{ab}			
64.29 ± 2.91 ^{ab}	65.24 ± 3.00^{ab}	64.41 ± 6.63^{ab}			
6.29 ± 0.88^{a}	$6.41 \pm 0.78^{\circ}$	5.69 ± 1.16 ^a			
22.88 ± 2.20 ^a	23.32 ± 2.97ª	25.43 ± 5.00 ^a			
5.39 ± 1.24ª	4.31 ± 0.54^{a}	3.83 ± 1.57ª			
$1.10 \pm 0.35^{\circ}$	0.80 ± 0.20^{a}	0.70 ± 0.26^{a}			
	$\begin{array}{c} Pregnant \\ dry cows \\ (Group1) \\ 10.42 \pm 0.42^{abc} \\ 4.57 \pm 0.27^c \\ 29.26 \pm 1.11^b \\ 65.10 \pm 2.27^{ab} \\ 23.36 \pm 1.51^{ab} \\ 35.58 \pm 1.04^a \\ 7.58 \pm 1.15^a \\ 8.10 \pm 0.76^{ab} \\ 64.29 \pm 2.91^{ab} \\ 62.9 \pm 0.88^a \\ 22.88 \pm 2.20^a \\ 5.39 \pm 1.24^a \end{array}$	$\begin{array}{c c c c c c c c c c c c c c c c c c c $			

Values bearing different superscripts in a row differ significantly (P<0.05) but sharing at least one superscript in a row differ nonsignificanlty

REFERENCE

Ahmad, I. (1995). Antibody titer and hematology following vaccination and immunopotentiation of Sahiwal cows in last trimester of pregnancy. PhD Thesis, Univ. Agri., Faisalabad, Pakistan. | Duncan, D. B. (1955). Multiple range and Multiple F test. Biometrics, 11: 1-42. | Dutta, J. C., Baruah, R. N., Dutta, L. and Talukar, S. C. (1988). Blood biochemical studies in anoestrus and normal cyclic cattle. Indian Vet. J., 65: 239–241 | Esievo, K. A. N. and Moore, W. E. (1979). Effect of dietary protein and stage of lactation on the haematology and erythrocyte enzymes activities of high producing dairy cattle. Res. Vet. Sci., 26: 53- 58. | Geneser, F. (1986). Textbook of Histology. 1st Ed., Munksgaard, Copenhagen, Denmark. | Kumar, B. and Pachauri, S. P. (2000). Haematological profile of crossbred dairy cattle to monitor herd health status at medium elevation in Central Himalayas. Res. Vet. Sci., 69: 141-145. | Mallard, B. A., Dekkers, J. C., Ireland, M. J., Leslie, K. E., Sharif, S., Lacey Vankampen, C., Wagter L. and Wilkie, B. N. (1998). Alteration in immune responsiveness during the peripartum period and its ramification on dairy cows and calf health. J. Dairy Sci., 81: 585–595. | Meglia, G. E., Johannisson, A., Agenas, S., Holtenius K. and Waller, K. P. (2005). Effects of feeding Intensity during the dry period on leukocyte and lymphocyte sub-populations, neutrophil function and health in periparturient dairy cows. Vet. J., 169: 376–384. | Pereira, J. L., Orden, M. A., Fernandez del Palacio, M. J., Barreiro, A., Diez, I. and Gonzalo, J. M. (1987). Haematological variations related to gestation and age in the autochthonous bovine breed Blanca Cacerena. Anales de Veterinaria de Murcia, 3: 93-97. | Richard, L. A. (1954). United States Lab. Staff. Agri. Hand Book. No 66 United States Depart Agri, p 129. | Saad, A. M., Concha C. and Astrom, G. (1989). Alterations in neutrophil phagocytosis and lymphocyte blastogenesis in dairy cows around parturition. J. Vet. Med., 36: 337–345. | Sastri, G. A. (1985). Veterinary Clinical Pathology. Salish Kumar Jain for CBS Publishers & Distributors, Delhi-32, India. | Steel, R. D. G. and Torrie, J. H. (1984). Principles and Procedures of Statistics. 2nd Ed., Mc-Graw Hill Koga Kusha Ltd. Book Co. Inc. New York, USA. | Steinhardt, M., Thielscher, H. H., von Horn, T., von Horn, R., Errogassen, K., Ladewig, J. and Smidt, D. (1994). The hemoglobin concentration in the blood of dairy cattle of different breeds and their offspring during the peripartum period. Tierarztl Prax., 22: 129-135. |