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Comparison of Otolith, Scale and Vertebrae for Age Estimation in Freshwater Exotic Fish Oreochromis mossambicus

KEYWORDS Comparison, age, longevity, exotic fish, Oreochromis mossambicus SWAPNALI B. LAD SURESH M. KUMBAR ABHIJIT B. GHADAGE Department of Zoology, Arts, Department of Zoology, Arts, Commerce and Science College Commerce and Science College Palus, Dist. Sangli, Maharashtra, Palus, Dist. Sangli, Maharashtra,

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ABSTRACT Otolith, scale and vertebrae sections of freshwater exotic fish O. mossambicus (N = 133) were compared to ascertain the best aging material. Different body sized fishes revealed the presence of 1 - 7 growth marks. Agreement between otolith and scale ages was 96.96% and otolith and vertebrae ages were 88.36% in 33 fishes used for comparative studies. Vertebrae ring counts (VRC) were under estimated the age in (13.6%) older individuals compared to otolith and scale ring counts (SRC). The results revealed that otoliths and scales are reliable calcified materials for estimating age compared to vertebrae in O. mossambicus. There was a highly positive correlation between body length (BL) and body weight (BW). Further, otoliths ring count (ORC), otoliths weight (OW) and otoliths diameter (OD) showed positive correlation with BL and BW. This fish may live for a maximum of 7 years in the natural population.

INTRODUCTION

Freshwater exotic fish O. mossambicus was noticed in the fish catch, probably on account of accidental entry with Indian major carp seed in 1990s. This fish population has been continuous increasing in the total number of caught in Indian major Rivers (Jain and Gupta, 1994) which cause of worry for the fishery biologists (Anon, 1995; Biju Kumar, 2000). Moreover, as per 2006 Global Invasive Species Database O. mossambicus is listed being as in the top 100 invasive alien species on the planet (Canonico et al., 2005). Invasive populations are now causing environmental and ecological problems in many countries including India (Canonico et al., 2005). O. mossambicus is widely distributed and now forms a part of fish fauna in the Godavari, Krishna, Cauvery, Yamuna and Ganga Rivers (Lakra et al., 2008).

Growth and age studies provide important demographic parameters to analyze and asses fish populations (Maceina & Sammons, 2006). Ageing of fishes from tropical regions have been reported through annual increments in calcified structures such as, scales (Werder and Soares, 1985; Mayekiso & Hecht, 1988; Sudarshan & Kulkarni, 2013), dorsal and pectoral spines (Bio & Ikusemiju, 1981; Pantulu, 1961), vertebral centra (Brown & Gruber, 1988; Bahuguna, 2013), and oto-liths (Fowler & Doherty, 1992; David & Pancharatna, 2003). Most of the earlier age estimation studies of Indian fishes have been concentrated on scales (Singh & Sharma, 1998; Dua & Kumar, 2006; Kanwal & Pathani, 2011; Ujjania et al., 2013) due to the easiest to collect, process and avoids sacrificing the specimens. Recent reviews have suspected on scale aging due to difficulties in reading annuli, low precision (Lowerre-Barbieri et al., 1994), and that scale ages may become inaccurate when growth becomes asymptotic (Beamish & McFarlane, 1987; Shepherd, 1988). Scales have generally been found to underestimate ages relative to other structures, especially for older individuals and in slow-growing populations (Campbell & Babaluk, 1979; Mills & Beamish, 1980; Erickson, 1983; Kocovsky and Carline, 2000). Since scale growth is assumed to be proportional to body growth (Whitney & Carlander, 1956; Hile, 1970; Bagenal, 1974; and Erickson, 1983), annuli become crowded on the scale edges in slow-growing populations and in older fish, making scale interpretation difficult. To overcome from these difficulties, comparison of various calcified materials have been performed in many species, including yellow perch Perca flavescens (Niewinski & Ferreri, 1999), river carp suckers Carpiodes carpio (Braaten et al., 1999), and white suckers Catostomus

commersonii (Scidmore & Glass, 1953; Ovchynnyk, 1969; Quinn & Ross, 1982; Sylvester, 2006). Although, some studies are also available on age and growth of O. mossambicus by using scales (Ujjania et al., 2013) and otoliths (Booth & Merron, 1996; Panfili & Tomas, 2001). There were no comparative studies for selection of reliable calcified material for aging in this species. Present investigation aims to select the most reliable calcified material for estimating the age of exotic freshwater fish O. mossambicus inhabiting Krishna River, Southern India.

MATERIALS AND METHODS

Different body sized freshwater fishes (O. mossambicus, N = 133) were collected from the Krishna River, Sangli District, Maharashtra, India with the help of local fisherman and purchased from local market in 2012 - 2013. Total body length (BL) of each fish was measured from the tip of snout to tip of the caudal fin (in cm). Body weight (BW) was recorded by using single pan balance nearest to 0.01gm, simultaneously lateral line scales, otoliths and vertebrae of each specimen was collected and preserved in different vials for further studies.

Scale study:

Lateral line scales of each specimen was collected with the help of forceps, cleaned in water by rubbing gently with fingers, fixed in 10% formalin solution for 24 hours and washed in water for 2 hours. Cleaned scale was kept in between two clean slides and slides were tied with rubber band on either side and observed under binocular microscope (Magnus MSZ-BI) for enumerating the ring counts present on the scales and photographed by digital camera (ABBOT DEC-2000).

Otolith Studies:

Otoliths were collected by taking an incision on the dorsal side of the head, to expose the brain on either side of which the otic capsules are located. The sagittal otoliths were removed from the otic capsules by opening the otic bulla. Both sagittae were retrieved intact from each specimen, washed in water and cleaned from all extraneous tissue. Then, each otolith was weighed to the nearest 0.001 mg the diameter was measured to the nearest 0.01 mm using a standard micro screw gage (Newman et al., 2000). Otoliths were then immersed in 50% glycerol and observed under binocular microscope (Magnus MLXB). Growth rings were clearly visible as alternate opaque and translucent zones that were enumerated.

Vertebrae Section:

Different body sized 33 (BL 6.4 - 35.5 mm) fishes were used for vertebrae sections. Central 5th and 10th vertebrae of each fish were excised, cleaned and fixed in 10% formaldehyde solution for 24 hours, then washed in water for 1 hour and decalcified with 10% nitric acid (duration of decalcification depends on size of vertebrae). Decalcified vertebrae were washed under running tap water for 24 hours to remove the traces of formaldehyde and nitric acid and preserved in 70% alcohol. These vertebrae were embedded in paraffin wax and sectioned (10µ thickness) by using a rotary microtome (Model GE-70). Mid-diaphyseal sections were stained with Harris haematoxyline and observed under compound microscope for enumeration of growth marks (Buckmeier et al., 2002).

Comparison of otoliths, scales and vertebrae sections of 33 fishes were undertaken for the selection of reliable calcified material for accurate assessment of age and longevity. For each fish, growth marks were counted on the aging structures independently by two readers without prior information of body length and body weight. Then percent agreement was calculated between otolith ring counts with scales and vertebrae ring counts.

Statistical analysis:

The relationship if any, between BL and BW, BL and OW, BL and OD, BW and OW, BW and OD was assessed by plotting scatter diagrams and using regression analysis (Zar, 1984). The relationship between ORC and BL, ORC and BW, ORC and OW and the ORC and OD was determined by calculating the correlation coefficient 'r' by Karl Pearson's method (Zar, 1984).

RESULTS

Monthly variation in ambient temperature and rainfall in the year 2012 - 2013 is shown in figure 1. Although the mean monthly temperature varied between 21 - 30°C and that of rainfall from 0 to 121 mm (Fig. 1)

The otoliths, scales and vertebrae of O. mossambicus showed annual rings, each ring composed of a faint broader growth zone and a dark opaque line (Lines of arrested growth) the LAG (Plate 1A-F). One to seven growth marks were observed in the otoliths, scales, and vertebrae of different body sized fishes (N = 133). Out of 133 fishes, 21 (15.79%) fishes with mean body length (13.9 cm) have not exhibited any LAG (Table 1, Plate 1A, C, E). Thirty fishes (22.56%) with mean body length (16.28 cm) showed 1 LAG (Table 1), 38 fishes (28.57%) with mean body length (16.71 cm) exhibited 2 LAGs, 29 fishes (21.80 %) with mean body length (15.57 cm) showed 3 LAGs, 8 fishes (6.02%) with mean body length (17.21cm) exhibited 4 LAGs, 4 fishes (3.00 %) with mean body length (18.95 cm) showed 5 LAGs and 3 fishes (2.26%) with mean body length (23.66 cm) exhibited 6 LAGs (Table 1 & Plate 1B, D, F). It was generally observed that in older fish growth marks appeared sharp at the periphery of the otoliths, scales, and vertebrae sections (Plate 1B, D, F).

Comparison of otolith ring count with scale and vertebrae ring count in 33 large body sized fishes were done, the number of rings remained identical in otoliths, scales and vertebrae sections of 26 (78.78%) fishes. In one (3.03%) fish, the scales showed short on one growth ring compared to otolith in 3rd year individual. In 4 (12.12%) fishes, the vertebrae showed one growth ring lesser compared to otolith rings in 3rd year age groups and 2 (6.06%) fishes were showed short on two growth rings in 4th and 7th year age group individuals.

Of the 133 fishes sampled, the body length (BL) ranged from 6.0 - 35.5 cm with an average of 20.75 cm and body weight (BW) varied between 5 and 760 g with a mean of 382.5 g (Table 1). There was a high degree of positive correlation (r = 0.736) between BL and BW (Fig. 2). The scatter diagrams indicated that BL correlated highly positively with OD (r =0.841; Fig. 3) and OW (r = 0.647; Fig. 4). Further, BW also showed positive correlation with OD (r = 0.628; Fig. 5) and OW (r = 0.759; Fig. 6). While, the ORC showed a lesser correlation with BW (r = 0.447), BL (r = 0.220), OW (r = 0.273) and OD (r = 0.144).

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DISCUSSION

Age and growth studies of fish provide information of rate of survival, growth, mortality, age composition, and reproduction rate (Polat et al., 2001; Campana, 2001). In the present study, one to seven broader light growth zones and one to six opaque sharp growth-arrested zones were found in fishes of varying body size (Table 1 and Plate 1). The comparative studies of otolith, scale, and vertebrae of freshwater exotic fish O. mossambicus revealed the presence of growth marks similar to that reported for other tropical species namely, *Sillago japonica* (Sulistiono *et al.*, 1999); Indian whiting, *S.* indica (David & Pancharatna, 2003); Catla catla (Ujjania, 2012); and Notopterus notopterus (Sudarshan & Kulkarni, 2013). The meteorological data indicate that, the annual temperature variation of Sangli is within 9°C, and the rainfall exhibited considerable annual variation (Fig. 1), further, during monsoon season there were increased temperature and maximum rainfall, consequently, cyclic pattern of growth in hard calcified material is common in fishes which is generally attributed to enforced seasonal feeding activity owing to marked variations in surrounding climatic factors, e.g. the extreme seasonal shifts in temperature of temperate areas, and the typical wet and dry conditions of the humidity cycle in the tropics (Johal & Tandon, 1992; Seshappa, 1999; Campana, 2001). Most of the freshwater fishes inhabiting Southern India exhibits a clear seasonality in body growth, breeding activities and gametogenesis (Saidapur et al., 1989). Even though, O. mossambicus is spawning in July - August (rainy season) when spawning of most of the Indian major carps (C. catla, L. rohita, and C. mrigala) occurred (Ganie, 2013). Similar observations on the other fishes have reported (Seshappa, 1958; 1972; Alam & Pathak, 2010). Moreover, experimental studies have been confirmed that the formation of growth marks are annual in many species including, Cirrhinus mrigala (Hanumantha Rao, 1974), O. niloticus (Abdel-Hadi et al., 2000); Clarias gariepinus (Wartenberg et al., 2011), Puntius conchonius (Bahuguna, 2013); and Labeo rohita (our personal observation). Therefore, number of growth marks present in scale, otolith and vertebrae section is directly depict the age of individual fish.

Among 133 fish samples, 21 fishes showed no LAG in its calcified materials indicating that the fishes may be in their first year of growth. One to third year age group individuals are dominant with varied body length (6.0 - 35.2 cm). Further, the positive correlation between body length and growth mark of these fishes indicate that larger individuals have experienced a greater number of growth marks and therefore may be older. Larger fishes exhibited increased number of growth marks, thus there was a positive correlation between OW, OD with BL and BW, whether the body length and weight influences age needs further elucidation.

Comparison of the three ageing structures within the current study revealed that otolith is the most precise material for age estimation in O. mossambicus due to clear and distinctness in growth marks. The suitability of otolith for age estimation is also supported by several workers (Seshappa, 1999; Campana, 2001) the fact that otolith do not show reabsorption and their growth is acellular rather than by calcification (Secor et al., 1995) and also because otoliths are reported to be metabolically inert and thus do not reflect physiological changes that may occur throughout the life of fish (Phelps et al., 2007). Otoliths continue to grow and form annuli even as body growth slows and asymptotic length is reached, and annuli reasbsorption does not appear to occur during periods of food limitation or stress (Devries & Frie, 1996). Several workers were reported that otoliths to be the most reliable ageing structure in a number of temperate as well as tropical species such as, Chelidonichthys kumu (Staples, 1971),

Capoeta capoeta umbla (Ekingen & Polat, 1987), Trachurus trachurus (Polat & Kukul, 1990), Pylodictis olivaris (Nash & Irwin, 1999), Indian whiting (David & Pancharatna, 2003) and Ictalurus punctatus (Buckmeier et al., 2002; Colombo et al., 2010). In the comparative studies, out of 33 fishes 32 fishes the number of growth marks was identical in scales and otoliths indicating that scale aging is reliable in > 97% of fishes. In other hand among 33 fishes 4 fishes one number of growth marks were short and another 2 fishes two growth marks were lesser in vertebrae sections compared to otolith ring counts in older individuals. Result indicating that vertebrae ring count is not that much reliable in this fish, this might be due to overcrowding of growth marks at the periphery of vertebrae sections, which was difficult to differentiate and count.

Our study found variation in the otolith, scale, and vertebrae ages of *O. mossambicus.* In this fish, vertebrae section underestimated the ages when compared with the otolith and scale ages. However, otolith and scale ring counts are more distinct and easy to enumerate, even in the older fish, than the vertebrae ring count. Therefore, we assume that the otolith and scale provide a reliable source for aging in *O. mossambicus* inhabiting Southern India.

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Table 1 Showing number of fishes, percentage, mean body weight, length, otolith weight, diameter and number of lines of arrested growths (LAGs) and age frequencies of freshwater exotic fish *O. mossambicus* (N = 133).

Number of Fishes	Percentage (%)	Body Weight (gm)	Body Length (cm)	Otolith Weight (mg)	Otolith Diameter (mm)	No. of LAGs	Age Frequency
21	15.79	83.57	13.9	21.11	0.276	0	1 st Year
30	22.56	108.9	16.28	27.53	0.350	1	2 nd year
38	28.57	151.94	16.71	39.35	0.366	2	3 rd year
29	21.80	179.20	15.57	39.87	0.344	3	4 th year
08	6.02	314.62	17.21	64.00	0.323	4	5 th year
04	3.00	249.25	18.95	70.67	0.417	5	6 th year
03	2.26	394.66	23.66	39.8	0.421	6	7 th year



Figure 1: Figure shows monthly variation in mean temperature and rainfall of Sangli District from January 2012 to December 2013.

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Figure 2: Relationship between Body Length (BL) and Body Weight (BW) in O. mossambicus.



Figure 3: Relationship between Body Length (BL) and Otolith diameter (OD) in O. mossambicus.



Figure 4: Relationship between Body Length (BL) and Otolith weight (OW) in *O. mossambicus.*



Figure 5: Relationship between Body Weight (BW) and Otolith diameter (OD) in *O. mossambicus.*

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Figure 6: Relationship between Body Weight (BW) and Otolith weight (OW) in *O. mossambicus.*





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A - Scale with no LAG, B - Scale with 6 LAGs C - Otolith with no LAG, D - Otolith with 6 LAGs E - Vertebra section with no LAG, F - Vertebra Section with 6 LAGs in *O. Mossambicus* Mc - Marrow Cavity, Pbm - Periosteal Bone Margin Arrow - LAG

F

REFERENCE

Abdel-Hadi, Y. M., Craig, J. F., Babaluk, J. A. and Wassle, R. 2000. Oxytetracycline marking studies of Tilapia, Oreochromis niloticus. N. Am. J. Fish. Manage. 7: 138 – 141. || Alam, M. and Pathak, R. K. 2010. Assessment of fecundity and gonadosomatic index of commercially important Fish Labeo rohita from Ramganga River. International Journal Pharma Bioscience.1 (3): 1-6. | Anon. 1995. Afriki Machhali Phailaw Se Jaisamand Jheel Ki Desi Machhalion Par Sankat. Rajasthan Patrika. 11. 12.1995. | Bagenal, T. B. and F.W. Tesch, 1974. Age and Growth in Methods for Assessment of Fish Production in Fresh Waters. In: Bagenal, T.B., (Ed.), 3rd Edn., Blackwell Scientific Publication, Oxford, pp: 101-136. || Bahuguna, P. 2013. Age determination and growth rate of freshwater fish Puntius conchonius (Ham.-Buch) by a use of trunk vertebrae. Periodic Research. 2: 46-51. || Beamish, R. J. and McFarlane, G. A. 1987. Current trends in age determination conchonius (Ham.-Buch) by a use of trunk vertebrae. Periodic Research. 2: 49-51. || Beamish, K. J. and McFariane, G. A. 1967. Current trends in age determination methodology. pp. 15-42. In: R.C. Summer felt & G.E. Hall (ed.) Age and Growth of Fish, Iowa State University Press, Ames, Iowa. || Biju Kumar, A. 2000. Exotic fishes and freshwater fish diversity. Zoos Print Journal. 15(11): 363-367. | Bio, E. and Ikusemiju, K. 1981. Age and growth determination in the catfish, Chrysichthys nigrodigitatus (Lacepede) by use of the dorsal spine. Journal of Fish Biology. 19: 345-351. || Booth, A. J. and Merron, G. S. 1996. The age and growth of the greenhead tilapia Oreochromis macrochir (Pisces: Cichidae) from the Okavango Delta, Botswana. Hydrobiologia, 321: 29-34. || Braaten, P. J., Doeringsfeld, M. R. and Guy, C. S. 1999. Comparison of age and growth estimates for river carp suckers using scales and dorsal fin ray sections. N. Am. J. Fish. Manage. 19:786–792. || Brown, C. A. and Gruber, S. H. 1988. Age assessment of the lemon shark, Negaprion brevirostris, using tetracycline validated vertebral centra. Copeia. 3: 747-753. [] Buckmeier, D. L., Irwin, E. R., Betsill, R. K. and Prentice, J. A. 2002. Validity of Otolith and Pectoral Spines for Estimating Ages of Channel Catfish. North Amer. J. Fish. Manage. 22: 934 Irwin, E. R., Betslill, R. K. and Prentice, J. A. 2002. Validity of Otolith and Pectoral Spines for Estimating Ages of Channel Cattish. North Amer. J. Fish. Manage. 22: 934 – 942. || Campana, S. E. 2001. Accuracy, precision, and quality control in age determination, including a review of the use and abuse of age validation methods. Journal of Fish Biology. 59:197–242. | Canonico, G. C., Arthington, A., McCrary, J. K. and Thieme, M. L. 2005. The effects of introduced tilapias on native biodiversity Aquat. Conserv. Mar. Freshw. Ecosyst. 15(5):463-483. || Colombo, R. E., Phelps, Q. E., Miller, C. M., Garvey, J. E., Heidinger, R. C. and Richards, N. S. | 2010. Comparison of channel catfish age estimates and resulting population demographics | using two common structures. N. Am. J. Fish. Manage. 30: 305-308. || David, A. and Pancharatna, K. 2003. Age determination of the Indian whiting, Sillago indica using otolith ring count. Indian Journal of Fisheries. 50(2): 215 – 22. | Devries, D. R. and Frie, R. V. 1996. Determination of age and growth. In: Murphy B. R., Willis D. W., editors. Fisheries Techniques. 2nd edn. Bethesda, MD: American Fisheries. Sologi. 216 – 248. Society, 1962. Determination of age and growth. In: Mulphy B. K., Wins D. W., editors. Fisience feelingues, 2nd edit. Between Based, MD. Anterican Fisience Society, 1962. Deta, A. Sana Kumar, K. 2006. Age and growth patterns in Channa marulius from Harike Wethand (A Ramsar site), Punjab, India. J. Environ. Biol. 27(2): 377-380. | Ekingen, G. and Polat, N. 1987. Age determination and length weight relations of Capoeta capoeta umbla (Heckel, 1843) in Lake Keban. Turk. J. Zool., 11, 515. || Erickson, C. M. 1983. Age determination of Manitoban walleyes using otoliths, dorsal spines, and scales. N. Am. J. Fish. Manage. 3176-181. || Fowler, A. J. and Doherty, P. J. 1992. Validation of annual growth increments in the otoliths of two species of damsel fish from the southern Great Barrier Reef. Australian Journal of Marine and Freshwater Research. 43: 1057-1068. || Ganie, M. A., Bhat, M. D., Khan, M. I., Parveen, M., Balkhi, M. H. and Malla, M. A. 2013. Invasion of the Mozambique tilapia, Oreochromis mossambicus (Pisces: Cichlidae; Peters, 1852) in the Yamuna River, Uttar Pradesh, India. J. Ecol. and the Nat. Environ. 5(10): 310-317. | | Hanumantha Rao, L. 1974. Studies on the biology of Cirrhinus mrigala (Ham.) of the river Godavari. Indian Journal of Fisheries. 21 (2): 303-322. | | Hile, R. 1970. Body-scale relation and calculation of growth in fishes. Transactions of the American Fisheries Society. 99:468-474. | Jain, A. K. and Gupta, A. K. 1994. Occurrence of Tilapia in Jaisamand Lake. A threat to indigenous fishery resources of Rajasthan. Fishing Chimes. 1994: 15. | Johal, M. S. and Tandon, K. K. 1992. Age and growth of the carp Catla catla from Northern India. Fisheries Reserch. 14:83-90. | Kanwal, B. P. S. and Pathani. S. S. 2011. Age-Growth, Length-Weight and Condition Factor of a Hill Stream Fish, Garra Lamta (Hamilton-Buchanan)of Kumaun Himalay, Nat. and Sci. 9(8): 199-206. | Kimura, D. K. and Lyons, J. J. 1991. Between-reader bias and variability in the age determination process. Fish Bull. U. S. 89: 53 - 60. || Kocovsky, P. M. and Carline, R. F. 2000. A comparison of methods for estimating ages of unexploited walleyes. N. Am. J. Fish. Manage. 20:1044-1048. || Lakra, W. S., Singh, A.K. and Ayyappan, S (eds.) 2008. Fish Introductions in India: Status, | Potential and Challenges. Narendra Publishers, New Delhi, India. || Lowerre-Barbieri S., Chittenden, M. E. Jr, and Jones, C. M. 1994. A comparison of a validated otolith method to age weak fish, Cynoscion regalis, with the traditional scale method. United States Fish Bulletin. 92: 555-568. || Maceina, M. J. and Sammons, S. M. 2006. An evaluation of different structures to age freshwater fish from a north¬eastern US river. Fish. Manage. Ecol. 13: 237-242. || Mayekiso, M. and Hecht, T. 1988. Age and growth of Sandelia bainsii Castelnau (Pisces: Anabantidae) in the Tyume River, Eastern Cape (South Africa). South African Journal of Zoology. 23: 295-300. || Mills, K. H., and Beamish, R. J. 1980. Comparison of fin-ray and scale age determinations for lake whitefish (Coregonus clupeaformis) and their implications for estimates of growth and annual survival. Canadian Journal of Fisheries and Aquatic Sciences. 37: 534-544. || Nash, M. K. and Irwin, E. R. 1999. Use of implications for estimates of growth and annual survival. Canadian Journal of Fisheries and Aquatic Sciences. 37: 534-544. [] Nash, M. K. and Iwin, E. R. 1999. Use of otoliths versus pectoral spines for aging adult flathead catfish. In Proceedings of the International Ictalurid Symposium (Irwin, E. R., Hubert, W. A., Rabeni, C. F., Schramm, H. L., Jr. & Coon, T., eds), pp. 309.316. American Fisheries Society, Symposium 24, Bethesda, Maryland | Newman, S.J., M. Cappo and D.M. Williams 2000. Age, growth and mortality of the stripey, Lutjanus carponotatus (Richardson) and the brown-striped snapper, L. vitta (Quoy and Gaimard) from the central Great Barrier Reef, Australia. Fisheries Research. 48: 263-275. || Ovchynnyk, M. M. 1969. On age determination with scales and bones of the white sucker, Catostomus commersoni (Lacepe'de). Zoologischer Anzeiger. 175:325–345. || Pantulu, V. R. 1961. On the use of pectoral spines for the determination of age and growth of Mystus gulio (H). Proceeding National Institute of Science India. 27B: 1-30. || Pantilu, J. and Tomas, J. 2001. Validation of age estimation and back-calculation of fish length based on otolith microstructures in tilapias (Pisces, Cichlidae). Fish. Bull. 99: 139-150. || Phelps, Q. E., Edwards, K. R. and Willis, D. W. 2007. Precision of five structures for estimating age of common carp. N. Am. J. Fish. Manage. 27: 103-105. || Polat, N. and Kukul, A. 1990. Age determination methods of Trachurus trachurus in the Black Sea. In: N. J. Zool. ZS: 441-446. |Quinn, S. P. and M. R. Ross, M. R. 1982. Annulus formation by white suckers and the reliability of pectoral fin rays for ageing them. N. Am. J. Fish. Manage. 2:204–208. || Saidapur, S. K. 1989. Reproductive cycles in Fishes. In: Reproductive cycles of Indian Vertebrates. P. 166-224. Saidapur, S. K. Ed. New Delhi, Allide Press. || Scidmore, W. J. and Glass, A. W. 1953. Use of pectoral fin rays to determine of otolith and scale-based ageing in mark-recaptured Chesapeake Bay striped bass, Morone saxatilis. Fish. Bull. determination of Indian tisnes using scales, otoliths, and other hard parts. Indian J. Fish. 46(1): 1 – 22. [] Singn, S., Sharma, L. L. and Saini, V. P. 1998. Age, growth and harvestable size of Labeo rohita (Ham.) from the lake Jaisamand, Rajasthan, India. Indian J. Fish. 45(2): 169 – 175.] Shepherd, G. R. 1988. Age determination methods for northwest Atlantic species. Weak fish Cynoscion regalis. NOAA Tech.71 – 76.] Staples, D. J. 1971. Methods of ageing red gumard (Teleosti: Triglidae) by fin rays and otoliths. New Zealand J. Mar. Freshwater Res., 5(1): 70-79.] Sudarashan, S. and Kulkarni, R. S. 2013. Age determination and age related biochemical changes in the hepatic tissue of the freshwater fish Notopterus notopterus. IJAR. 3(11): 556-558.] Sulistiono, M. Yokota, S. Kitada and Watanabe, S. (1999). Age and growth of Japanese whiting Sillago japonica in Tateyama Bay. Fisheries Science (Tokoyo). 65(1): 117-122.] | Sullivan, S. O., Moriarty, C., FitzGerald, R. D., Davenport, J. and Japanese Whiting Sillago Japonica in Tateyama Bay. Fisheries Science (Tokoyo), 55(1): 117-122. || Sullivian, S. O., Monarty, C., FitzGerald, K. D., Davenport, J. and Mulcahy, M. F. 2003. Age, growth and reproductive status of the European conger eel, Conger conger (L) in Irish coastal waters. Fisheries Research, 64: 55-69. | Sylvester, R. 2006. Comparison of White Sucker Age Estimates from Scales,Pectoral Fin Rays, and Otoliths. N. Am. J. Fish. Manage. 26:24–31. || Ujjania N. C., Sharma L. L. and Srivastva, R. M. 2013. Assessment of age and growth of exotic fish tilapia (Oreochromis mossambicus p.) In lake Jaisamand, India. Indian Journal of Fundamental and Applied Life Sciences. 3 (4): 27-34. || Ujjania, N. C. 2012. Comparative age and growth of Indian major Carp (Catla catla ham. 1822) in Selected water bodies of Southern Rajasthan, India. Research Journal of Recent Sciences. 1: 17-22. || Wartenberg, R., Booth, A. J. and Weyl, O. L. F. 2011. A comparison of three techniques for fluorochrome marking of juvenile Clarias gariepinus otoliths. African Zoology. 46(1): 72–77. || Werder, U. and Soares, G. M. 1985. Age determination by sclerite numbers, and scale variations in six species from the central Amazon (Osteichthyes, Characoidei). Animal Research and Development. 21: 23-46. || Whitney, R. R., and Carlander, K. D. 1956. Interpretation of body-scale regression for computing body length of fish. J. Wildlife Manag. 20: 21-27. || Zar, J. H 1984. Biostatistical Analysis scored edition. prentice... Hall: Englewood cliffs. N. 178 PP. H. 1984. Biostatisical Analysis second edition, prentice – Hall, Englewood cliffs. NJ, 718 PP. |