RESEARCH PAPER

Biology



Effect of Malathion on Protein Content, Reduced Glutathione and Lipid Peroxidation Level on Liver of *Bufo melanostictus* Schneider, 1799

KEYWORDS	Bufo melanostictus, malathion, reduced glutathione, lipid peroxidation, liver				
Puspanjali Parida		Priyadarsini Naik	Lakshmipriya Mohanta		
P.G. Department of Zoology, North Orissa University, Baripada-757003 (Odisha),India		P.G. Department of Zoology, North Orissa University, Baripada-757003 (Odisha),India	P.G. Department of Zoology, North Orissa University, Baripada-757003 (Odisha),India		

ABSTRACT Bufo melanostictus (n=20) of various sizes (body weight: 90g- 120g) were divided into four groups of 5 animals each. Group I (control) animals received distilled water; Group II-IV (experimental) treated orally with 0.1µI malathion /g body weight. The treated animals were sacrificed after the time intervals of 24 hour, 48 hour and 72 hour (Group II-IV) whereas the control animal was sacrificed immediatetly (0h) and the liver was dissected out and kept at 00C. The protein content, reduced glutathione level (GSH) and lipid peroxidation (LPX) level were measured in the liver of B. melanostictus in both control and experimental group. Variation of different parameteres of liver were observed at different time intervals. On the basis of the results, It is concluded that malathion even at low dose altered the biochemical parameters and induces oxidative stress.

INTRODUCTION

There is an increasing concern about the ecological effect of the use of pesticides. It has been reported that pesticide ingestion either by direct or indirect exposure may lead to generation of reactive oxygen species (ROS), which are detrimental to the health of humans and non-target organisms (Otitoju and Onwurah, 2007). A good bioindicator should have a well-known taxonomy and ecology, be distributed over a broad geographic area, have a high degree of ecological specialization as specialized species are far more vulnerable to environmental perturbations compared to generalists, be cost-effective and relatively easy to survey (Rainio and Niemela 2003; Brischoux et al. 2009).

Although pesticides have long been suggested as a possible cause of amphibian declines (Carey and Bryant 1995, Stebbins and Cohen 1995, Drost and Fellers 1996, Lips 1998), there have been few toxicological studies on declines. Malathion is registered for use on food, feed, and ornamental crops and in mosquito, boll weevil and fruit fly eradication programs (Ware, 2000).

Malathion is toxic through skin contact, ingestion, and inhalation exposure (Tomlin, 2006). In the present study generation of ROS in liver of Bufo melanostictus in response to malathion were estimated by measuring reduced glutathione and lipid peroxidation level after different time intervals of 24 hour, 48 hour and 72 hour and compared against the control (0 hour).

MATERIALS AND METHODS

B. melanostictus were collected locally near the North Orissa University campus, during night and early morning time. They were acclimatized for seven days prior to the experiment. Bufo melanostictus (n=20) of various sizes (body weight: 90g- 120g) were divided into four groups of 5 animals each. Group I (control) animals received distilled water; Group II-IV (experimental) treated orally with 0.1μ l malathion /g body weight. The treated animals were sacrificed after the time intervals of 24 hour, 48 hour and 72 hour (Group II-IV) whereas the control animal was sacrificed immediatetly (0h).

The liver of both control and experimental group were dissected out quickly and kept at 0°C. A 20% homogenate was prepared with phosphate buffer (pH 7.4). The tissue homogenate was centrifuged at 4000 rpm for 10 minutes.

Protein

Protein estimation of the sample were made according to the method of Lowry et. al.,(1961). To 0.1ml suitably homogenate of tissue 0.4ml 0f distilled water was added. Then 5 ml of biuret reagent (containing alkaline Na₂CO₃, 0.5% CuSO₄ solution and 1% Sodium potassium tartarate solution in the ratio 100:2:2) was added and properly mixed up. After 10 minutes of incubation at room temperature 0.5ml of Folin Ciocalteau phenol reagent (the commercial reagent diluted three times with distilled water) was added and incubated at 37°C for 30 minutes at room temperature. Absorbance was measured at 700 nm against an appropriate blank

Reduced Glutathione

Reduced glutathione of the sample were estimated by Ellman (1959) method. 0.7ml of the tissue homogenate was added to 0.7ml of TCA. Then the substances in the tubes were centrifuged at 4000 rpm for 10 minutes. 0.5ml supernatant was added to 2.5ml of DNTB (DNTB 30 mM) was diluted in PO_4 buffer 100 times. The absorbance was taken at 412 nm with in between 5 to 30 minutes against a appropriate blank.

Lipid Peroxidation

Lipid peroxidation of the sample is estimated as thiobarbiturate acid reacting substance (TBARS) by thiobarbituric acid (TBA) according to the method of Ohkawa et. al. (1979). 3.8ml of TBA reagent contain (2ml of 8.1% SDS , 1.5ml of 20% acetic acid of pH 3.5, 1.5ml of 0.8% aqueous solution of TBA, 5ml of distilled water and 1ml of BHT) was added to 0.2ml of suitably diluted post nuclear supernatant. After mixing thoroughly, the test tube's substance was boiled in water bath for 1 hour. The tubes were cooled down to the room temperature. Then the tube substances ware centrifuged at 4000 rpm for 10 minutes. The absorbance of the supernatant was measured at 532 nm against a appropriate blank.

RESEARCH PAPER

Table1: Comparison of protein content (mg/g tissue), GSH level (µ mol/g tissue), LPX level(n mol TBARS/mg protein)of liver in Bufo melanostictus after treatment of malathion (0.1µl/g body weight) at different time interval. The value are expressed in Mean± S.D.

		-		
treatm		Protein con- tent (mg/g tissue)	GSH level (µ mol/ g tissue)	LPX level (n mol TBARS/ mg protein)
0h (I)	Group-	63.89±0.379	0.16±0.001	62.46±1.147
24h (11)	Group-	65.31±2.035	0.24±0.017	20.072±0.069
48h (111)	Group-	47.28± 0.794	0.32±0.012	80.04±0.893
72h (IV)	Group-	40.79±0.979	0.14±0.007	31.561±1.383

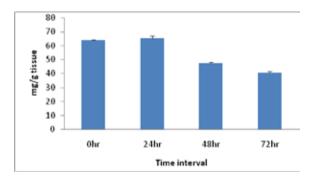


Fig : 1 Comparison of protein content (mg/g tissue) of liver in Bufo melanostictus treated with malathion (0.1µl/g body weight) at different time interval.

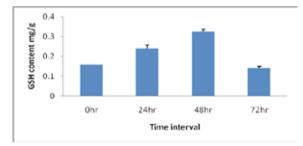


Fig : 2 Comparison of GSH level (µ mol/g tissue) of liver in Bufo melanostictus treated malathion (0.1µl/g body weight) at different time interval.

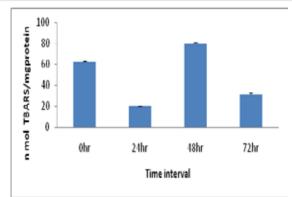


Fig: 3 Comparison of LPX level (n mol TBARS/mg protein) of liver in Bufo melanostictus treated with malathion (0.1µl/g body weight) at different time interval.

RESULTS AND DISCUSSION

Protein content (mg/g tissue) in the liver of B. melanostictus treated with malathion were 63.89±0.379 mg/g tissue, , 65.31±2.035 mg/g tissue , 47.28± 0.794 mg/g tissue and 40.79±0.979 mg/g tissue at 0h, 24h, 48h and 72 h respectively (table 1 and fig.1). In other words the protein content of liver of B. melanostictus was almost same to that of 24h but, decreases gradually at 48h and 72h of time interval.

GSH content (µ mol/g tissue) in liver tissue of B. melanostictus treated with malathion were 0.16±0.001 µ mol/g tissue, 0.24±0.017 µ mol/g tissue, 0.32±0.012 µ mol/g tissue and 0.14±0.007 at 0h, 24h, 48h and 72 h respectively (table 1 and fig. 2). In other words the GSH content of liver of B. melanostictus increased with increase of time intervals upto 48h and protect the animal from oxidative stress but after 72h the GSH content gradually decreased.

The LPX level (n mol TBARS/mg protein) in liver tissue of B. melanostictus treated with malathion is 62.46±1.147 n mol TBARS/mg protein, 20.072±0.069 n mol TBARS/ mg protein, 80.04±0.893 n mol TBA-RS/mg protein and 31.561±1.383 n mol TBARS/mg protein at 0h, 24h, 48h and 72 hour (table 1 and fig. 3). The LPX content was highest at 48h may be due to decrease in GSH content. But again drastically at 72h which indicate, there must be some other antioxidant which taken care to combat the liver from oxidative stress.

From the above it is concluded that Bufo has well developed antioxidant defence mechanism to resist the pesticide insult though all the parameters of antioxidant system were not measured.



Brischoux F., Bonnet X. & Legagneux P. 200). Are sea snakes pertinent bio-indicators for coral reefs? A comparison between species and tes. Marine Biology 156: 1985-1992. || Carey, C., and C. J. Bryant. 1995. Possible interrelations among environmental toxicants, amphibian development, and decline of amphibian populations. Environmental Health Perspectives 103:13-17. || Drost, C. A., and G. M. Fellers. 1996. Collapse of a regional frog fauna in the Yosemite area of the California Sierra Nevada, USA. Conservation Biology 10:414–425. || Ellman, G.L. 1959. Tissue sulphydryl groups. Arch. Biochem. Biophy. pp 82:70-77. || Lips, K. R. 1998. Decline of a tropical montane amphibian fauna. Conservation Biology 12:106–117. || Lowry, O.H., Resbrough, N.J., Farr, A.L. Randoll, R.J. 1951. Protein measurement with the Folin-phenol reagent. J.Biol. Chem. 19: 265-275. || Ohkawa H., Ohishi, N. and Yagi, K. 1979. Assay of LPX in animal terms to kindender in a conservation 2007. Clutetiking Stermere (NET) activity on existence of the california conservation Biology 20106–117. || Lowry, O.H., Resbrough, N.J., Farr, A.L. Randoll, R.J. 1951. Protein measurement with the Folin-phenol reagent. J.Biol. Chem. 19: 265-275. || Ohkawa H., Ohishi, N. and Yagi, K. 1979. Assay of LPX in animal terms to kindender in a conservation Biology C. Chemere (NET) activity on the environmental NET. 2007. Clutetiking Stereofference (SET) and Clutetiking Activity of the server (NET) activity on the environmental NET. 2007. Clutetiking Stereofference (SET) and Server (NET) activity on the internet in the server (NET) activity on the internet internet (NET) activity on the internet (NET) activ tissues by thiobarbituric acid reaction. Anal Biochem. Physiol.118C (1):33-37. || Otitoju O, Onwurah INE. 2007. Glutathione S-transferase (GST) activity as a biomarker in ecological risk assessment of pesticide contaminated environment. Afr. J. Biotechnol., 6: 1455-1459 || Rainio J. & Niemela J. (2003). Ground beetles (Coleoptera: Carabidae) as bioindicars. Biodiversity and Conservation 12: 487-506. [Stebbins, R. C., and N. W. Cohen. 1995. A natural history of amphibians. Princeton University Press, Princeton, New Jersey, USA. [] Tomlin, C. D. S. 2006. The Pesticide Manual, A World Compendium. 14th ed.; British Crop Protection Council: Alton, Hampshire, UK. pp 642-643. || Ware, G. W. 2000. The Pesticide book. Frenso C A: Thompson Publications. pp.52-67. |