



Larvicidal Action of *Garcinia Gummi-Gutta*. Robs. Var. *Gummi-Gutta* on Dengue Victim *Aedes Aegypti*

KEYWORDS

Aedes aegypti, flavonoids, *Garcinia gummi-gutta*, larvicidal, leaf extract, percentage mortality

Dhanya. P

Department of Zoology, St. Thomas College, Pala, Kottayam, Kerala.

Benny. P. J.

Department of Zoology, St. Thomas College, Pala, Kottayam, Kerala.

ABSTRACT Mosquitos are the vectors of many vector-borne diseases that outbursts at rainy seasons. Of these dengue fever is a common one caused by *Aedes aegyptii*. As a solution, many experiments were conducted in this study to evaluate the larvicidal potential of *Garcinia gummi-gutta*. Crude methanolic leaf extracts of *Garcinia gummi-gutta* were tested on *Aedes aegypti* under *in vitro* condition. The results indicated that the leaf extract of *Garcinia gummi-gutta* showed more cytotoxic effect (inhibitory) on the third or fourth instar larva at higher concentrations as compared to control, while the lower concentrations of leaf extract showed negative larvicidal (stimulatory) effect on growth parameters of mosquito larva. Presence of flavonoids have been observed and, LC/MS analysis was carried out on methanol leaf extract to find out the active component.

Introduction

Mosquitos are insects which are capable of transmitting diseases like malaria, filariasis, dengue, Japanese encephalitis, etc. [Becker et al., 2003]. These diseases are disastrous to human life. Among these diseases, dengue remains the most serious vector-borne disease affecting millions of people throughout the world and it is reported to infect more than 100 million people every year in more than 110 countries in the tropics [Halstead, 2000]. *A. aegypti* is the principal vector of dengue fever and dengue haemorrhagic fever. At present no any promising medicines are available for dengue. New way of reducing the incidence is to block disease transmission by either killing, preventing mosquitoes to bite human beings (by using repellents) or by causing larval mortality in a large scale at the breeding centres of the vectors. Many approaches have been developed to control mosquito, among which the current approach is based on synthetic insecticides. Even though they are effective, they created many problems like insecticide resistance [Lixin et al., 2006], pollution and toxic side effects on human beings [Liu et al., 2005]. In addition, they have also caused the problem of pesticide resistance in some mosquitoes [Su and Mulla, 1995].

Thus the use of natural products like phytochemicals and biopesticides, has recently been given much emphasis against the use of chemical fungicides and pesticides since they are eco-friendly. Many plant extracts or allelochemicals show a broad spectrum of activity against pests and such products have long been used as attractive alternatives to synthetic chemical pesticides for pest management because they pose little threat to the environment or to human health. The effects of botanical derivatives against mosquito have been reviewed [Sukumar et al., 1991].

Garcinia gummi-gutta var. *gummi-gutta* is a plant genus of the family Clusiaceae native to Asia, Australia, tropical and South Africa, and Polynesia. They are commonly known as Kodampuli or Malabar Tamarind. The Clusiaceae family is a rich source of secondary metabolites. Four major classes of compounds are found in the Clusiaceae family: xanthones, coumarins, biflavonoids, and benzophenones [Tharachand et al., 2013]. *Garcinia gummi-gutta* has been investigated extensively by many researchers in reference to its medicinal properties and various other cytotoxic effects [Nimbalet al., 1996b] Although many researchers have confirmed its antifungal [Duke et al., 1997], anticancer [Acuna et al., 2009], antimicrobial [Shara et al., 2004] and antiviral effects [Danziel

JM, 1937], making the plant a good source of medicine, only few researches have been done on its allelopathic aspect. So the purpose of the present study was to elucidate the allelopathic potential of *Garcinia gummi-gutta* leaf extracts on some common agricultural crops.

Materials and Methods

The leaves of *Garcinia gummi-gutta* were collected, chopped into small pieces, oven-dried and were coarsely powdered using in a table model grinder. The ground plant leaves (50 g) were dipped in methanol solvents and were extracted with 250 ml of methanol using a Soxhlet extractor for 72 h at a temperature not exceeding the boiling point of the solvent. The extracts were filtered using filter paper (Whatman No. 1) and then concentrated *in vacuo* at 40°C using a Rotary Evaporator. The extract was considered as stock solution. The extracts obtained were stored in a freezer at -80°C until further tests [Fuji et al., 2003].

About 25 ml of the crude extract containing 2.430 g residue was subjected to purification by means of Normal Phase Column chromatography using silica gel (60-120 mesh size) in successive elution with CHCl_3 -MeOH in the ratio 2:1 (30 ml each). The process of purification was repeated 2-3 times to increase the concentration. The fractions found to contain flavonoids (through phytochemical analysis) were pooled together (fraction 14-19), evacuated to dryness, thus obtained a concentration of 162.19 mg. Redissolved in 50 ml methanol. From this, a 5 ml containing 13.05 mg was taken for larvicidal test.

Treatment of larvae with leaf extract : For the assay, dry extracts of the fraction (13.05 mg in 5 ml methanol) isolated from methanolic extract of the leaves were dissolved in 10 ml dimethylsulphoxide (DMSO) to prepare graded series of concentrations. so as to solubilize the compound in water. From the stock solution, different concentrations ranging from 0.05, 1.0, 2, 3, 4 and 5 ml /lit were prepared. Control medium was also maintained to 100 ml by adding 95 ml tap water and 5 ml DMSO. Four replicates were maintained for each concentration.

Larvicidal bioassay was carried in 500 ml glass beakers containing 250 ml of water and 20 numbers of late III or early IV instar mosquito larvae for various concentrations. For the treatment of larvae with the leaf extract of *Garcinia gummi-gutta*, 100 ml of tap water was kept in a series of glass beak-

ers (of 200 ml capacity). Twenty numbers of 3rd or 4th instar larvae of *A. aegypti* were separately introduced into control and different concentrations of leaf extract. The mortality rate was evaluated 24 hours after the beginning of the experiment, verifying the number of dead larvae. After treatment, the larvae were considered dead if, at the end of 24 hrs, they showed no sign of swimming movements even after gentle touching with a glass rod, as described in the World Health Organization's technical report series [WHO,1981]. The number of larvae surviving at the end of 24 h was recorded and the per cent mortality values were calculated.

Results and Discussions

In the present investigation, the larvicidal potential of flavonoids isolated from methanolic extracts of leaves of the plant *Garcinia gummi-gutta* at varying concentrations against the larval stage of dengue vector *Aedes aegypti* has been tested. The data were recorded and mortality values were calculated. Results shows that the plant is possessing the activity at higher concentrations (inhibitory), whereas a negative larvicidal effect (stimulatory) was observed at lower concentrations. Of the seven concentrations used against *Aedes aegypti* larvae of 95.6%, has been observed with a concentration of 3mg/ml. A 50% larvicidal effect (LC50) was exhibited by a concentration of 2.5mg/ml. Lower concentrations did not exhibit any larvicidal effect. The activity increased with an increase in the concentration of the compound (Table.1). So we can suggest that flavonoid compound isolated from *G.gummi-gutta* is possessing the ability to kill mosquito larvae at higher concentrations.

Table.1.Larvicidal Effect of the Flavonoids isolated from *Garcinia gummi-gutta*

Concentration of methanolic extract of leaf of <i>G.gummi-gutta</i> (mg/ml)	No. of larva dead after 48 hr (out of 20 larva)		Percentage mortality (%)	
	Control	Test	Control	Test
0.25	0.5±0.58	0.5±0.58	2.5	2.5
0.5	0.5±0.58	0.5±0.58	2.5	2.5
1	0.5±0.58	0.5±0.58	2.5	2.5
1.5	0.5±0.58	0.5±0.58	2.5	2.5
2	0.5±0.58	4.75±0.96	2.5	23.75
2.5	0.5±0.58	10.75±0.96	2.5	53.75
3	0.5±0.58	19.12±0.96	2.5	95.6

Four replicates were done. Values are expressed as mean±SD.

Flavonoids which are the pharmacologically active constituents in many herbal plant medicines are known to possess cytotoxic effects. Larvicidal effect of the plant *G.gummi-gutta* has not been tested before. The studies conducted on larvicidal effect of *Azadirachta indica* (Meliaceae) against *Aedes aegypti* supports the idea [Dua et al., 1995]. Similar dose dependant effect of *Lantana camera* extracts on third instar larvae of *C. quinquefasciatus* showed larvicidal activity and cause maximum mortality rate at 3.0mg/ml concentration [Sathish Kumar and Maneemegalai, 2008]. Larvicidal activity of ethyl acetate, butanol, and petroleum ether extracts of five species of *Euphorbiaceae plants*, *Jatropha curcas*, *Pedilanthus tithymaloides*, *Phyllanthus amarus*, *Euphorbia hirta*, and *Euphorbia tirucalli*, were tested against the early fourth instar larvae of *Aedes aegypti* L. and *Culex quinquefasciatus* [Rahuman et al.,2008].

Plant extracts have offered many beneficial uses in applications ranging from pharmaceuticals to insecticides. The combined effect of phenolic acids such as caffeic acid, vanillic acid, anisic acid, p-anisic acid, chlorogenic acid and parahydroxy benzoic acid isolated from the leaf extract of *P. hysterocephus* revealed the larvicidal and pupicidal property on *A. aegypti* and *C. Quinquefasciatus* [Narasimhan and Keshava murthy,1984]. Parthenium isolated from the lant *Parthenium hysterocephus* is also reported as promising remedy against hepatic amoebiasis [Sharma and Bhutani,1988; Dominguez and Sierra,1970]. The botanical insecticides are generally safer, readily biodegradable non-toxic. But they are found active against the insect pest, lack toxicity to higher animals and they do not leave any phytotoxic residues in the environment. Thus the observation made in the present study have come as yet another evidence for the significant influence of the plant derived botanical pesticide like *G.gummi-gutta* in the control of mosquito.

From the above results, it has been observed that that the flavonoids isolated from the leaf extracts of the plant *G.gummi-gutta* are promising as larvicidal agent against *Aedes aegypti*. Further studies are needed to determine its mode of action, toxicity, stability, its impact on human health and non target organisms in mosquito feeding habitats when exploited as a larvicide etc.

REFERENCE

1. Becker N, Petraie D, Zgomba M, Boase C, Dahl C, Lane J, Kaiser A. 2003. Mosquitoes and their control. New York: Kluwer Academic Plenum Publisher; p. 1–16. | 2. Halstead SB. 2000. Global perspective on Dengue Research. *Dengue Bull.* 24: 77–82. | 3. Lixin S, Huiqin D, Chongxia G, Jin, Q, Jing S, Lei M. and Chang liang Z. 2006. Larvicidal activity of extracts of Ginkgo biloba Exocarp for three different stages of *Culex pipiens pallens*. *Journal of Medical Entomology*, 43(2): 258 - 261. | 4. Li H, Xu Q, Zhang L, Liu N. 2005. Chlorpyrifos resistance mosquito *Culex quinquefasciatus*. *J Med Entomol* 42:815–820 | 5. Su, T and Mulla MS.1998. Ovicidal activity of neem products(Azadirachtin) against *Culex tarsalis* and *Culex quinquefasciatus* (Diptera: Culicidae). *Journal of American Mosquito Control Association*, 14, 204–209. | 6. Sukumar K, Perich MJ and Boobar LR. 1991. using steam distillation against *Aedes* mosquito bite. Botanical derivatives in mosquito control a review. J. | In this present study the leaf and flower extracts of Am. Mosq. Control Assoc., 7: 210–237. | 7. Tharachand, Immanuel Selvaraj, Mythili Avadhani.2013. Medicinal Properties of Malabar Tamarind (*Garcinia Cambogia* (Gaertn.) DESR.). *Int. J. Pharm. Sci. Rev. Res.*, 19(2), n 20, 101–107. | 8. Ulyana Munoz Acuna . 2011. Phenolic Constituents from *Garcinia intermedia* and Related Species. ProQuest Dissertations and Theses. | 9. Duke SO, F.E. Dayan, A. Hernández, M.V. Duke, and H.K. Abbas. 1997. Natural products as leads for new herbicide modes of action. In Proceedings of The 1997 Brighton Crop Protection Conference—Weeds. Vol. 2, eds.; pp. 579–586 | 10. Acuna, Ulyana M.; Jancovski, Nikola; Kennelly, Edward J. 2009. Polyisoprenylated benzophenones from Clusiaceae: potential drugs and lead compounds. *Current Topics in Medicinal Chemistry*, Volume 9, Number 16, , pp. 1560–1580(21). | 11. Shara M, Ohia SE, Schmidt RE, Yasmin T, Zaedetto-Smith A, Kincaid A, Bagchi M, Chatterji A, Bagchi D, Stohs SJ. (2004). Physico-chemical properties of a novel (-)-hydroxycitric acid extract and its effect on body weight, selected organ weights, hepatic lipid peroxidation and DNA fragmentation, hematology and clinical chemistry, and histopathological changes over a period of 90 days. *Mol Cell Biochem* 260:171–186. | 12. Dalziel JM. 1937. The Useful Plants of West Tropical Africa. Crown Agents for the colonies, London. | 13. Fujii Y, Parvez SS, Parvez MM, Ohmae Yand Iida O. 2003. Screening of 239 medicinal plant species for allelopathic activity using sandwich method. *Weed Biol. Mang.* 3: 233–241. | 9. WHO. 1981. Instructions for determining the susceptibility/resistance of mosquito larvae to insecticides. WHO/VBC/81.807 Geneva: World Health Organization; p. 7. | 10. Dua VK, Nagpal BN, Sharma VP. 1995. Repellent action of neem cream against mosquitoes. *Indian J. Malariol.* 32: 47–55. | 11. Sathish Kumar M and Maneemegalai S. 2008. Evaluation of Larvicidal Effect of *Lantana* [camara Linn against Mosquito Species *Aedes aegypti* and *Culex quinquefasciatus*. *Ad. Bio. Res.*, 2: 39–43. | 12. Rahuman AA, Gopalakrishnan G, Venkatesan P, Geetha K. 2008. Larvicidal activity of some Euphorbiaceae plant extracts against *Aedes aegypti* and *Culex quinquefasciatus* (Diptera: Culicidae). *Parasitol Res. Apr*; 102(5):867–73. | 13. Narasimhan TR and Keshava murthy BS. 1984. Characterization of a toxin from *Parthenium hysterophorus* and its mode of excretion in animals. *Journal of Bioscience.* 6:729–738. | 14. Sharma GL and Bhutani KK. 1988. Plant based antiamebic drugs. Part II. Amoebicidal activity of parthenin isolated from *Parthenium hysterophorus*. *Planta Medica.* 54:20–22. | 15. Dominguez, XA and Sierra A. 1970. Isolation of a new diterpene alcohol and parthenin from *Parthenium hysterophorus*. *Planta Medica* 18:275–277. |