



## Larvicidal Activity of *Citrullus colocynthis* (L.) Schrad (Cucurbitaceae) isolated fractions Against *Aedes aegypti* (L.), *Anopheles stephensi* Liston and *Culex quinquefasciatus* Say (Diptera: Culicidae)

## KEYWORDS

*Citrullus colocynthis*, larvicidal activity, *Aedes aegypti*, *Anopheles stephensi*, *Culex quinquefasciatus*

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**ABSTRACT** Vector-borne diseases are one of the greatest contributors to human mortality and morbidity in tropical and subtropical regions. Vector control remains the most effective measure and is often the only way to prevent disease outbreaks since there are no vaccines for many vector-borne diseases. The extensive and repeated use of synthetic organic insecticides have led to disrupted natural biological control systems heading to resurgence and resistance in insects and destruction of non-target beneficial fauna, in turn resulting in environment imbalance and human health concerns. Plants have co-evolved with insects that have equipped them with a plethora of chemical defense, which can, in turn be used against insects. In the present study, the isolated fractions of *Citrullus colocynthis* dichloromethane whole plant extract were evaluated for larvicidal activity against the vector mosquitoes viz., *Aedes aegypti*, *Anopheles stephensi* and *Culex quinquefasciatus*. Eight fractions viz., A, B, C, D, E, F, G and H were obtained from the residue of dichloromethane extract by column chromatography. Standard WHO protocols with minor modifications were adopted for the larvicidal bioassay. Larvicidal activity was evaluated against the isolated fractions at concentrations of 25, 50, 75 and 100 ppm. Larval mortality was observed for 24 hours. Amongst the isolated fractions tested, fraction 'C' showed 94.4%, 96.0% and 98.4% mortality against third instar larvae of *Aedes aegypti*, *Anopheles stephensi* and *Culex quinquefasciatus* at 100 ppm and  $LC_{50}$  values were 18.57, 23.48, 19.26 ppm and  $LC_{90}$  values were 98.65, 92.26, 84.84 ppm, respectively. Further investigations are needed to explore the larvicidal activity of the isolated fraction 'C' of dichloromethane whole plant extract of this plant and also the active ingredient(s) responsible for larvicidal activity.

## INTRODUCTION

The mosquitoes, commonly called "flying syringes", are sanguivorous vectors, which cause more sufferings to human than any other organism (Thomas *et al.*, 2014). Vector-borne diseases are one of the greatest contributors to human mortality and morbidity in tropical and subtropical regions. Every year, more than one billion people are infected and more than one million people die from vector-borne diseases including malaria, dengue, yellow fever, and lymphatic filariasis (WHO, 2014). Vector control remains the most effective measure and is often the only way to prevent disease outbreaks since there are no vaccines for many vector-borne diseases and drug resistance is an increasing threat. As stated by Amer and Mehlhorn (2006), larval stages of mosquitoes are attractive targets for pesticides to control mosquito populations. The advantage of targeting the larval stages are that mosquitoes are killed before they disperse to human habitations, and larvae, unlike adults, cannot change their behavior to avoid control activities (Killeen *et al.* 2002) and also to reduce overall pesticide use in control of adult mosquitoes by aerial application of adulticidal chemicals (Gleiser and Zygadlo, 2007). The use of conventional larvicides in the aquatic environment poses serious threats viz., undesirable action on human health and other beneficial organisms, environmental sustainability, and higher rate of biological magnification (Brown, 1986). Due to the limitations, such as insecticide resistance, environmental pollution, and high cost of synthetic insecticides, current studies are focused to find out natural substances particularly from plants to control the disease transmitting vectors. Various reports on the use of natural plant products against mosquito vectors have been reported (Sukumar *et al.*, 1991; Arivoli *et al.*,

1999, 2012a, b, 2015; Shaalan *et al.*, 2005; Sakthivadivel and Daniel, 2008; Ghosh *et al.*, 2012; Raveen *et al.*, 2012, 2014; Samuel *et al.*, 2012a, b; Vargas, 2012; Samuel and William, 2014).

*Citrullus colocynthis* (L.) Schrad a medicinal plant belonging to the family Cucurbitaceae is commonly called 'bitter apple' or 'wild water melon' (Mahmoud *et al.*, 2009), in Tamil as Paedikari attutummatti', in Hindi as 'Indrayan' and in Sanskrit as 'Indravaruni' (Pravin *et al.*, 2013). The plant is found throughout India and Ceylon, both wild and cultivated. It is also indigenous in the West Asia, tropical Africa and Mediterranean regions (Pravin *et al.*, 2013). In traditional medicine, the plant has been utilized as an abortifacient (Duke, 2006), purgative (Aburjai *et al.*, 2007), in curing tumors, leucoderma, elephantiasis, ulcers and in removing kidney stones (Shah *et al.*, 1989). The powerful medicinal values of the pulp are due to the presence of an amorphous glucosid 'colocynth'. The plant also possesses anti-inflammatory (Belsem *et al.*, 2011), anticandidal and antibacterial (Rasool and Jahanbakhsh, 2011), antioxidant (Saba and Oridupa, 2010), analgesic (Marzouk *et al.*, 2010, 2011), hypoglycemic (Agarwal *et al.*, 2012), hypolipidemic (Rahbar and Nabipour, 2010), antialopecia (Dhanotia *et al.*, 2011), antidiabetic (Huseini *et al.*, 2009) and antifertility (Chaturvedi *et al.*, 2003) properties. The phytochemical compounds present in the plant include stearic acid, myristic acid, palmitic acid, oleic acid, linoleic acid, linolenic acid, flavonoid glycoside quercetin, flavone-3-glucoside viz., iso-vitexin, iso-orientine and isoorientine-3-methyl ether, colocynthoside A and B, cucurbitacin E 2-O-beta-D-glucoside, cucurbitacin E, 2-O-beta-D-glucopyranosyl-cucurbitacin B, 2, 25-di-O-beta-D-glucopyranosyl-cucurbitacin L,

colocynthin, colocynthein, colocynthetin, pectin gum and albuminoids (Nayab et al., 2005; Yoshikawa et al., 2007; Gurudeepan et al., 2010). Further, *Citrullus colocynthis* gained increasing attention as a natural insecticide and its activity have been evaluated against many important insect species. It possesses deterrent, antifeedant, growth regulating and fertility reducing properties in insects (Prabuseenivasan et al., 2004) and insecticidal effect against the aphid *Aphis craccivora* (Torkey et al., 2009). The whole plant extracts of *Citrullus colocynthis* were assayed for their toxicity against the larvae of *Culex quinquefasciatus* (Rahuman et al., 2008). In addition, Arivoli and Samuel (2011) have reported the larvicidal activity of the crude dichloromethane extracts of *Citrullus colocynthis* whole plants against *Aedes aegypti*, *Anopheles stephensi* and *Culex quinquefasciatus* and recently Satti and Edriss (2015) have reported the larvicidal activity of petroleum ether fruit extract of *Citrullus colocynthis* against *Anopheles arabiensis*. Therefore, the present study was carried out to study the larvicidal activity of isolated fractions of *Citrullus colocynthis* dichloromethane whole plant extracts against the vector mosquitoes viz., *Aedes aegypti*, *Anopheles stephensi* and *Culex quinquefasciatus*.

## MATERIALS AND METHODS

### Plant collection and preparation of crude extract

*Citrullus colocynthis* whole plants collected in and around Tamil Nadu, India were brought to the laboratory, shade dried under room temperature and powdered using an electric blender. Dried and powdered whole plants (1 kg) was subjected to extraction using 3 L of dichloromethane for a period of 72 hours to obtain the crude extracts using rotary vacuum evaporator. The dichloromethane crude extract thus obtained was refrigerated at 4 °C.

### Isolation and fractionation of crude extracts by column chromatography

The residue from the crude dichloromethane extract of *Citrullus colocynthis* (32.844g) was mixed with silica gel (60 -120 mesh, 100g) as admixture, subjected to column chromatography (si gel, 100 -200 mesh 300g) to obtain eight fractions viz., A, B, C, D, E, F, G and H by increasing polarity of eluents viz., hexane and ethyl acetate in the ratio of 100:0; 80:20; 60:40; 40:60; 20:80; 0:100 and finally ethyl acetate and acetone in the ratio of 50:50 and 0:100, respectively.

### Larvicidal bioassay

Bioassay was carried out against laboratory reared vector mosquitoes free of exposure to insecticides. Standard WHO (2005) protocol with minor modifications was adopted for the study. The tests were conducted in glass beakers. Mosquito immature particularly third instar larvae were obtained from laboratory colonized mosquitoes of F<sub>1</sub> generation. Concentrations of 25, 50, 75 and 100 ppm were prepared. Twenty five healthy larvae were released into each 250 ml glass beaker containing 200 ml of water and test concentration. Larval mortality was observed for 24 hours after treatment. Larvae were scored as dead when they showed no signs of movement. A total of five trials with three replicates per trial for each concentration were carried out. Distilled water as control was run simultaneously. The larval per cent mortality was calculated and when control mortality ranged from 5-20% it was corrected using Abbott's formula (Abbott, 1925). SPSS version 11.5 was used for determination of LC<sub>50</sub> and LC<sub>90</sub> values (SPSS, 2007). The percentage data obtained was angular transformed. Data from mortality and effect of concentrations were subjected to two way ANOVA followed by Tukey's

test (P<0.05) to determine the difference in larval mortality between concentrations.

## RESULTS

The dichloromethane extract of *Citrullus colocynthis* was subjected to column chromatography with varying proportions of hexane: ethyl acetate and ethyl acetate: acetone. Results revealed that among eight fractions (A, B, C, D, E, F, G and H), fraction 'C' showed 98.4% larval mortality against *Culex quinquefasciatus* followed by 96.0% and 94.4% against *Anopheles stephensi* and *Aedes aegypti*, respectively at 100 ppm. Other fractions showed minimum mortality (Table 1; Figure 1). No mortality was observed in eighth (H) fraction as well as in control. The fraction 'C' exhibited LC<sub>50</sub> values of 18.57, 19.26, 23.48 ppm against *Aedes aegypti*, *Culex quinquefasciatus* and *Anopheles stephensi* respectively and LC<sub>90</sub> values of 84.84, 92.26 and 98.65 ppm, against *Culex quinquefasciatus*, *Anopheles stephensi* and *Aedes aegypti*, respectively (Table 2).

## DISCUSSION

In a preliminary investigation, different solvent extracts (hexane, diethyl ether, dichloromethane and ethyl acetate) of *Citrullus colocynthis* whole plants were tested against *Aedes aegypti*, *Anopheles stephensi* and *Culex quinquefasciatus* for larvicidal activity where dichloromethane extract was found to be active (Arivoli and Samuel, 2011) and in the present study the active substances of the dichloromethane extract were found in the 'C' fractionated group, which was indicated by the lowest LC<sub>50</sub> value reported. The finding of the present study is in line with the high potential of non-polar (dichloromethane, chloroform and hexane) extracts (Chaaib, 2004; Krishnappa et al., 2012) demonstrated against mosquito larvae.

Rahuman et al. (2008) reported that the bioassay-guided fractionation of *Citrullus colocynthis* petroleum ether leaf extract led to the separation and identification of fatty acids; oleic acid and linoleic acid which were isolated and identified as mosquito larvicidal compounds. Oleic and linoleic acids were quite potent against fourth instar larvae of *Aedes aegypti* (LC<sub>50</sub> 8.80 and 18.20 ppm), *Anopheles stephensi* (LC<sub>50</sub> 9.79 and 11.49 ppm) and *Culex quinquefasciatus* (LC<sub>50</sub> 7.66 and 27.24 ppm). Similarly, according to Sayed et al. (1973), *Citrullus colocynthis* contains oleic and linoleic acids. These substances are known to be more effective against aquatic insects. Alkaloids as well as saponins have also been detected in extracts of this plant and were preferably biologically active against aquatic insects (Hatim et al., 1989) may justify to the action of larvicidal activities. In addition, triterpenoids and saponins in chloroform; saponins in hexane; steroids, saponins, tannins and alkaloids in methanol extracts of *Adansonia digitata* had revealed their toxicity against *Aedes aegypti* and *Culex quinquefasciatus* larvae (Krishnappa et al., 2012).

Basheer (2014) tested the *Ricinus communis* ethyl acetate leaf extract fractions (F1-F7) against the third instar larvae of *Anopheles arabiensis* after 24 hours of exposure and found the fraction F3 to exhibit the highest larvicidal activity whose LC<sub>50</sub> value was 125.0 ppm. da Silva et al. (2007) indicated out of eight fractions each for the hexane (CRH1-CRH8) and methanol (CRM1-CRM8) extracts of *Copaifera reticulata* tested against the third instar larvae of *Aedes aegypti*, two hexane (CRH1, CRH5) and two methanol (CRM1, CRM5) fractions showed the highest toxicity against *Aedes aegypti* larvae and their LC<sub>50</sub> values were 2.3, 13.9 and 0.8, 10.5 ppm respectively.

Albaba et al. (2015) tested the n-hexane leaf extract fractions (CF1-CF11) of *Chromolaena odorata* against *Aedes vittatus* and found the fractions CF3, CF5 and CF6 to exhibit the highest larvicidal activities and their LC<sub>50</sub> values were 1.66, 1.81 and 1.30 ppm respectively. Chintem et al. (2014) stated that *Datura stramonium* leaf methanol extract fractions (DSEE-F1 to DSEE-F7) when tested against the third instar larvae of *Culex quinquefasciatus*, highest mortality rate was achieved in DSEE-F1 and its LC<sub>50</sub> value was 4.39 ppm after 24 hours. Nzelibe and Chintem (2013) tested the *Ocimum gratissimum* leaf n-hexane fractions (F1-F6) against *Culex quinquefasciatus* larvae, and found the fraction F1 to be effective with a LC<sub>50</sub> value of 1.49 ppm. Tomass et al. (2011) tested the fractions (F1-F3) of *Jatropha curcas* methanolic leaf extract against *Anopheles arabiensis* and found fraction F1 and F2 to exhibit high larvicidal activity with LC<sub>50</sub> values of 28.65 and 30.40 ppm respectively. Fraction F3 gave a LC<sub>50</sub> value of 80.70 ppm.

All fractions (hexane, dichloromethane, acetone, ethyl acetate and methanol) of the methanolic leaf extract of *Spondias mombin* were very effective against *Aedes aegypti*. Hexane, dichloromethane and acetone fractions recorded maximal mortality against *Anopheles gambiae* and it was only hexane fraction that registered maximal mortality against *Culex quinquefasciatus* (Eze et al., 2014). The bioassay-guided fractionation of *Achyranthes aspera* led to the separation and identification of a saponin as a potential mosquito larvicidal compound with LC<sub>50</sub> value of 18.20 and 27.24 ppm against *Aedes aegypti* and *Culex quinquefasciatus* respectively (Bagavan et al., 2008). The compounds, ecobolin A and ecobolin B isolated from the ethyl acetate extract of *Ecbolium viride* root showed larvicidal activity against the third instar larvae of *Culex quinquefasciatus* (Cecilia et al. 2014).

Botanical pesticides have the advantage of providing novel modes of action against insects that can reduce the risk of cross-resistance as well as offering new leads for design of target specific molecules (Zhou et al., 2012). Commonly a connection is extrapolated between plant activity based on traditional experience and insecticidal activity against mosquitoes. In general, extracts of the plants derived from specific solvents can influence the bioactivity, probably because of the active components present in large quantities (Oliveira et al., 2010). In addition, insecticidal effects of plant extracts vary not only according to plant species, mosquito species, geographical varieties and parts used, but also due to extraction methodology adopted and the polarity of the solvents used during extraction (Ghosh et al., 2012). A considerable number of plants have been extensively screened/ studied for their mosquito larvicidal activity. Though several plant extracts have been reported for mosquitocidal activity, only a few botanicals have been studied for the isolation of active molecule responsible for the activity. In that way, the results of the present study offer a possible way for further investigations to find out the active mosquito larvicidal compound from the dichloromethane extract of *Citrullus colocynthis*. The fractions of dichloromethane extract in the present study contained one or more phytochemical compounds and hence, the larvicidal activity might be due to the presence of those phytoconstituents.

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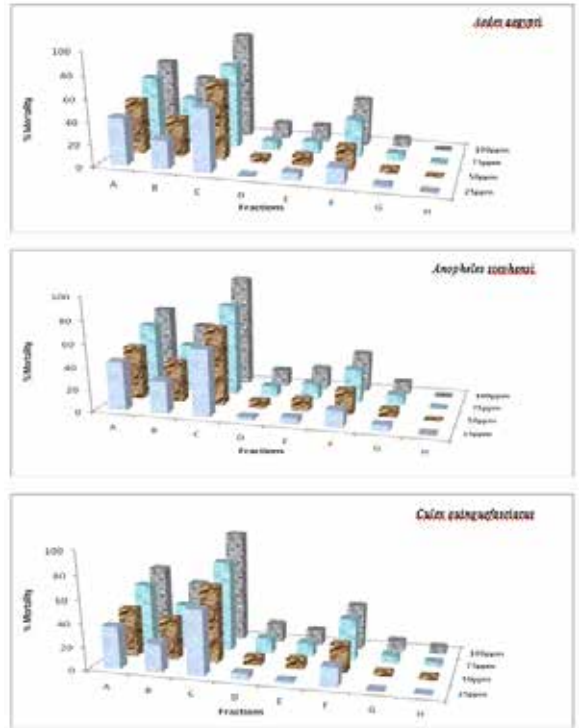
**Table 1. Per cent larvicidal activity of *Citrullus colocynthis* isolated fractions of dichloromethane whole plant extract against vector mosquitoes**

Concentration (ppm)	Isolated fractions							
	A	B	C	D	E	F	G	H
<b><i>Aedes aegypti</i></b>								
25	42.4 ±2.19 (40.6) <sup>e</sup>	25.6 ± 2.19 (30.4) <sup>d</sup>	56.8 ±4.38 (48.9) <sup>f</sup>	0.0 ±0.0 (0.0) <sup>a</sup>	06.4 ±2.19 (14.7) <sup>b</sup>	14.4 ±2.19 (22.3) <sup>c</sup>	02.4 ±2.19 (8.9) <sup>ab</sup>	0.0 ±0.0 (0.0) <sup>a</sup>
50	48.0 ±2.82 (43.9) <sup>e</sup>	33.6 ±2.19 (35.4) <sup>d</sup>	68.8 ±5.21 (56.0) <sup>f</sup>	04.8 ±1.78 (12.7) <sup>ab</sup>	08.8 ±1.78 (17.3) <sup>b</sup>	20.0 ±2.82 (26.6) <sup>c</sup>	3.2 ±3.34 (10.3) <sup>ab</sup>	0.0 ±0.0 (0.0) <sup>a</sup>
75	59.2 ±3.34 (50.3) <sup>e</sup>	42.4 ±2.19 (40.6) <sup>d</sup>	75.2 ±5.21 (60.1) <sup>f</sup>	08.0 ±2.82 (16.4) <sup>b</sup>	10.4 ±2.19 (18.8) <sup>b</sup>	32.8 ±3.34 (34.9) <sup>c</sup>	5.6 ±2.19 (13.7) <sup>ab</sup>	0.0 ±0.0 (0.0) <sup>a</sup>
100	66.4 ±4.56 (54.6) <sup>f</sup>	52.8 ±3.34 (46.6) <sup>e</sup>	94.4 ±3.57 (76.3) <sup>f</sup>	13.6 ±2.19 (21.6) <sup>bc</sup>	14.4 ±3.57 (22.3) <sup>c</sup>	42.4 ±3.57 (40.6) <sup>d</sup>	7.2 ±3.34 (15.6) <sup>b</sup>	0.0 ±0.0 (0.0) <sup>a</sup>
Control	0.0 ±0.0 (0.0) <sup>a</sup>	0.0 ±0.0 (0.0) <sup>a</sup>	0.0 ±0.0 (0.0) <sup>a</sup>	0.0 ±0.0 (0.0) <sup>a</sup>	0.0 ±0.0 (0.0) <sup>a</sup>	0.0 ±0.0 (0.0) <sup>a</sup>	0.0 ±0.0 (0.0) <sup>a</sup>	0.0 ±0.0 (0.0) <sup>a</sup>
<b><i>Anopheles stephensi</i></b>								
25	36.0 ±2.82 (36.9) <sup>d</sup>	24.0 ±2.82 (29.3) <sup>c</sup>	55.9 ±5.93 (48.4) <sup>e</sup>	04.8 ±1.78 (12.7) <sup>a</sup>	01.6 ±2.19 (7.39) <sup>a</sup>	16.8 ±3.34 (24.2) <sup>b</sup>	0.0 ±0.0 (0.0) <sup>a</sup>	0.0 ±0.0 (0.0) <sup>a</sup>
50	41.6 ±4.56 (40.2) <sup>c</sup>	35.2 ±3.34 (36.4) <sup>c</sup>	65.6 ±6.69 (54.1) <sup>d</sup>	07.2 ±1.78 (15.6) <sup>a</sup>	08.0 ±2.82 (16.4) <sup>a</sup>	24.0 ±2.82 (29.3) <sup>b</sup>	03.2 ±1.78 (10.3) <sup>a</sup>	02.4 ±2.19 (8.9) <sup>a</sup>
75	54.4 ±5.36 (47.5) <sup>d</sup>	38.2 ±4.56 (38.2) <sup>c</sup>	78.4 ±4.56 (62.3) <sup>e</sup>	12.8 ±3.34 (20.9) <sup>b</sup>	11.2 ±1.78 (19.6) <sup>b</sup>	35.2 ±1.78 (36.4) <sup>c</sup>	06.4 ±2.19 (14.7) <sup>ab</sup>	03.2 ±3.34 (10.3) <sup>a</sup>
100	61.6 ±4.56 (51.7) <sup>e</sup>	49.6 ±6.06 (44.8) <sup>d</sup>	96.0 ±4.00 (78.5) <sup>f</sup>	16.0 ±2.82 (23.6) <sup>b</sup>	12.8 ±3.34 (20.9) <sup>ab</sup>	38.4 ±4.56 (38.3) <sup>c</sup>	08.8 ±1.78 (17.3) <sup>ab</sup>	05.6 ±2.19 (13.69) <sup>a</sup>
Control	0.0 ±0.0 (0.0) <sup>a</sup>	0.0 ±0.0 (0.0) <sup>a</sup>	0.0 ±0.0 (0.0) <sup>a</sup>	0.0 ±0.0 (0.0) <sup>a</sup>	0.0 ±0.0 (0.0) <sup>a</sup>	0.0 ±0.0 (0.0) <sup>a</sup>	0.0 ±0.0 (0.0) <sup>a</sup>	0.0 ±0.0 (0.0) <sup>a</sup>
<b><i>Culex quinquefasciatus</i></b>								
25	43.2 ±3.34 (41.6) <sup>d</sup>	28.8 ±3.34 (32.5) <sup>c</sup>	59.2 ±5.21 (50.3) <sup>e</sup>	02.4 ±3.57 (8.9) <sup>a</sup>	04.8 ±1.78 (12.7) <sup>a</sup>	14.4 ±2.19 (22.3) <sup>b</sup>	04.8 ±3.34 (12.7) <sup>a</sup>	0.0 ±0.0 (0.0) <sup>a</sup>
50	47.2 ±5.21 (43.3) <sup>e</sup>	36.0 ±4.00 (36.9) <sup>d</sup>	68.8 ± 4.38 (56.0) <sup>f</sup>	05.6 ±2.19 (13.7) <sup>ab</sup>	09.6 ±2.19 (18.1) <sup>b</sup>	20.8 ±3.34 (27.1) <sup>c</sup>	06.4 ±3.57 (14.7) <sup>ab</sup>	0.0 ±0.0 (0.0) <sup>a</sup>
75	59.2 ±1.78 (50.3) <sup>e</sup>	42.4 ±2.19 (40.6) <sup>d</sup>	80.8 ± 5.93 (64.0) <sup>f</sup>	08.8 ±1.78 (17.3) <sup>b</sup>	13.6 ±2.19 (21.6) <sup>b</sup>	29.6 ±3.57 (32.9) <sup>c</sup>	08.8 ±1.78 (17.3) <sup>b</sup>	0.0 ±0.0 (0.0) <sup>a</sup>
100	66.4 ±6.06 (54.6) <sup>f</sup>	52.8 ±4.38 (46.6) <sup>e</sup>	98.4 ± 3.57 (82.7) <sup>f</sup>	14.4 ±2.19 (22.3) <sup>bc</sup>	18.4 ±3.57 (25.4) <sup>c</sup>	35.2 ±3.34 (36.4) <sup>d</sup>	10.4 ±3.57 (18.8) <sup>b</sup>	0.0 ±0.0 (0.0) <sup>a</sup>
Control	0.0 ±0.0 (0.0) <sup>a</sup>	0.0 ±0.0 (0.0) <sup>a</sup>	0.0 ±0.0 (0.0) <sup>a</sup>	0.0 ±0.0 (0.0) <sup>a</sup>	0.0 ±0.0 (0.0) <sup>a</sup>	0.0 ±0.0 (0.0) <sup>a</sup>	0.0 ±0.0 (0.0) <sup>a</sup>	0.0 ±0.0 (0.0) <sup>a</sup>

Values are mean (%) of the five-replicates of three trials  $\pm$  standard deviation. Figures in parenthesis are angular transformed. Different superscript alphabets indicate statistical significant difference at  $P < 0.05$  level by two way ANOVA followed by Tukey's test.

**Table 2. Probit analysis of larvicidal activity of activity of *Citrullus colocynthis* isolated fractions of dichloromethane whole plant extract against vector mosquitoes**

Vector mosquitoes	<i>Aedes aegypti</i>				<i>Anopheles stephensi</i>				<i>Culex quinquefasciatus</i>									
	LC <sub>50</sub> (ppm)	95% CL LL	95% CL UL	LC <sub>90</sub> (ppm)	LC <sub>50</sub> (ppm)	95% CL LL	95% CL UL	LC <sub>90</sub> (ppm)	LC <sub>50</sub> (ppm)	95% CL LL	95% CL UL	LC <sub>90</sub> (ppm)	95% CL LL	95% CL UL				
Fractions																		
A	50.35	42.36	56.99	200.87	173.78	244.23	67.02	60.93	73.55	207.14	180.61	248.24	50.12	41.90	56.89	203.67	175.62	248.99
B	93.62	86.13	103.87	226.42	197.57	270.49	102.62	93.14	116.66	250.76	214.57	309.10	94.07	85.41	106.61	249.68	212.52	310.68
C	18.57	3.96	28.17	98.65	88.23	114.76	23.48	11.55	31.70	92.26	83.16	105.79	19.26	4.68	28.66	84.84	75.79	98.71
D	163.81	146.94	190.35	242.14	210.95	291.77	204.78	172.41	264.83	343.75	279.32	464.32	190.91	158.64	238.22	301.19	240.27	429.83
E	281.67	213.11	466.36	497.89	361.46	867.49	200.27	162.31	287.71	323.18	250.53	492.53	190.86	163.35	239.30	323.69	267.41	423.84
F	115.06	106.33	127.09	221.11	196.95	255.82	126.06	113.23	145.84	265.53	227.28	326.77	138.13	123.04	161.92	277.18	236.07	343.83
G	300.10	203.50	864.79	477.23	307.56	1475.68	187.31	162.11	232.21	275.25	230.76	355.27	326.91	220.01	903.78	560.74	358.89	1655.92
H	-	-	-	-	-	-	221.34	166.51	434.44	321.76	229.92	682.50	-	-	-	-	-	-



**Figure 1. Larvicidal activity of *Citrullus colocynthis* isolated fractions of dichloromethane whole plant extract against vector mosquitoes**

## REFERENCE

- Abbott, W.S. 1925. A method of computing the effectiveness of an insecticide. *Journal of Economic Entomology*, 18: 265-267. | Aburjai, T., Hudaib, M., Tayyem, R., Yousef, M., and Qishawi. M. 2007. Ethno-pharmacological survey of medicinal herbs in Jordan, the Ajloun heights region. *Journal of Ethnopharmacology*, 110: 294-304. | Agarwal, V., Sharma, A.K., Upadhyay, A., Singh, G. and Gupta, R. 2012. Hypoglycemic effects of *Citrullus colocynthis* roots. *Acta Poloniae Pharmaceutica*, 69(1):75-79. | Albaba, S.U., Nzelibe, H.C., Inuwa, H.M., Chintem, D.G.W., Abdullahi, A.S. and Dingwoke, J.E. 2015. Larvicidal activity of *Chromolaena odorata* leaf extracts against *Aedes vittatus* mosquito. *International Journal of Applied Research and Studies*, 4(2): 1-6. | Amer, A. and Mehlhorn, H. 2006. Larvicidal effects of various essential oils against *Aedes*, *Anopheles*, and *Culex* larvae (Diptera, Culicidae). *Parasitology Research*, 99:466-472. | Arivoli, S., Narendran T. and Ignacimuthu. S. 1999. Larvicidal activity of some botanicals against *Culex quinquefasciatus* Say. *Journal of Advanced Zoology*, 20(2):19-23. | Arivoli, S., Ravindran, K.J., Raveen, R. and Samuel, T. 2012a. Larvicidal activity of botanicals against the filarial vector *Culex quinquefasciatus* Say (Diptera: Culicidae). *International Journal of Research in Zoology*, 2(1):13-17. | Arivoli, S., Ravindran, K.J. and Samuel, T. 2012b. Larvicidal efficacy of plant extracts against the malarial vector *Anopheles stephensi* Liston (Diptera: Culicidae). *World Journal of Medical Sciences*, 7(2):77-80. | Arivoli, S., Raveen, R., Samuel, T. and Sakthivadivel, M. 2015. Adult emergence inhibition activity of *Cleistanthus collinus* (Roxb.) Euphorbiaceae leaf extracts against *Aedes aegypti* (L.), *Anopheles stephensi* Liston and *Culex quinquefasciatus* Schrad. (Diptera: Culicidae). *International Journal of Mosquito Research*, 2(1): 24-28. | Arivoli, S. and Samuel, T. 2011. Bioefficacy of *Citrullus colocynthis* (L.) Schrad. (Cucurbitaceae) whole plant extracts against *Anopheles stephensi*, *Aedes aegypti* and *Culex quinquefasciatus* (Diptera: Culicidae). *International Journal of Current Research*, 3(4): 296-304. | Bagavan, A., Rahuman, A.A., Kamaraj, C. and Geetha, K., 2008. Larvicidal activity of saponin from *Achyranthes aspera* against *Aedes aegypti* and *Culex quinquefasciatus* (Diptera: Culicidae). *Parasitology Research*, 103(1): 223-229. | Basheer, A.G.M. 2014. *Ricinus communis* (Castor) as larvicide on *Anopheles arabiensis* Patton. *International Journal of Advances in Pharmacy, Biology and Chemistry*, 3(2): 319-328. | Belsam, M., Zohra, M., Ehsen, H., Manel, T., Abderrahman, B., Mahjoub, A. and Nadia, F. 2011. Anti-inflammatory evaluation of immature fruit and seed aqueous extracts from several populations of Tunisian *Citrullus colocynthis* Schrad. *African Journal of Biotechnology*, 10(20):4217-4225. | Brown, A.W.A. 1986. Insecticide resistance in mosquitoes: pragmatic review. *Journal of the American Mosquito Control Association*, 2: 123-140. | Cecilia, K.F., Ravindhran, R., Gandhi, M.R., Reegan, A.D., Balakrishna, K. and Ignacimuthu, S. (2014) Larvicidal and pupicidal activities of ecbolin A and ecbolin B isolated from *Echobolium viride* (Forssk.) Alston against *Culex quinquefasciatus* Say (Diptera: Culicidae). *Parasitology Research*, 113:3477-3484. | Chaib, K.F. 2004. Investigation phytochimique d'une brosse à dents africaine *Zanthoxylum zanthoxyloides* (Lam.) Zepernick et Timleryns. *Fagarazanthoxyloides* L. (Rutaceae)- Thèse de doctorat. Lausanne : Université de Lausanne, p. 211. | Chaturvedi, M., Mali, P.C. and Ansari, A.S. 2003. Induction of reversible antiferility with a crude ethanol extracts of *Citrullus colocynthis* Schrad fruit in male rats. *Pharmacology*, 68(1):38-48. | Chintem, D.G.W., Nzelibe, H.C., James, D.B., Albaba, S.U. and Grillo, H.T. 2014. Larvicidal potential of leaf extracts and purified fraction of *Datura stramonium* against *Culex quinquefasciatus* mosquitoes. *International Journal of Natural Sciences Research*, 2(12): 284-293. | da Silva, H.H.G., Geris, R., Filho, E.R., Rocha, C. and da Silva, I.G. 2007. Larvicidal activity of oil-resin fractions from the Brazilian medicinal plant *Copaifera reticulata* Ducke (Leguminosae- Caesalpinoideae) against *Aedes aegypti* (Diptera: Culicidae). *Revista da Sociedade Brasileira de Medicina Tropical*, 40(3):264-267. | Dhanotia, R., Chauhan, N.S., Saraf, D.K. and Dixit, V.K. 2011. Effect of *Citrullus colocynthis* Schrad fruits on testosterone-induced alopecia. *Natural Product Research*, 25(15):1432-1443. | Duke. 2006. *Phytochemical and Ethnobotanical Databases, Ethnobotanical uses of Citrullus* (Cucurbitaceae). <http://www.ars-grin.gov/cgi-bin/duke/ethnobot>. | Eze, E.A., Danga, S.P.Y. and Okoye, F.B.C. 2014. Larvicidal activity of the leaf extracts of *Spondias mombin* Linn. (Anacardiaceae) from various solvents against malarial, dengue and filarial vector mosquitoes (Diptera: Culicidae). *Journal of Vector Borne Diseases*, 51: 300-306. | Ghosh, A., Chowdhury, N. and Chandra, G. 2012. Plant extracts as potential larvicides. *Indian Journal of Medical Research*, 135:581-598. | Gleiser, R.M. and Zygadlo, J.A. 2007. Insecticidal properties of essential oils from *Lippia turbinata* and *Lippia polystachya* (Verbenaceae) against *Culex quinquefasciatus* (Diptera: Culicidae). *Parasitology Research*, 101: 1349-1354. | Gurudeeban, S., Satyavani, K. and Ramanathan, T. 2010. Bitter Apple (*Citrullus colocynthis*): An overview of chemical composition and biomedical potentials. *Asian Journal of Plant Sciences*, 1: 1-8. | Hatim, N.A.R., Whiting, D.A. and Yousif, N.J. 1989. Cucurbitacin glycosides from *Citrullus colocynthis*. *Phytochemistry*, 24(4): 1268-1271. | Huseini, H.F., Darvishzadeh, F., Heshmat, R., Jafarizadeh, Z., Raza, M. and Larjani, B. 2009. The clinical investigation of *Citrullus colocynthis* (L.) Schrad fruit in treatment of Type II diabetic patients: a randomized, double blind, placebo controlled clinical trial. *Phytotherapy Research*, 23(8):1186-1189. | Killeen, G.F., Fillingier, U. and Knols, B.G. 2002. Advantages of larval control for African malaria vectors: low mobility and behavioural responsiveness of immature mosquito stages allow high effective coverage. *Malaria Journal*, 1:8. | Krishnappa, K., Elumalai, K., Dhanasekaran, S. and Gokulkrishnan, J. 2012. Larvicidal and repellent properties of *Adansonia digitata* against medically important human malarial vector mosquito *Anopheles stephensi* (Diptera: Culicidae). *Journal of Vector Borne Diseases*, 49: 86-90. | Mahmoud, A., Mohammad, D., Fahaid, A., Nabil, B., Mohammad, A., Riyadh, E., and Mohammad, K., 2009. *In vivo*, acute, normo-hypoglycemic, antihyperglycemic, insulinotropic actions of orally administered ethanol extract of *Citrullus colocynthis* (L.) Schrad. *American Journal of Biochemistry and Biotechnology* 5(3): 118-125. | Marzouk, B., Marzouk, Z., Fenina, N., Bouraoui, A. and Aouni, M. 2011. Anti-inflammatory and analgesic activities of Tunisian *Schrad*. *American Schrad*. Immature fruit and seed organic extracts, *European Review for Medical and Pharmacological Sciences*, 15(6):665-672. | Marzouk, B., Marzouk, Z., Haloui, E., Fenina, N., Bouraoui, A. and Aouni, M. 2010. Screening of analgesic and anti-inflammatory activities of *Citrullus colocynthis* from southern Tunisia. *Journal of Ethnopharmacology*, 128(1):15-19. | Nayab, D., Ali, D., Arshad, N., Malik, A., Choudhary, M. and Ahmed, Z. 2006. Cucurbitacin glycoside from *Citrullus colocynthis*. *Natural Product Research*, 20(5): 409-413. | Nzelibe, H.C. and Chintem, D.G.W. 2013. Larvicidal potential of leaf extracts and purified fraction of *Cimicum gratissimum* against *Citrullus colocynthis* mosquito larva. *International Journal of Science and Research*, 4(2): 2254-2258. | Oliveira, P.V., Ferreira, J.C. Jr, Moura, F.S., Lima, G.S., de Oliveira, F.M., Oliveira, P.E.S., Conserva, L.M., Giulietti, A.M. and Lemos, R.P.L. 2010. Larvicidal activity of 94 extracts from ten plant species of northeastern of Brazil against *Aedes aegypti* L. (Diptera: Culicidae). *Parasitology Research*, 107:403-407. | Prabuseenivasan, S., Jayakumar, M., Raja, N., and Ignacimuthu, S. 2004. Effect of bitter apple, *Citrullus colocynthis* (L.) Schrad seed extracts against pulse beetle, *Callosobruchus maculatus* Fab. (Coleoptera: Bruchidae). *Entomology*, 29: 81-84. | Pravin, B., Tushar, D., Vijay, P. and Kishanchand, K. 2013. Review on *Citrullus colocynthis*. *International Journal of Research in Pharmacy and Chemistry*, 3(1): 46-53. | Rahbar, A.R. and Nabipour, I. 2010. The hypolipidemic effect of *Citrullus colocynthis* on patients with hyperlipidemia. *Pakistan Journal of Biological Sciences*, 13(24):1202-1207. | Rahuman, A.A., Venkatesan, P., and Gopalakrishnan, G. 2008. Mosquito larvicidal activity of oleic and linoleic acids isolated from *Citrullus colocynthis* (L.) Schrad. *Parasitology Research*, 103: 1383-1390. | Rasool, K. and Jahanbakhsh, T. 2011. Anticandidal screening and antibacterial of *Citrullus colocynthis* in South East of Iran. *Journal of Horticulture and Forestry*, 3(13): 392-398. | Raveen, R., Dhayanithi, P., Dhinamala, K., Arivoli, S. and Samuel, T. 2012. Larvicidal activity of *Pedilanthusthymaloides* (L.) Poit (Euphorbiaceae) leaf against the dengue vector *Aedes aegypti* (L.) (Diptera: Culicidae). *International Journal of Environmental Biology*, 2(2): 36-40. | Raveen, R., Kamakshi, K.T., Deepa, M., Arivoli, S. and Samuel, T. 2014. Larvicidal activity of *Nerium oleander* L. (Apocynaceae) flower extracts against *Culex quinquefasciatus* Say (Diptera: Culicidae). *International Journal of Mosquito Research*, 1(1): 36-40. | Saba, A.B. and Oridupa, A.O. 2010. Search for a novel antioxidant, anti-inflammatory/analgesic or anti-proliferative drug: Cucurbitacins hold the ace. *Journal of Medicinal Plants Research*, 4(25): 2821-2826. | Sakthivadivel, M. and Daniel, T. 2008. Evaluation of certain insecticidal plants for the control of vector mosquitoes, viz., *Culex quinquefasciatus*, *Anopheles stephensi* and *Aedes aegypti*. *Applied Entomology and Zoology*, 43(1):57-63. | Samuel, T., Ravindran, K.J. and Arivoli, S. 2012a. Bioefficacy of botanical insecticides against the dengue and chikungunya vector *Aedes aegypti* (L.) (Diptera: Culicidae). *Asian Pacific Journal of Tropical Biomedicine*, 2:S1842-S1844. | Samuel, T., Ravindran, K.J. and Arivoli, S. 2012b. Screening of twenty five plant extracts for larvicidal activity against *Culex quinquefasciatus* Say (Diptera: Culicidae). *Asian Pacific Journal of Tropical Biomedicine*, 2:S1130-S1134. | Samuel, T. and William, S.J. 2014. Potentiality of botanicals in sustainable control of mosquitoes (Diptera: Culicidae). In: *Achieving Sustainable Development: Our Vision and Mission*, Ed. William, S.J. Loyola College, Chennai, Tamil Nadu, India, p 204-227. | Satti, A.A. and Edriss, A.E. 2015. Preliminary phytochemical screening and activities of *Citrullus colocynthis* (L.) Schrad. as mosquito larvicides. *World Journal of Pharmaceutical Research*, 4(1): 188-203. | Shaalan, E.A.S., Canyon, D., Younes, M.W.F., Abdel-Wahaba, H. and Mansoura, A.H. 2005. A review of botanical phytochemicals with mosquitocidal potential. *Environment International*, 3: 1149-1166. | Shah, A.H., Mahia, A.M., Ageel, M. and Qureshi, S. 1989. Cytological studies on some plants used in Traditional Arab medicine. *Fitoterapia*, 60(2): 171-173. | SPSS. 2007. SPSS for windows, Version 11.5. SPSS, Chicago, Illinois, USA. | Sukumar, K., Perich, M.J. and Boobar, R. 1991. Botanical derivatives in mosquito control: A review. *Journal of the American Mosquito Control Association*, 7(2):210-237. | Thomas, J., Govindan, M.S. and Kurup G.M. 2014. Isolation and characterisation of mosquito larvicidal compound from *Gliricidia sepium* Jacq. *International Journal of Pharma Research and Health Sciences*, 2(2): 173-178. | Tomass, Z., Hadis, M., Teye, A., Mekonnen, Y. and Petros, B. 2011. Larvicidal effects of *Jatropha curcas* L. against *Anopheles arabiensis* (Diptera: Culicidae). *MEJS*, 3(1):52-64. | Torkey, H.M., Abou-Yousef, H.M., Abdel, A.A.Z., and Hoda, E.A.F. 2009. Insecticidal effect of Cucurbitacin E glycoside isolated from *Citrullus colocynthis* against *Aphis craccivora*. *Australian Journal of Basic and Applied Sciences*, 3(4): 4060-4066. | Vargas, M.V. 2012. An update on published literature (period 1992-2010) and botanical categories on plant essential oils with effects on mosquitoes (Diptera: Culicidae). *Boletín de Malariología y Salud Ambiental*, 2(2): 143-193. | W.H.O. 2005. Guidelines for laboratory and field testing of mosquito larvicides. Geneva. | W.H.O. 2014. A global brief on vector-borne diseases. WHO/DCO/WHO/2014.1. | Yoshikawa, M., Morikawa, M., Kobayashi, T., Kobayashi, H., Nakamura, A., Matsuhira, K., Nakamura, S. and Matsuda, H. 2007. Bioactive saponins and glycoside. XXVII. Structure of new Cucurbitane - type triterpene glycoside and anti-allergic constituents from *Citrullus colocynthis*. *Chemical and Pharmaceutical Bulletin*, 55(3): 428-434. | Zhou, H.Y., Zhao, N.N., Du, S.S., Yang, K., Wang, C.F., Liu, Z.L. and Qiao, Y.J. 2012. Insecticidal activity of the essential oil of *Lonicera japonica* flower buds and its main constituent compounds against two grain storage insects. *Journal of Medicinal Plants Research*, 6(5): 912-917.