



Effect of α -Cyclodextrin on Gefitinib

KEYWORDS

α -Cyclodextrin, Gefitinib, Absorption, Emission, Phase solubility

A.GloryPunitha

Dr.J.PremaKumari

Department of Chemistry, Scott Christian College, (Autonomous).Nagercoil.TamilNadu, India

Department of Chemistry, Scott Christian College, (Autonomous).Nagercoil.TamilNadu, India

ABSTRACT Gefitinib is a drug used in the treatment of metastatic epithelial cell lung cancer. This Study examined the complexation of Gefitinib with α -Cyclodextrin in liquid State. The inclusion processes are discussed based on absorption, emission and phase solubility studies. The absorption and emission maxima of gefitinib appear at 263nm and 402nm. The stability constant [Kst] of Gefitinib is calculated. The formation constant [K] value is calculated by analysing in the intensities of absorption and emission maxima with the α -CD concentration.

Introduction

Gefitinib is a drug used for the treatment of epithelial cell lung cancer. It is taken in 250mg oral doses once daily, higher doses showing no improvement but increased toxicity. It is an anilinoquinazoline with chemical name N-[3-chloro-4-fluoro-phenyl-7-methoxy-6-[3-morpholin-4-yl propoxy] quinazoline 4-amine. It has the molecular formula $C_{22}H_{24}ClFN_4O_3$, a relative molecular mass of 446.9⁽¹⁾

Gefitinib is the first selective inhibitor of epidermal growth factor receptor's [EGFR] tyrosine kinase domain. Thus gefitinib is an EGFR inhibitor^(2,3)

Cyclodextrins

Cyclodextrins (CD) are a group of structurally related cyclic oligosaccharides that have a polar cavity and hydrophilic external surface. The most commonly used host molecules are cyclodextrins⁽⁴⁾. Cyclodextrins are cyclic oligosaccharides of 6, 7 or 8 -D-glucopyranose units with alternatively hydrophobic central cavity and hydrophilic outer surface^(5,6). The hydrophobic CDs innercavity forms inclusion complexes with a wide range of guest molecules^(7,8) while the hydrophilic exterior enhances CD solubility in water. CDs are classical examples of compounds that can form inclusion complexes⁽⁹⁾

Experimental

Apparatus

- Double beam spectrophotometer-2203
- Jascospectrofluorometer FP-8200
- Rotary shaker

Procedure for preparation of liquid inclusion complex of Gefitinib with α -CD

The solutions of the stock of gefitinib was transferred into 10ml volumetric flasks containing 0.002, 0.004, 0.006, 0.008, and 0.01 mol dm⁻³ α -CD solution. The mixed solution was diluted to 10ml with double distilled water and shaken thoroughly. The absorption and fluorescence spectra were recorded.

Phase solubility studies:

Phase solubility studies were performed according to the method reported by Higuchi and Connors⁽¹⁰⁾. Guest compound in constant amounts that exceeded its solubility was transferred to screw capped vials containing

15ml of aqueous solution of α -CD. The contents were stirred on rotary shaker for 72hrs at 37°C. The time duration was fixed based on pilot experiment and found to be sufficient to achieve equilibrium of mixture. After reaching equilibrium samples were filtered through the Whatmann No.1 filter paper and analysed by UV-Visible Spectrophotometer. Solubility studies were performed in triplicate.

Results and Discussion

Table:1

Absorption and Fluorescence maxima of Gefitinib at different concentration of α -CD

α -CD con	λ max	Absorbance	λ flu	Intensity	1/(α CD)	Log ϵ
0	263.0	0.392	402	102.275		4.00
0.002	262.0	0.443	404	106.316	500	4.05
0.004	261.0	0.543	405	107.632	250	4.14
0.006	260.5	0.601	407	109.820	166.66	4.18
0.008	259.5	0.633	409	110.654	125	4.21
0.01	259	0.705	411	111.542	100	4.25

Table (1) and figure (1,2) shows the absorption and fluorescence maxima of Gefitinib solutions containing various concentrations of α -CD. The absorption maxima of gefitinib appear at 263.0nm. The absorption intensities are increased with the increasing concentration of α -CD. As the concentration of α -CD increases the absorption wavelength is blue shifted from 263nm-259nm. It is already reported that the anions are blue shifted in the α -CD medium⁽¹¹⁾. Here because of the presence of halide ions the absorption maxima is blue shifted. The fluorescence maxima of gefitinib appear at 402nm. Upon increasing the concentration of α -CD it is shifted from 402nm to 411nm. By the addition of α -CD the fluorescence and their intensities are red shifted.

Figure.1
Absorption spectra of Gefitinib at different concentration of α -CD

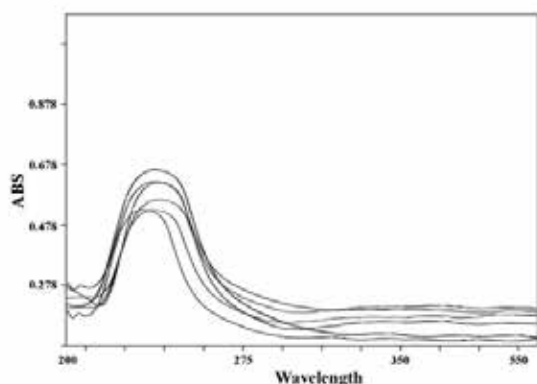


Figure.2
Fluorescence spectra of Gefitinib at different concentration of α -CD

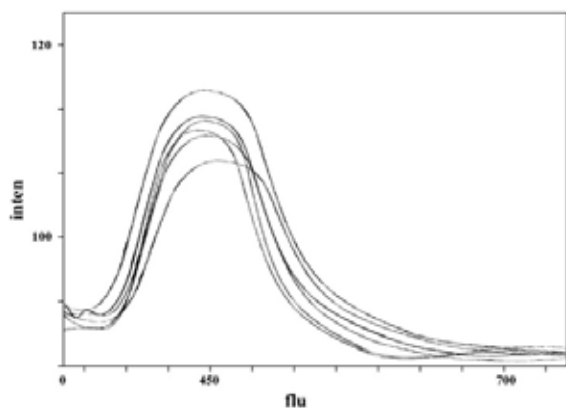
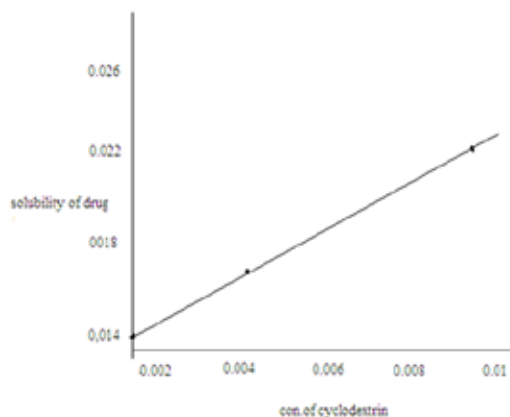


Figure.3
Phase solubility of Gefitinib at different concentration of α -CD



The phase solubility diagram of gefitinib as a function of concentration of various CDs at room temperature is shown in fig.3. The solubility of gefitinib with an increase in concentration of CDs gives an AL type of phase solubility diagram. The stability constant (Kst) of the complex was calculated from the slope and intercept (S_0) of the phase solubility diagram according to the equation

$$K_{st} = \frac{\text{slope}}{S_0(1-\text{slope})}$$

The Kst value of gefitinib α -CD complex was calculated to be $87.30M^{-1}$. The Kst values of α -CD gefitinib complex make them suitable for practical applications in terms of improving the drug permeability solubility related oral bioavailability.

REFERENCE

1. Iressa label, U.S., (2007) Food and Drug Administration. Available at <http://www.fda.gov/cder/foi/label/2005/02139950081b/bdf> | 2. Pao, W., Miller, V., Zakowski, M., et al. (2004) "EGF receptor gene mutations are common in lung cancers from "never smokers" and are associated with sensitivity of tumors to gefitinib and erlotinib". Proceedings of the National Academy of Sciences of the United States of America 101 [36]113306-11 | 3. Sordella, R., Bell, D.W., Haber, D.A., Settleman, J. (2008) "Gefitinib-Sensitizing EGFR mutations in lung cancer activate anti-apoptotic pathways". Science 305 [5687] 1163-67. | 4. Loftsson, T., (2007) Cyclodextrins and the Biopharmaceutics Classification System. J. Inclusion Phenomena and Macrocyclization, 44, 63-67 | 5. Brewster, M.E. & Loftsson, T. (2007) Cyclodextrins as pharmaceutical solubilizers. Advanced Drug Delivery Reviews, 59, 645-666. | 6. Alvariza, C., Usero, R. and Mendicuti, F. (2010) Binding of dimethyl 2, 3-naphthalenedicarboxylate with α , β and γ cyclodextrins in aqueous solution. Spectrochimica Acta A, 67, 420-429. | 7. Calabró, M. L., Tommasini, S., Donato, P., Raneri, D., Stacanelli, R., Ficarra, P. (2011) Effects of α - and β -cyclodextrin complexation on the physico-chemical properties and antioxidant activity of some 3-hydroxyflavones. Journal of Pharmaceutical and Biomedical Analysis 35 (2), 365-377. | 8. Lucas-Abellán, C., Forte, I., López-Nicolás, J.M., Núñez-Delgado, E. (2009) Cyclodextrins as resveratrol carrier system. Food Chemistry, 104, 39-44. | 9. Szejtli, J., (2008) Cyclodextrins as food ingredients. Trends in Food Science and Technology, 15, 137-142 | 10. Higuchi, K., Connors, K.A., (1965) Phase Solubility Techniques. Adv in Analytical Chemistry, 4, 117-212 | 11. Prema Kumari, J., Allan Gnana Raj G., Rajendran, N., (2009) "Study on the spectral characteristics of 4-hydroxy-3-methoxy benzoic acid in different pH and α -Cyclodextrin" J. Indian Chem. Soc., 86, 53-57.