

Protozoan Parasites of Some Grasshoppers of Manipur, India.

KEYWORDS

Manipur, Grasshopper, Gregarine, Species.

Indira Yumnamand N. Mohilal

Parasitology Section, Department of Life Sciences, Manipur University, Canchipur

ABSTRACT Protozoan parasites infest a wide range of Arthropods including Insects like Grasshoppers. During the course of a survey in Manipur, Grasshopper were collected and examined for protozoan parasites. Gre-garines were found from the mid-gut region. The taxonomy of Gregarine Species are based on the trophozoite morphology, epimerite morphology, timing of gamont association, gametocyst morphology and the method of gametocyst dehiscence (Clopton 2009; Léger 1892; Smith and Cook, 2008). In the present Communication five species of Septate gregarine trophozoites belonging to Brustiospora, Lepismatophila, Retractocephalus, Nematopsis and Amphiplatyspora genera are reported from three species of grasshoppers found in Manipur. The morphological details supported with photomicrographs are provided in the manuscript.

Introduction

Protozoa are a diverse and poorly understood group of unicellular organisms. Among them apicomplexans parasites inhabit the intestines, coeloms and reproductive vesicles of marine, freshwater and terrestrial invertebrates. Of 6,000 described apicomplexan species, 1,600 are gregarines and many more are estimated to exist (Adl et al.2007; Hausmann, Hullsmann, Levine 1988; Morrison, 2009; Perkins et al. 2000). The description of gregarine species has historically been based on the morphological species concept that emphasized features like trophozoite morphology, epimerite morphology (mainly septate eugregarines), timing of gamont association, gametocyst morphology, and the method of gametocyst dehiscence (Clopton, 2009;Le'ger, 1892; Smith and Cook, 2008). Host association has also been used as a taxonomic criterion because many gregarines are inferred to be stenoxenous, being able to mature in only a single host genus or species Levine, 1979; Perkins et al 2000). Nonetheless, some gregarine species can infect different host species, and one host species can be infected with several different gregarine species (A`bro, 1974; Rueckert and Leander, 2009). Moreover, the shape, size, and ultrastructure of trophozoites are often highly variable depending on different developmental stages of the parasite and host environmental conditions. In the present Communication five species of Septate gregarine are reported.

Material and Methods

The Adults of Oxya hyla hyla, Chondracis rosea and Choreodocus robustawere collected from various grass fieldsof Manipur (24° 44' N,93° 58' E) in the morning between 6 to 8 a.m. with the help of net,were kept in glass tubes and brought alive to the laboratory for investigation from April, 2011 to December, 2014. These were decapitated, their guts carefully dissectedout under a dissecting microscope and gently pressed to expel theparasites from the gut lumen. Thin smear preparations were fixedin Schaudinn's fixative and subsequently stained with Heidenhain'shaematoxylin (Kudo, 1966). Gametocysts were recovered from thehind gut and placed in moist chambers (> 80% relative humidity)for sporulation (Sprague, 1941). The structure of the oocystswere studied by using Lugol's iodine solution. Figures of stained specimens were drawn with the aid of a camera lucida. Measurements of fresh

materials were taken using an ocular micrometer calibrated with a stage micrometer. All measurements, unless otherwise mentioned, are in micrometers. Forty specimens each of mature gamonts and associations were randomly measured from the infected hosts. Similarly, thirty gametocysts and fifty individual oocysts were measured. Measurements were taken from widest part of protomerite, deutomerite, nucleus, gametocyst and oocyst and are presented in this paper as range values, followed by means, standard errors and sample sizes in parentheses. Blue filters were used for measurements and daylight filters were used for observation of colour in living specimens. Nomenclature for shapes used in this manuscript conforms to (Clopton, 2004).

Results and discussion

Several motile trophozoites were observed in the midgut of infectedOxya hyla hyla, Chondracis rosea andChoreodocus robusta. The specimens show five forms of trophozoites i.e. Trophozoites of Brustiosporaindicola, Lepismatophila cruszi, Retractocephalus aulacophorae, Nematopis annulipes and Amphiplatyspora striata as shown in Figure 1. Plate 1.

Brustiosporaindicola Kundu and Haldar, 1981 Measurement: Table.1.

Description: The trophozoite were encountered from within the lumen of the host's midgut. The body is elongated and measures 68.2µm-146.7µm (121.6). The protomerite is more or less cylindrical and it measures 6.52µm-27.9µm \times 9.0µm-29.7µm (20.7). The deutomerite is elongated and measures 36.7µm-118.8µm (98.4) × 11.7µm-42.5µm (20.4), gradually narrows to a rounded posterior extremity. The pellicle is very thin and the cytoplasm is uniformly granulated. The nucleus is almost spherical, situated just behind the protomerite - deutomerite septum and measures 6.2µm in diameter.

Host: Oxya hyla hyla Habitat: Mid gut and hepatic caeca

Incidence: 3 out of 7 are infected

Lepismatophila cruszi Kundu and Haldar, 1984 Measurement: Table.1.

Description: The detailed structures of a trophozoite measures 35.9μ m-149.0 μ m (82.8). The body as usual bears epimerite, prortomerite and deutomerite. The epimerite is petalloid or has two horned structure that pushes through the host epithelium. The protomerite is sub-conical or hemispherical in shape, measures 6.4μ m-24.0 μ m (15.1) × 49.9 μ m-11.7 μ m (28.1). It is followed by the deutomerite and measures 125 μ m-45 μ m (69.5) × 15.0 μ m-54 μ m (30.2). It is highly granulated and is separated from the protomerite by a clear septa. The nucleus is circular and measures 12.7 μ m in diameter, lodged immediately behind the septum, possesses a distinct nuclear membrane and encloses a big endosome and few chromatin granules in it.

Host: *Chondracis rosea* Habitat: Hepatic caeca Incidence: 4 out of 8 are infected

Retractocephalus aulacophorae Haldar, Chakraborty and Kundu, 1982

Measurement: Table.1.

Description: The intracellular form of the parasite is somewhat oval in outline. Its cytoplasm appears opaque white when observed in fresh smear preparation. It may reach upto a length 139 μ m -309 μ m (154.1). The protomerite is somewhat conical in shape and is pointed towards the apex and measures 10.9 μ m-65.0 μ m × 26.0 μ m-97 μ m (39.9). The deutomerite is the largest segment of the body and is separated from the protomerite by a deep constriction. The deutomerite is obese in shape, and is broadest slightly behind the septum, measure 123 μ m-244 μ m (150.7) × 18.7 μ m-139 μ m (43.6). It tapers posteriorly and ends in a rounded proximity. The nucleus is spherical, situated almost at the center of the deutomerite and measures 7.5 μ m in diameter.

Host: *Oxya hyla hyla* Habitat: Mid gut and hepatic caeca Incidence: 5 out of 13 are infected

Nematopsis annulipes Prasadan and Janardanan, 2001 Measurement: Table.1.

Description: Development extracellular, smallest observed trophozoite, narrow, elongate and ovoid. It may reach up to 198.7 μ m-315 μ m (272.7). Protomerite shape subspherical to ovoid, measures 29.7 μ m-64 μ m (38.4) × 11.7 μ m-48 μ m (29.7). Epicyte uniformly thick and striated. Endocyte granulated. Septa circular, convex towards deutomerite. Deutomerite narrow behind septum, gradually dilates, posterior end broadly round or flat and measures 11.9 μ m-252 μ m (127.7) × 33.2 μ m-72 μ m (49.4); nucleus spherical, variable in position and measures 7.5 μ m in diameter.

Host: *Choreodocus robusta* Habitat: Mid gut Incidence: 6 out of 21 are infected.

Amphiplatyspora striata Kundu and Haldar, 1984 Measurement: Table.1.

Description: The trophozoite encountered in the smear preparation is cylindrical in shape, with a large ovoidal protomerite and elongate deutomerite. The trophozoite varies markedly in shape and size in both protomerite and deutomerite. It measures 110μ m- 411.9μ m (255.5). The trophozoite has a globular, fusiform, rectangular, semi-lunar or sickle-shaped, hemispherical or a hat shaped, Protomerite measuring 22.9μ m- 79.7μ m (41.9) × 29.7μ m- 158.9μ m (79.1). The deutomerite is cylindrical flask shaped, vermiform, cylindro -conical with broadly rounded anterior and gradually tapering posterior extremity and pitcher shaped body measuring 79.1μ m- 332.2μ m (220.7) × 29.8μ m- 169μ m (89.7). The nucleus is situated at the deutomerite portion and measure 17.5μ m in diameter. The longitudinal striation converge at a point near the posterior tip of the body. The various shape of the protomerite and deutomerite are probably due to the presence of such strong striations in the body.

Host: *Chondracis rosea* Habitat: Mid gut and hepatic caeca Incidence: 4 out of 18 are infected

In the course of the present investigation 20*Oxya hyla hyla*, 26 *Chondracis rosea* and 21 *Choreodocus robust*awere examined of which 8, 8 and 6 respectively were infectedby eugregarines. The present findings showing a number of septate gregarines from the Grasshoppers of Manipur, is an added knowledge to the study of the biodiversity of protozoan groups.

The present specimens of *Brustiospora indicola* shows similarity in the size and shape of Protomerite, Deutomerite, Gametocyst, Spore, with the specimens of (Kundu and Haldar, 1981). But there is slight variation in the total length of the Trophozoite. The present specimen is also reported from a new host i.e. *Oxya hyla hyla*. The host are obtained specially in the months of September to November and the infection is carried mostly during these months. At an average 42% are infected by the parasite.

On the other hand Lepismatophila Cruszi was originally described by Kundu and Haldar in 1984 from Silver fish of West Bengal. In the present work it has been described from Chondracis rosea. The morphological details and measurements are within the range described in the original work. 50% of the host examined were infected with the parasite. Retractocephalus aulocophorae is another gregarine that is prevalent in grasshoppers. In the original description of Haldar, (Chakraborty and Kundu, 1982) it has been described from the midgut of the Beetle, Lema sp. The present specimen is described from Oxya hyla hyla. The Trophozoite is a little smaller in the present specimen. The ratio of LP:TL and WP:WD show much similarity. In the presence of intracellular developmental phase, Simple globular epimerite and barrel shaped spore they are much similar. The parasite availability coincides with the abundance of the host i.e. during the months of June to September, 39 % of the hosts are infected with the parasite during this period. Another gregarine Nematopsis annulipes has been recovered from Choreodocus robusta while the original specimens of (Prasadan and Janardanan, 2001) were from the Crustacean Uca annulipes. The prevalence rate in the present case is much higher than the original. In addition the size range of the present specimen are much higher. In other morphological details like those of Sporadins, Epimerite, Protomerites, presence of satellites, structure of Gametocyts there are close similarities. From Chondracis rosea another gregarine Amphiplatyspora striata had been recovered. (Kundu and Haldar, 1984) described the species from an Orthopteran Insect, Pteronemobius concolor. Comparison of the morphological characteristics and measurements show close resemblance

RESEARCH PAPER

Volume : 5 | Issue : 3 | March 2015 | ISSN - 2249-555X

withthe present specimens. The intensity of infection is low ranging around 27% which is close to 22% in case of the original specimen.



Figure 1. Photomicrographs of the different septate gregarines A-Trophozoite of Bruzitozpora indicola, B-Trophozoite of Lapinmaphila cruzit, C- Trophozoite of Raractocephalus aulacophorae, D- Trophozoite of Nomalopis amultaes, E- Trophozoite of Amphipalapspora striata

Table: 1	١.	Measurements	of	Five	Species	of	Septate	gre-
garines								

Charac- ters	Brustio- sporaindi- cola, Kundu and Haldar, 1981	Lepis- matoph- ila cruszi, Kundu and Haldar, 1984	Retracto- cephalus aulacopho- rae, Haldar et al. 1982	Nematop- sis an- nulipes, Parasa- dan and Janardan, 2001	Amphi- platys- pora striata, Kundu and Haldar, 1984
Total length	68.2μm- 146.7μm	35.9μm- 149.0μm	139.0µm- 309µm	198.7µm- 315µm	110µт- 411.9µт
Protomer- ite	More or less Cylin- drical	Sub- Conical or hemi- spherical	Somewhat Conical	Subspheri- cal to Ovoidal	Sickle shaped, hemispheri- cal or hat shaped
Deu- tomerite	Elongated and gradu- ally narrows to posterior extremity	-	Obese, distinct epicyteal striations	Rounded to flat	Cylindrical flask Shaped
Nucleus	Spherical	Circular	Spherical, one or two Karyosomes	Spherical	Spherical to semi lunar

Sporadin	Typically Solitary	Solitary	Biassocia- tive	Asso- ciation of three Sporadin	Solitary
Gameto- cyst	Elliptical or egg shaped	Bean- Shaped with one side convex	Spherical	Spherical to Ovoid	Ovoidal with prominent ectocyst: dehisces by simple rupture
Oocyst	Spherical	Boat- Shaped	Barrel- Shaped	Spherical uninucle- ated	Spore cylindrical with polar thickenings: extruded in chains
LP:TL	1:5.0	1:6.2	1:4.7	1:4.9	1:5.2
WP:WD	1:1.0	1:1.7	1:1.3	1:1.5	1:0.9
Host	Oxya hyla hyla	Chondracis rosea	Oxya hyla hyla	Choreo- docus robusta	Chondracis rosea
Locality	Manipur, India	Manipur, India	Manipur, India	Manipur, India	Manipur, India

REFERENCE A'bro, A. 1974. The gregarine infection in different species of Odonata from the same habitat. Zool. Scripta, 3:111–120.IAdl, S. M., Leander, B. S., Simpson, A. G. B., Archibald, J. M., Anderson, O. R., Bass, D., Bowser, S. S., Brugerolle, G., Farmer, M. A., Karpov, S., Kolisko, M., Lane, C. E., Lodge, D. J., Mann, D. G., Meisterfeld, F., Mendoza, L., Moestrup, Ø., Mozley-Standridge, S. E., Smirnov, A. V. & Spiegel, F. 2007. Diversity, nomenclature, and taxonomy of protists. Syst. Biol., 56:684–689.IClopton RE 2004. Standard nomenclature and metrics of plane shapes for use in gregarine taxanomy. Comperative Parasitology; 71:130-40.IClopton, R. E. 2009. Phylogenetic relationships, evolution, and systematic revision of the septate gregarines (Apicomplexa: Eugragrinorida: Septatorina). Comp. Parasitol., 76:167–190.IHausmann, K., Hu'Ismann, N. & Radek, R. 2003. Protistology, 3rd completelyrevised ed. E. Schweizerbart/sche Buchhandlung (Na"gele u. Obermiller), Stuttgart.IKundu 1966. Studies on Nyctanthes arbor-tristis Linn. Proc. Indian Sci, Congr Assn. Part III. Candigarh.ILe'ger, L. 1892. Recherches sur les Gre'garines. Tabl. Zool., 3:1–183.ILevine, N. D. 1979. New genera and higher taxa of septate gregarines (Protozoa, Apicomplexa). Protozool., 26:532–536. ILevine, N. D. 1988. Progress in taxonomy of the apicomplexan protozoa. J. Protozool., 35:518–520. IMorrison, D. A. 2009. Evolution of the Apicomplexa: where are we now? Trends Paragites in taxoning of the apicomplexa. In TotoCoro, 33.10–32.00. How Strends Paragites and the approximation of the apicomplexa in the apicompl particular reference to the chromosome cycle III. Biol. Mongor.18: 5 - 57.