



Extraction of Coelomic Fluid from the Earthworm *Polypheretima elongata* for the Antibacterial Activities

KEYWORDS

Earthworm, Coelomic fluid, Antibacterial activity

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ABSTRACT It is proved that earthworm coelomic fluid is having antibacterial activity. It was our interest to know the antibacterial activity of the coelomic fluid of *Polypheretima elongata* against pathogenic bacteria like *Escherichia coli*, *Staphylococcus aureus*, *Pseudomonas*, *Proteus* and *Bacillus*. *Polypheretima elongata* is a local species of earthworm present in Gulbarga. The antibacterial activity of the coelomic fluid of the earthworm was tested in comparison with the commercial antibiotics available in the market. The antibacterial activity was determined by the zone of inhibition by coelomic fluid of earthworm *Polypheretima elongata* against bacterial strains. The zone of inhibition varies from species to species, it is species specific particular bacteria. The best inhibitory effect of coelomic fluid of *Polypheretima elongata* on the growth of *Staphylococcus aureus* is 10mm, *Escherichia coli* and *Proteus* is 9mm, *Bacillus* is 7mm and *Pseudomonas* is 5mm.

I) Introduction:

Invertebrates exhibit different immune mechanisms against environmental pathogens. In earthworms, coelomocytes (leucocytes) located in coelomic cavity are responsible for innate cellular immune functions such as phagocytosis and encapsulation against parasites and pathogens. The coelomocyte population has been divided into different subpopulations. The coelomocytes possess immuno-defense related biological functions. The effector cells participate mainly in cellular mechanism but chloragocytes and the granulocytes of coelomocytes may produce humoral factors which may mediate the cellular and humoral responses as well. Microbial killing results from the combined action of the phagocytic process with humoral immune factors such as agglutinin (viz., lactin), lysosomal enzymes (viz., acid phosphatases) and various cytotoxic and antimicrobial molecules.

Invertebrates have developed innate immune mechanism that detects pathogens by recognizing conserved molecular patterns. Molecules responsible for the recognition of foreign material have been named as pattern recognition proteins (PRPs) (Medzhitov and Janeway, 1997) because the host's primitive effector cells would recognize molecular pattern rather than particular structure of the invading microorganisms. Examples of pathogen-associated molecules, which are not found in the multicellular organisms, are lipopolysaccharides (LPS) or peptidoglycans from bacterial cell walls β -1, 3-glucan of fungal cell walls and double stranded RNA of viruses.

Immune mechanism of earthworms includes both cellular and humoral components. Earthworm coelomocytes respond to the presence of pathogens by phagocytosis, encapsulation/brown body formation and N K cells activity (Cooper et al., 2001). Humoral components include lectin, antimicrobial peptides, pore forming proteins, phenoloxidases and proteases. They include sequestration of antigenic material by agglutination, cytotoxicity and antibacterial activity. Cytolysin of *Eisenia fetida* coelomic fluid, named eiseniapore, was found to cause pore like structure at the target membranes. Formation of pore proteins is the result of oligomerization of eiseniapore monomers (Lange et al., 1999).

There are many investigations on the presence of antibacterial substances in coelomic fluid of earthworms since bacterolytic molecules were identified as lysozyme like molecules, active only against gram positive bacteria (Jolles and Zuili, 1960; Scherbert and Messner, 1997; Lassegues, 1986). The

coelomic fluid of earthworm, *Eisenia fetida* (Oligochaeta: Lumbricidae) was demonstrated to possess an antibacterial activity directed against earthworm pathogenic bacteria –namely: Gram negative *Aeromonas hydrophila* and Gram-positive *Bacillus megaterium* by Valembos et al. (1992).

Our present study is aimed to know the antibacterial activity of coelomic fluid (crude form) of the tropical earthworms *Polypheretima elongata* on pathogenic bacteria like *Escherichia coli*, *Staphylococcus aureus*, *Pseudomonas*, *Proteus*, and *Bacillus*. This antibacterial activity of the coelomic is fluid compared with that of commercially available antibiotics like Streptomycin, Norfloxacin, Ciprofloxacin, Gentamicin and Amoxicillin.

II) Materials and Methods:

a) Bactericidal studies

The bacterial strains used for determining antibacterial activity are *Escherichia coli*, *Staphylococcus aureus*, *Pseudomonas*, *Proteus* and *Bacillus* which were obtained from Department of Microbiology, Gulbarga University, Gulbarga. The nutrient agar media was prepared in the laboratory by adding agar to the nutrient broth (Hi-Media make). Whatman no. 1 filter paper was used to prepare the discs. Commercially available antibiotic discs (Streptomycin, Norfloxacin, Ciprofloxacin, Gentamicin and Amoxicillin) were purchased from Hi-media. Sterile swabs were purchased from local medical distributors. Nutrient agar media was purchased from Hi-media.

b) Method of Extraction of Coelomic Fluid for Antibacterial Activities :

Earthworms were washed, kept on soft paper to absorb surface body moisture and then transferred to Petri dish, excited with 6 volt electrical stimulation, which induced them to extrude coelomic fluid through epidermal dorsal pores. The fluid extracted was collected in the small tubes and used for antibacterial tests. Sterilized blotting paper (about 6 mm diameter), free from any antibacterial activity, were impregnated with 20 μ l of the fraction to be tested and placed on agar dishes inoculated with bacterial strain.

c) Disk method:

The target bacterial species were separately cultured in the nutrient broths to yield 10^7 cfu/ml and 18-24 hr cultures were separately spread plated on nutrient agar plates. Then, the 6 mm paper disks impregnated with 20 μ l coelomic fluid fraction and incubated upright at 37°C in an incubator for 24 h. The bactericidal activity of the fractions was measured in

terms of mm zone of inhibition. Antibacterial activity of the coelomic fluid of the *Polypheretima elongata*, was compared with the commercial antibiotics available in the market viz., Streptomycin, Norfloxacin, Ciprofloxacin, Gentamicin and Amoxillin.

III)Results:

The influence of coelomic fluid of *Polypheretima elongata* on the *in vitro* growth of bacterial cultures was evaluated against pathogenic bacteria such as *Escherichia coli*, *Proteus*, *Pseudomonas*, *Staphylococcus aureus* and *Bacillus*. The best inhibitory effect of coelomic fluid of *Polypheretima elongata* on the growth of *Escherichia coli*, *Proteus* and *Staphylococcus aureus* was seen to be 9 mm to 10 mm zone of inhibition. The inhibitory effect is comparatively less against *Pseudomonas*, and *Bacillus*, exhibiting 5 mm and 7 mm inhibitory zones in *Pseudomonas* and *Bacillus* respectively (Table – 1, Plate 1 to 5).

Table – 1: Zone of inhibition (antibacterial activity) obtained by the coelomic fluid of *Polypheretima elongata* and commercial antibiotics Amoxillin, Streptomycin, Norfloxacin, Ciprofloxacin and Gentamicin against the growth of pathogenic bacteria like *Escherichia coli*, *Staphylococcus aureus*, *Pseudomonas*, *Proteus* and *Bacillus*.

Bacteria used	Zone of inhibition (mm) <i>P. elongata</i>	Amoxillin (mm)	Streptomycin (mm)	Norfloxacin (mm)	Ciprofloxacin (mm)	Gentamycin (mm)
<i>Escherichia coli</i>	9	12	24	2	25	15
<i>Proteus</i>	9	18	2	22	23	2
<i>Pseudomonas</i>	5	–	24	32	38	18
<i>Staphylococcus aureus</i>	10	14	22	25	34	24
<i>Bacillus</i>	07	–	3	3	33	19



Plate – 1: Zone of suppression impregnated with coelomic fluid of *Polypheretima elongata* on *Escherichia coli*.

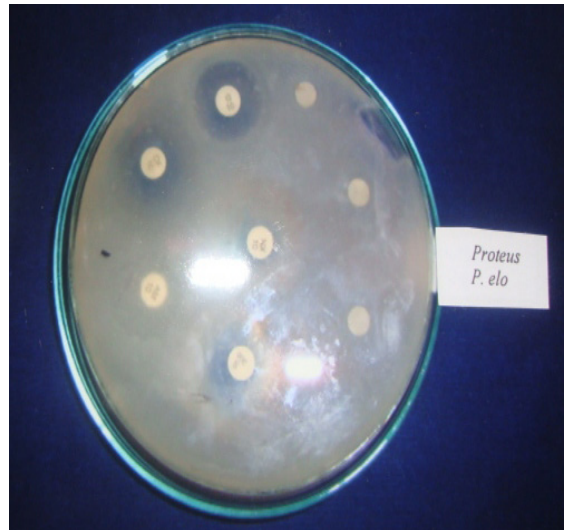


Plate – 2: Zone of suppression impregnated with coelomic fluid of *Polypheretima elongata* on *Proteus*.

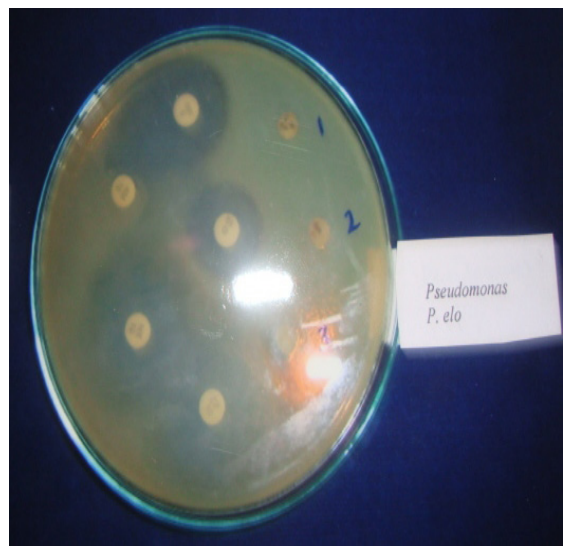


Plate – 3: Zone of suppression impregnated with coelomic fluid of *Polypheretima elongata* on *Pseudomonas*.

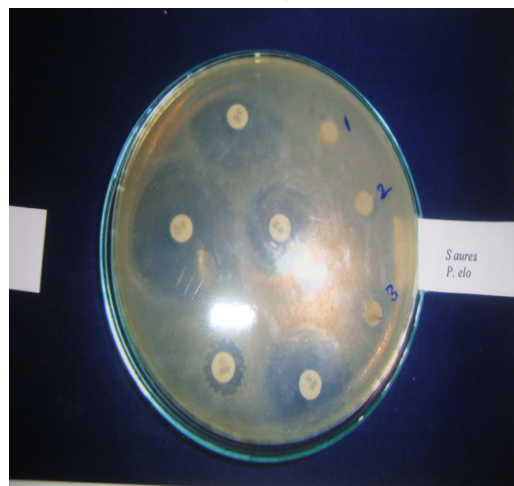


Plate – 4: Zone of suppression impregnated with coelomic fluid of *Polypheretima elongata* on *Staphylococcus aureus*.

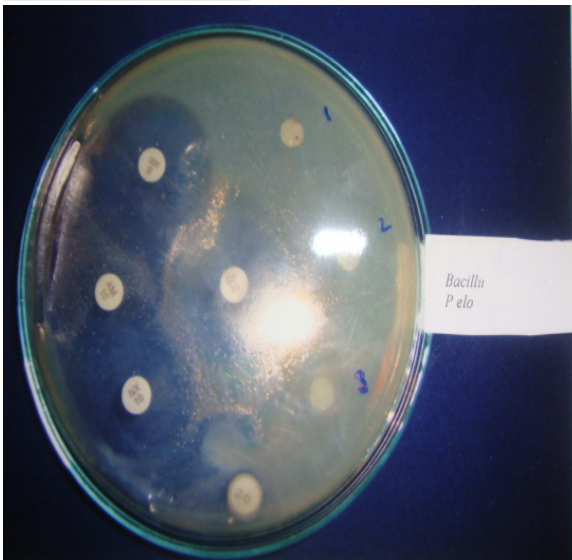


Plate – 5: Zone of suppression impregnated with coelomic fluid of *Polypheretima elongata* on *Bacillus*.

IV) Discussion:

Preparations made out of materials of animal origin (placenta, snake and bee poison, dags, sea and river hydrocoels) were used extensively in folk and modern medicine for treating many human diseases. These preparations are also gaining importance now-a-days, though some of them are not used in modern medicine because of resource deterioration. Renewable natural medical materials of animal origin and pharmaceutical preparations on their basis as well as biologically active food supplements are the future. This area of human activity and economics should be developed and improved in every possible way. In many South-East Asia countries earthworms have already been used for nearly 2300 years to cure different human diseases. In Vietnam a dry earthworm powder produced according to different techniques that guarantee its purity is the compulsory ingredient of pharmaceuticals or "magic medicines that save life with in 60 minutes". These pharmaceuticals are often used to treat various organ dysfunctions-the result of bacterial and viral infections.

Darwin too commented that tissue fluid of earthworm can dissolve fibrin, many Japanese researchers have extracted fibrin dissolving enzymes from *Lumbricus rubellus*.

Our results show that coelomic fluid of earthworm *Polypheretima elongata* applied in the certain concentrations could decrease growth of pathogenic bacteria.

In clinical experience, the inflection of skin wounds often occurs by Staphylococci causing a decrease in the proliferation of epithelial cells and fibroblasts in the wounds (Chang *et al.*, 1997). Maja *et al.* (2005) have reported G-90; a glycolipoproteins mixture obtained from tissues homogenate of earthworm *Eisenia fetida* and has shown the antibacterial activity *in vivo* and *in vitro* in different concentrations of growth on non pathogenic and facultative pathogenic bacteria. The bacteriostatic effect of the G-90 was 21% stronger for facultative pathogenic bacteria than that observed for non-pathogenic bacteria.

Upon the inoculation of bacteria into the coelomic cavity of earthworm, the coelomocytes initiate the process of connecting with each other by their adhesive structures around the bacteria and form so called brown bodies (Valembois *et al.*, 1992; Cooper *et al.*, 1999). At the same time the coelomocytes intensively synthesize and secrete proteins that adhere to the bacteria, forming aggregations and may inhibit their further proliferation. One of these proteins is agglutination of 56 kDa molecular mass, which attaches to the lectin like monosaccharide of the cellular membrane of the bacteria.

Rejnek (1991), Tuckova (1991) and Valembois *et al.* (1993) demonstrated significant antibacterial activity of the earthworms and that they were assisted besides coelomocytes by the chloragocytes, i.e., cell from the intestinal tract of earthworms, in resistance mechanisms. The chloragocytes secrete two proteins with a molecular mass of 40 and 45 kDa, which share 35% similarities with immunoglobulins. They adhere to the bacteria making them suitable for phagocytosis by coelomocytes in coelomic fluid (Lassegues *et al.*, 1997). The chloragocytes synthesize and secrete the protein lysozyme (33 kDa), which binds specially to phospholipids of the cell membrane and causes cytolysis (Kobayashi *et al.*, 2000; Ohit *et al.*, 2000).

With the knowledge, it is difficult to define which molecule of coelomic fluid of earthworms is responsible for its antibacterial activity. Theoretically such activities could be ascribed to some of the following molecular masses: 33, 40, 42, 45 and 60 kDa, which are detected by SDS-PAGE in G-90 (Hrzenjak *et al.*, 1992). By immunochemical analyses these proteins are shown to be belonging to the immunoglobulin super family (Popovic *et al.*, 1998).

From the results of our study it is clear that antibacterial activity is exhibited at different levels by coelomic fluids of *Polypheretima elongata*. Coelomic fluid of *Polypheretima elongata* is exhibit highest antibacterial activity against *Escherichia coli* and *Staphylococcus aureus*.

These observations are made based on zone of inhibition *in vitro*. However, further investigations on the presence of antibacterial molecules in the coelomic fluid and/or coelomocytes are needed which may help to explore the medicinal value of coelomic fluids of local species of earthworms.

V) Summary and Conclusion

Earthworms are known for their medicinal value since ancient times all over the world. Earthworms have blood and the coelomic fluid that contains both haemocytes and coelomocytes in their fluids. Coelomic fluids as well as blood of the earthworm have coelomocytes and haemocytes which play an important role in fighting against pathogens. This quality of earthworms made us to find out the immunological and antibacterial activity. To know the antibacterial activity of the coelomic fluid of the earthworms *Polypheretima elongata* against pathogenic bacteria like *E. coli*, *Proteus*, *Pseudomonas*, *S. aureus* and *Bacillus*. Results reveal that crude form of coelomic fluids of all the species of earthworm exhibited antibacterial activity. *Polypheretima elongata* exhibit highest antibacterial activity against *E. coli* and *S. aureus*. These results are based on the zone of inhibition *in vitro* growth of bacterial cultures. However, further investigations on the presence of antibacterial molecules in coelomic fluid of earthworms are necessary.

REFERENCE

- Chang, T.H., Patel, M., Watford, A., Freundlich, L., Steinberg, J.J. and Levenson, S.M., 1997. Single local instillation of Nonviable *Staphylococcus aureus* or its peptidoglycan ameliorates, glucocorticoid induced impaired wound healing. *Rep. Rweg.*, 5:184-190. | Cooper, E.L., Cossarizza, A., Kauschke, E. and Franceschi, C., 1999. Cell adhesion and the immune system: A case study using earthworm. *Microse. Res. Tech.*, 44: 237-253. | Cooper, E.L., Kauschke, E., Cossarizza, A., 2001. Annelid humoral immunity: Cell lysis in earthworms. *Adv. Exp. Med. Biol.*, 484: 169-183. | Hrzenjak, T., Hrzenjak, M., Kasuba, V., Efenbergez-Marinculic, P. and Levanat, S., 1992. A new source of biologically active compounds earthworms tissue (*Eisenia foetida*, *Lumbricus rubellus*). *Comp. Biochem. Physiol.*, 102A (3): 441-447. | Jolles, P. and Zuili, S., 1960. Purification et etude compare de nouveaux lysozymes extraits du poumon de poule et de *Nephtys hombergi*. *Biochem. Biophys. Acta*, 39: 212-217. | Kobayashi, H., Sekizawa, Y., Aizu, M. and Umeda, M., 2000. Lethal and non lethal responses of spermatozoa from a wide variety of vertebrates and invertebrates to lysenin, a protein from the coelomic fluid of the earthworm *Eisenia foetida*. *J. Exp. Zool.*, 286: 538-549. | Lange, S., Kauschke, E., Mohrig, W. and Cooper, E.L., 1999. Biochemical characteristics of Eiseoiapore, a pore forming protein in the coelomic fluid of earthworms. *Eur. J. Biochem.*, 262: 547-556. | Lassegues, M., 1986. Etude des activites antibacteriennes humorales et cellulaires du lombricien *Eisenia foetida andrei*. These doctorates - Sciences Universite de Bordeaux. | Lassegues, M., Milochau, A., Doingnon, F., Du Pasquier, L. and Valembos, P., 1997. Sequence and expression of an *Eisenia foetida* derived cDNA clone that encodes the 40-kDa fetidin antibacterial protein. *Eur. J. Biochem.*, 246: 756-762. | Maja Popuic, Mira Grdisa and Terezija Mihaela Hrzenjak, 2005. Glycolipo protein G-90 obtained from the earthworm *Eisenia foetida* exerts antibacterial activity. *Verterinarska Arhiv.*, 75(2):119-128. | Medzhitov, R. and Janeway, C.A., 1997. Innate immunity: the virtues of a non clonal system of recognition. *Cell*, 91: 295-298. | Ohita, N., Shioda, S., Sekizawa, Y., Nakai, Y. and Kobayashi, H., 2000. Sites of expression of mRNA for lysenin, a protein isolated from the coelomic fluid of the earthworm *Eisenia foetida*. *Cell Tissues Res.*, 302: 263-270. | Popovic, M., Hrzenjak, T., Grdisa, M. and Vukovic, S., 1998. Adhesins of immunoglobulin like super family from earthworm *Eisenia foetida*. *Gen. Pharmacol.*, 30: 795-800. | Rejnek, J., Tuckova, J., Zikan, M. and Tomana, 1991. The interaction of a protein from the coelomic fluid of earthworms with *Staphylococcal* protein A. *Develop. Comp. Immunol.*, 15: 269-277. | Scherbert, I. and Messner, B., 1997. Untersuchungen über das vor kommenvone Lysozym bei anneliden, *Zool. Jahrb. Physiol. Biol.*, 76: 36-50. | Tuckova, L., Rejnek, J., Bilej, M. and Pospisil, R., 1991. Characterization of antigen binding protein in earthworms *Lumbricus terrestris* and *Eisenia foetida*. *Develop. Comp. Immunol.*, 15: 263-268. | Valembos, P., Lassegues, M. and Roch, P., 1992. Formation of brown bodies in the coelomic cavity of the earthworm *Eisenia foetida Andrei* and attendant changes in shape and adhesive capacity of constitutive cells. *Develop Comp. Immunol.*, 16: 95-101. | Valembos, P., Lassegues, M., Hirigoyenberry, F. and Seymour, J., 1993. Clearance and breakdown of pathogenic bacteria injected into the body cavity of the earthworm *Eisenia foetida Andrei*. *Comp. Biochem*