



Radiographical&Clinical Study by Applying Laser&Vegf on Bone Remodeling in Experimental Tooth Movement

KEYWORDS

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ABSTRACT *Aim of the Study* The main aim of this study:

To evaluate the effect of the Low Level Laser Therapy (LLLT) on the rate of post experimental tooth movement clinically, biologically and radiographically, and on the bone remodeling histologically and immunohistochemically. Radiographic evaluation of the effect of local application of LLLT and VEGF (alone and in combination) on orthodontic movement. The present study shows the LLLT Laser and VEGF exerts a fundamental role in remodeling periodontal ligament and is also involved in bone resorption and formation.

Introduction

Low Level Laser Therapy (LLLT) is a simple and inexpensive method that can be used easily in the dental practice for different purposes .The stimulatory effect of low-level laser therapy is well known and includes enhancement in tissue growth and tissue regeneration, resolution of inflammation (1.Chiari,2015) , pain reduction(2.Stein et al.,2015) ,enhancement of wound healing(3.Martins et al.,2015). Some studies investigated the efficacy of low power lasers in reducing burning mouth pain (4.Arbabi-Kalati et al.,2015) ,promoting bone regeneration in the midpalatal suture during expansion and stimulating tooth movement(5.Marquezan et al.,2013).

The vast majority of therapeutic lasers are semiconductor lasers today(6.Dalaie et al.,2015). There are three diode types:

1. Indium, Gallium-Aluminum-Phosphide (InGaAlP) laser
2. Gallium-Aluminum Arsenide (GaAlAs) semiconductor laser
3. Gallium-Arsenide (GaAs) semiconductor laser

In recent research projects, the effect of laser therapy was tested regarding the stimulatory effect on bone remodeling with the potential to influence the tooth movement rate as related to photo-biological responses of oral tissue after application of laser(7.Seifi &Vahid-Dastjerdi 2015).

There are three effects that commonly occur as a result of tissue exposure to laser photons (8.Torri &Weber ,2013). They are:

Primary effects of photoreception are a result of the interaction of photons and cell mitochondria which capture, direct, and transduce photon energy to chemical energy used to regulate cellular activity.

Secondary effects occur in the same cell in which photons

produced the primary effects and are induced by these primary effects. Secondary effects include cell proliferation, protein synthesis, degranulation, growth factor secretion, myofibroblast contraction and neurotransmitter modification—depending on the cell type and its sensitivity.

Tertiary effects are the indirect responses of distant cells to changes in other cells that have interacted directly with photons. They are the least predictable because they are dependent on both variable environmental factors and intercellular interactions.They are, however, the most clinically significant. Tertiary effects include all the systemic effects of phototherapy. Primary, secondary, and tertiary events summate to produce phototherapeutic activity.

Orthodontic tooth movement occurs in the presence of a mechanical stimuli sequenced by remodeling of the alveolar bone and periodontal ligament (PDL). Bone remodeling is a process of both bone resorption on the pressure site and bone formation on the tension site(9.Baloul, 2015). Orthodontic tooth movement can be controlled by the size of the applied force and the biological responses from the PDL .The force applied on the teeth will cause changes in the microenvironment around the PDL due to alterations of blood flow(10.McCormack et al.,2014), leading to the secretion of different inflammatory mediators such as cytokines, growth factors, neurotransmitters, colony-stimulating factors, and arachidonic acid metabolites (11.Kumar et al.,2015). As a result of these secretions, remodeling of the bone occurs .The most important growth factor is Vascular endothelial growth factor (VEGF) ,this protein is a member of the PDGF/VEGF growth factor family and encodes a protein that is often found as a disulfide linked homodimer. This protein is a glycosylated mitogen that specifically acts on endothelial cells and has various effects, including mediating increased vascular permeability, inducing angiogenesis, vasculogenesis and endothelial cell growth, promoting cell migration, and inhibiting apoptosis.VEGF is involved in tissue neo-formation that is strictly correlated with the presence of blood vessels(12.Wu. et al.,2009).

During orthodontic tooth movement, compressive forces induce angiogenesis of periodontal ligament together with the role of mediator of the VEGF. The localization of VEGF was analyzed in many in vivo researches and illustrated an increment in its expression in periodontal tissue during experimental tooth movement (13. Salomão et al., 2014). Therefore, VEGF exerts a fundamental role in remodeling periodontal ligament and is also involved in bone resorption and formation.

Material and Method

Orthodontic Appliance Design and LLL therapy

The orthodontic appliance consists of orthodontic bands, arch wires, and NiTi open-coil spring. The bands were customized for each rabbit. Briefly, the animals were anesthetized with general anesthesia, induced by an intramuscular injection of a ketamine (50 mg/ml) at a dose of 50mg/kg body weight and muscle relaxant Orbarcaine 2% at a dose of 5mg/kg body weight. The two drugs were mixed at the ratio of 2:1 (Ketamine: Orbarcaine), impression for mandibular central incisors (MCIs) of each rabbit was taken first with silicone material, study stone models of the MCIs and the surrounding region were made, which used for preparing of individual resin trays for each rabbit; that then used to take precise final impressions with alginate material and the master stone models. Orthodontic bands were prepared to fit the teeth sizes, using band strips (Dentaurum-Germany) and then welded under pressure by using of Welder device. Then a round buccal tube with wings welded to the hand made bands in a horizontal direction and used as labial tube. The bands were cemented to its co-related MCIs after the removing of the orthodontic elastic separator, so that the superior border of the cemented bands was 3mm away from the incisal edge to allow for wear of the teeth and the lower border about 2mm away from the cervical area to avoid a trauma of the surrounding tissue. Orthodontic tooth movement was generated by the insertion of a stainless steel arch wire with diameter of 0.016" and 15 mm in length through the labial tubes and the NiTi open-coil spring (ORTHO. TECHNOLOGY USA) with 3-4 mm in length (about 4-6 coils) was inserted along the arch wire from the non-bend end and subjected to constrict pressure with tucker in order to be inserted between the labial tubes, so that it will apply a pushing force on both MCIs (in distal direction) with a total orthodontic force of (100 gm.) divided into two teeth so that each incisor receive a light continuous force of (50gm) according to Proffit et al. (6). This force was measured by pressure-gauge (CORBLX, Dent arum -Germany). Two coils at both ends of the arch wire were made in one plan, and it serves as stopper for the arch wire and as non-traumatic end. Experimental tooth movement was conducted for 21 days. The experimental group

(A) Was received the LLL therapy at 810 nm, with an output of 250 mW

, and exposure of 20 s for each 7 days. While the experimental group

(B) was received the 0.1µm of VEGF in pressure side and about 0.2 mm subgingivally, for each 7 day.

Experimental group (C) treated with a combination of LLL therapy and 0.1µm VEGF and for each 7 days.

Radiographical Study

An intra-oral radiograph will be taken for the three groups

before and after application of LLLT and VEGF separately and in combination. Itan conventional Mono path size 2 ultra-speed type radiographic dental film.

Results

1 Results Of the Clinical Measurements

The data of the all experimental periods include the experimental tooth movement measurements (cumulative Measurements) and percentage, both of them were analyzed using both Descriptive and Inferential statistics as follows:

1.1 Measurements of distance (mm) between MCIs

A bodily tooth movement was observed in all groups. The means of the weekly measurements of the distance between the mandibular central incisors (MCIs) of each group and their comparisons among groups and within group are shown in **Table (1)**, and **(Figure 1)**. It was found that, there was gradual increase in the amount of tooth movement from the 1st week till the 3rd week as follow:

After 1 week: the mean ± SD values of separation were increased and the highest one was (0.79±0.01), in combination group, then (0.66±0.01 mm) in laser, then (0.34±0.02 mm) in VEGF, and the least one was (0.32±0.01 mm) in control.

After 2 week: the mean ± SD values of separation were increased and the highest one was (1.22 ±0.01), in combination group, then (1.08±0.04 mm) in laser, then (0.78 ±0.01 mm) in VEGF, and the least one was (0.51 ±0.01 mm) in control.

After 3 week: the mean ± SD values of separation were increased and the highest one was (1.95 ±0.11), in combination group, then (1.78 ±0.03 mm) in laser, then (1.23±0.01 mm) in VEGF, and the least one was (0.92±0.01 mm) in control.

Figure (2) illustrates Stem-Leaf Plots for clinical measurements of distance (mm) between MCIs among studied groups at the 3rd week.

By using Receiver Operating Characteristic Curve (ROC Curve) analysis was listed in table (2) & figure (3) that show area under the curve (ROC) for clinical measurements of distance (mm) between MCIs in contrast of control group at different periods, for 1 week the area (0.056) with sensitivity (100) and specificity (88.9), while for the 2 & 3 week they illustrate area (0.00) with sensitivity (100) and specificity (100).

Table (3) & figure (4) record (0.191) area under the curve (ROC) for distance (mm) between MCIs in contrast of control group with other groups.

Area under the curve (ROC) for clinical measurements of distance (mm) between MCIs in contrasts of all pair wise groups was reported in table (4) & figure (5). Wide area (0.66) was recorded for laser group pair wise VEGF group, with a high 95% Confidence Interval (LB 0.486-UB 0.847).

1.2 percentages of movement (P.M.)

Descriptive statistics for percentages of movement (P.M.) for each contrast's periods at different groups and Comparisons significant was recorded in table (5). High percentage recorded in **1w. & 3w.** and specifically to group VEGF (**261.76**).

The results that shows in table (6) illustrates that most pairs comparisons of contrasts of periods of (P.M.) factor are accounted highly significant at $P < 0.01$, except between (1w. & 2w. X 2w. & 3w.) at $P > 0.05$.

Figures (6,7) show a Photograph for Biopsies that show the difference in the distance that remained between MCIs after 3 weeks duration of the experiment.

1.3 Radiographic evaluation

At the end of the experiment period (at the end of 3rd week) all the study groups were subjected to x-ray to evaluate the distance that remained between MCIs as show in figure(8).

It was notice that, the radiographical measurement results of the distance that remained between MCIs at the end of experiment was coincide with the results of the clinical measurement at the same period, with statistically no significant difference ($P > 0.05$) between the two measurements of each group, as show in table(7) and figure(9).

1.4 The Width of the Mandibular Suture (Mand.S)

The highest mean values of the Mand.S width at three points that measured through radiographs at the end of the experiment was in combination group, mean± SD was(0.492±0.04mm), and in laser(0.455±0.01mm), while-VEGF records a low value (0.242 ±0.02mm) and control is the least one (0.215±0.01mm) ,as show in figure(10).

Table (3.8)&figure (3.11) illustrate the Area(0.056) under the curve (ROC) for amount of width mandibular suture (mm) in contrast of control group with sensitivity (100) and specificity (94.4).

1.5 The Width of the Periodontal Ligament (PDL):

The mean± SD values of the width of PDL (means of three points) were shown as follow

Figure (3.12)illustrates Stem-Leaf Plots for Width of PDL of Right and Left mesial side among studied groups. The highest values was recorded by combination group ,M±SD for the mesial right side was(0.285±0.01mm) and for left side(0.263±0.02mm).

Figure (3.13) shows Stem-Leaf Plots for Width of PDL of Right and Left distal side among studied groups. The highest values was recorded by combination and laser groups ,M±SD for the mesial right side was(0.222±0.02mm) that equal for both groups ,while for the left side ,laser group records a high value (0.232±0.01mm),followed by combination (0.227±0.01mm).

Table (9)and figure(14) show Area under the curve (ROC) for Width of PDL of Right and Left Mesial Side and Distal Side in contrast of control group at different intervals.

For right mesial side the area was (0.11 6)with sensitivity (100) and specificity (61.1),and for the left mesial side the area was (0.199) with sensitivity (100) and specificity (50).

For rightdistal side the area was (0.287)with sensitivity (100) and specificity (44.4),and for the left distal side the area was (0.199) with sensitivity (100) and specificity (55.6).

Clinical Measurements of distance (mm) between MCIs

Table 1: Descriptive statistics of Clinical Measurements of distance (mm) between MCIs among studied groups

at different intervals with comparisons significant

| Marker | Groups | Mean | SD | SE | ANOVA test | |
|--------|-------------|------|------|------|------------|------------------------|
| | | | | | F-test | P ^(*) Value |
| Week 1 | Control | 0.32 | 0.01 | 0.01 | 1290 | 0.000 HS |
| | Lazar | 0.66 | 0.01 | 0.00 | | |
| | VEGF | 0.34 | 0.02 | 0.01 | | |
| | Combination | 0.79 | 0.01 | 0.01 | | |
| Week 2 | Control | 0.51 | 0.01 | 0.00 | 1111 | 0.000 HS |
| | Lazar | 1.08 | 0.04 | 0.02 | | |
| | VEGF | 0.78 | 0.01 | 0.01 | | |
| | Combination | 1.22 | 0.01 | 0.00 | | |
| Week 3 | Control | 0.92 | 0.01 | 0.00 | 462 | 0.000 HS |
| | Lazar | 1.78 | 0.03 | 0.01 | | |
| | VEGF | 1.23 | 0.01 | 0.00 | | |
| | Combination | 1.95 | 0.11 | 0.04 | | |

(*) HS: Highly Sig. at $P < 0.01$; S: Sig. at $P < 0.05$; NS: Non Sig. at $P > 0.05$

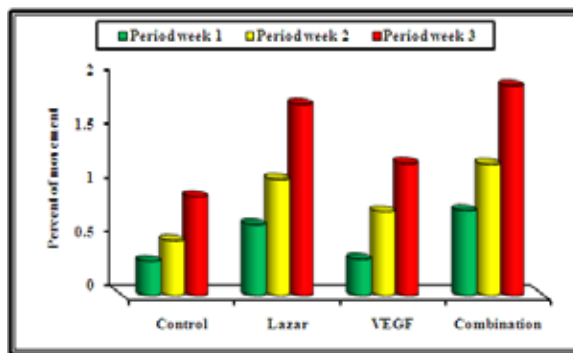


Figure 1: Cluster Bar Chart concerning Mean values for Clinical Measurements of distance (mm) between MCIs among studied groups at different intervals

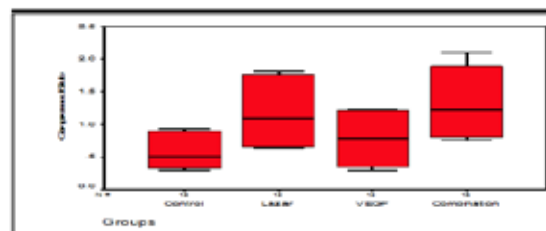


Figure 2: Stem-Leaf Plots for Clinical Measurements of distance (mm) between MCIs among studied groups at 3rd week.

Table 2: Area under the curve (ROC) for Clinical Measurements of distance (mm) between MCIs in contrast of control group at different periods

| Area Under the Curve | | | | | | | |
|--------------------------------------|-------|------------|------------------|----------|-------|------|-------|
| Test Result :working side- intervals | | | | | | | |
| Periods | Area | Std. Error | As-ymptotic Sig. | 95% C.I. | | Sen. | Spec. |
| | | | | L.B. | L.B. | | |
| 1 week | 0.056 | 0.047 | 0.001 | -0.037 | 0.148 | 100 | 88.9 |
| 2 week | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 100 | 100 |
| 3 week | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 100 | 100 |

Cutoff Point (0.33) at 1 week; Cutoff Point (0.52) at 2 week; Cutoff Point (0.93) at 3 week

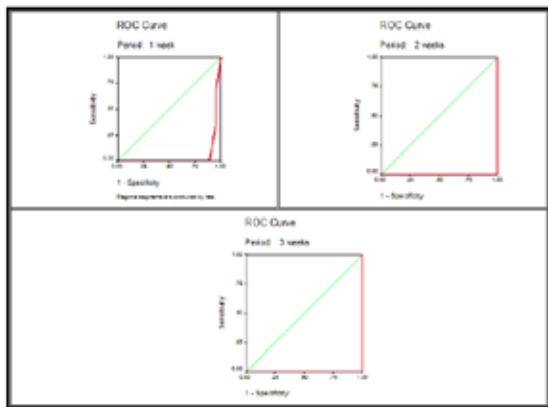


Figure 3: Receiver Operating Characteristic (ROC) curve for Clinical Measurements of distance (mm) between MCIs in contrast of control group at different intervals

Table 3: Area under the curve (ROC) for Clinical Measurements of distance (mm) between MCIs in contrast of control group at different sources of variation

| Area Under the Curve | | | | | |
|----------------------------------|-------|------------|-----------------|-------------------------|-------------|
| Test Result: working side-groups | | | | | |
| Control with others groups | Area | Std. Error | Asymptotic Sig. | 95% Confidence Interval | |
| | | | | Lower Bound | Upper Bound |
| | 0.191 | 0.055 | 0.000 | 0.084 | 0.299 |

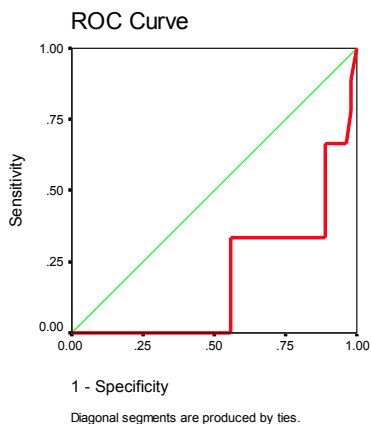


Figure 4: Receiver Operating Characteristic (ROC) curve Clinical Measurements of distance (mm) between MCIs in contrast of control group at different sources of variation

Table 4: Area under the curve (ROC) for Clinical Measurements of distance (mm) between MCIs in contrasts of all pair wise groups at different sources of variation

| Area Under the Curve | | | | | |
|--|-------|------------|-----------------|-------------------------|-------------|
| Test Result: working Side – study groups | | | | | |
| Contrasts | Area | Std. Error | Asymptotic Sig. | 95% Confidence Interval | |
| | | | | Lower Bound | Upper Bound |
| Control X Lazar | 0.111 | 0.054 | 0.000 | 0.006 | 0.217 |
| Control X VEGF | 0.352 | 0.094 | 0.129 | 0.169 | 0.535 |
| Control X Comb. | 0.111 | 0.054 | 0.000 | 0.006 | 0.217 |

| | | | | | |
|---------------|-------|-------|-------|-------|-------|
| Lazar X VEGF | 0.667 | 0.092 | 0.088 | 0.486 | 0.847 |
| Lazar X Comb. | 0.336 | 0.092 | 0.094 | 0.156 | 0.517 |
| VEGF X Comb. | 0.230 | 0.077 | 0.006 | 0.078 | 0.382 |

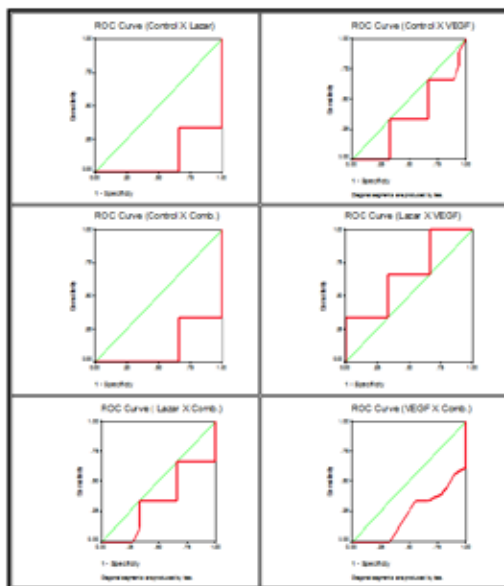


Figure 5: Receiver Operating Characteristic (ROC) curve for Clinical Measurements of distance (mm) between MCIs in contrasts of all pair wise groups at different sources of variation

Table (5): Descriptive statistics for percentages of movement (P.M.) for each contrast's periods at different groups and Comparisons significant

| S.O.V. | Per-iods | Groups | Mean of P.M. | Std. Dev. | ANOVA | | ANOVA | |
|--|----------|--------|--------------|-----------|---------|-------------------------|---------|-------------------------|
| | | | | | Periods | P ^(*) -value | Groups | P ^(*) -value |
| 1w. & 2w. Combination | Control | | 59.38 | 35.33 | F=23.90 | 0.001 HS | F=2.715 | 0.138 NS |
| | Lazar | | 63.64 | | | | | |
| | VEGF | | 129.41 | | | | | |
| 2w. & 3w. Combination | Control | | 80.39 | 10.2484 | F=23.90 | 0.001 HS | F=2.715 | 0.138 NS |
| | Lazar | | 64.81 | | | | | |
| | VEGF | | 57.69 | | | | | |
| 1w. & 3w. Combination | Control | | 187.5 | 49.74 | F=23.90 | 0.001 HS | F=2.715 | 0.138 NS |
| | Lazar | | 169.7 | | | | | |
| | VEGF | | 261.76 | | | | | |
| | | | 146.84 | | | | | |
| R Squared = 0.903 (Adjusted R Squared = 0.822) | | | | | | | | |

(*) HS: Sig. at P<0.05; NS: Non Sig. at P>0.05

Table (6): Multiple comparisons by (LSD) among all pairs of contrast's periods for (P.M.)parameter

| Parameter | Statistical tests | periods | periods | Mean Diff. | Sig. | C.S. (*) |
|---------------|-------------------|-----------|-----------|------------|-------|----------|
| Contrasts LSD | | 1w. & 2w. | 2w. & 3w. | 11.03 | 0.604 | NS |
| | | 2w. & 3w. | 1w. & 3w. | -114.74 | 0.001 | HS |
| | | 1w. & 3w. | 2w. & 3w. | -125.77 | 0.001 | HS |

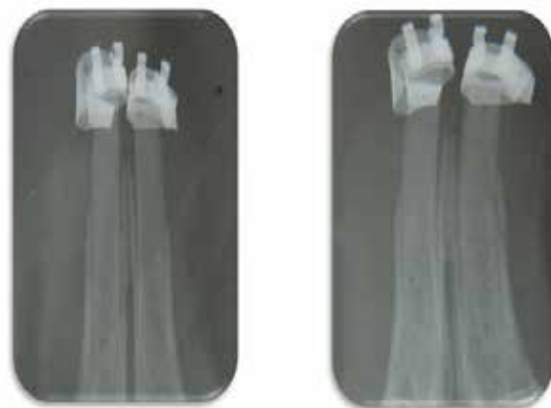
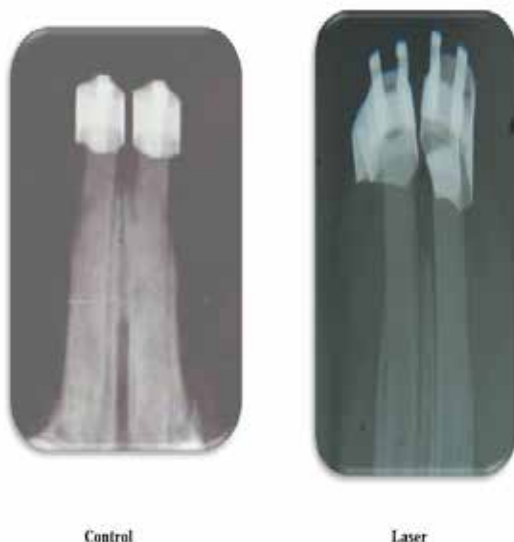
(*) HS: Highly Sig. at P<0.01; NS: Non Sig. at P>0.05



Figure(6)Photo for Biopsies shows the difference in the distance that remained between MCIs after 3 weeks duration of experiment.



Figure(7) Photo for Biopsies shows the difference in the distance that remained between MCIs after 3 weeks duration of experiment and after removing of orthodontic appliance.



VEGF Combination

Figure(8) Radiographical images for the study groups show the difference in the distance that remained between MCIs .

Table (7): Descriptive statistics of Clinical Measurements of distance (mm) between MCIs among studied groups at different intervals with comparisons significant

| Marker | Groups | Mean | SD | SE | Student test | |
|--------------|--------------|-------|------|------|--------------|------------------------|
| | | | | | t-value | P ^(*) Value |
| Control | Radiographic | 0.917 | 0.01 | 0.00 | -0.319 | 0.756 NS |
| | Clinical | 0.918 | 0.01 | 0.00 | | |
| Lazar | Radiographic | 1.768 | 0.03 | 0.01 | -0.592 | 0.567 NS |
| | Clinical | 1.778 | 0.03 | 0.01 | | |
| VEGF | Radiographic | 1.207 | 0.01 | 0.00 | -4.568 | 0.551 NS |
| | Clinical | 1.225 | 0.01 | 0.00 | | |
| Combina-tion | Radiographic | 1.933 | 0.10 | 0.04 | -0.221 | 0.830 NS |
| | Clinical | 1.947 | 0.11 | 0.04 | | |

(*) HS: Highly Sig. at P<0.01; S: Sig. at P<0.05; NS: Non Sig. at P>0.05

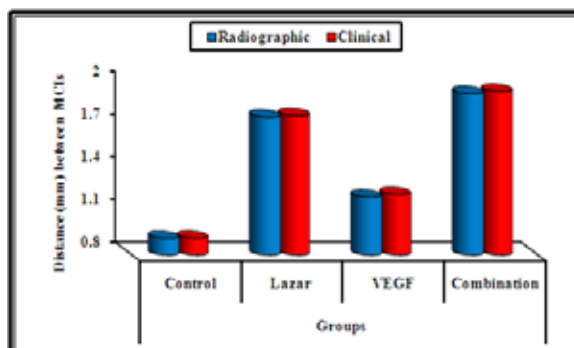


Figure (9): Cluster Bar Chart concerning Mean values for Clinical Measurements of distance (mm) between MCIs among studied groups at the end of third weeks

Amount of width of mandibular suture (mm)

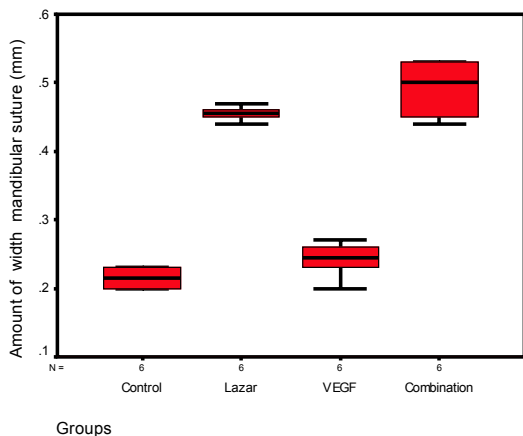


Figure (10): Stem-Leaf Plots for Amount of width mandibular suture (mm) among studied groups

Table (8): Area under the curve (ROC) for Amount of width mandibular suture (mm) in contrast of control group

| | | | | | | | |
|---|-------|------------|------------------|----------|------|------|-------|
| Area Under the Curve | | | | | | | |
| Test Result: Amount of width mandibular suture (mm) | | | | | | | |
| Control with others groups | Area | Std. Error | As-ymptotic Sig. | 95% C.I. | | Sen. | Spec. |
| | 0.056 | 0.049 | 0.001 | L.B. | L.B. | 100 | 94.4 |

Cutoff Point (0.23)

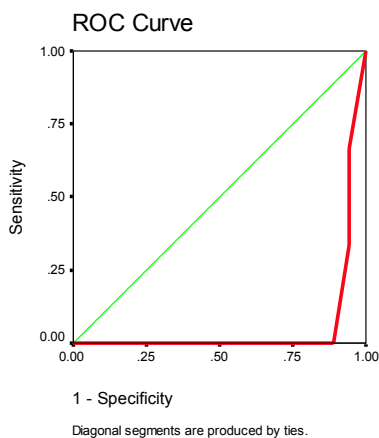


Figure (11): Receiver Operating Characteristic (ROC) curve for Amount of width mandibular suture (mm) in contrast of control group

Width of PDL of Right and Left Mesial & Distal Side

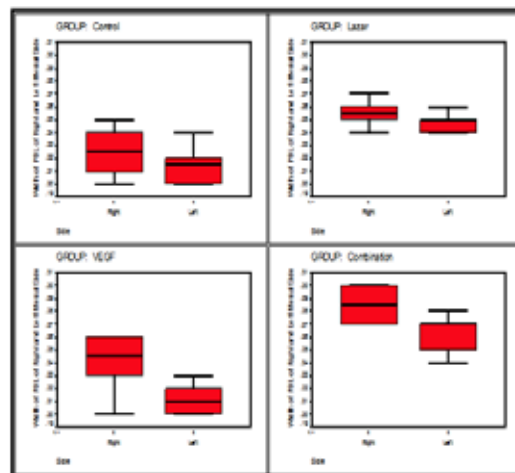


Figure 12: Stem-Leaf Plots for Width of PDL of Right and Left Mesial Side among studied groups

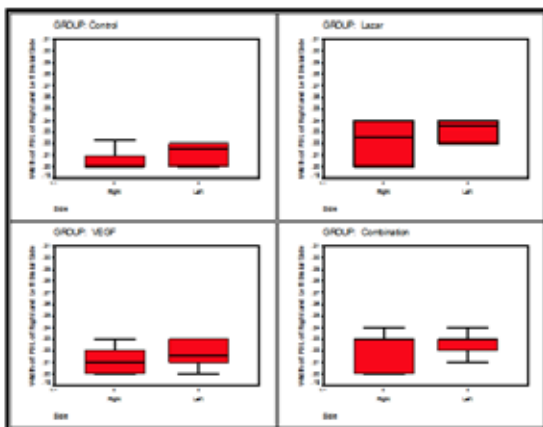


Figure 13: Stem-Leaf Plots for Width of PDL of Right and Left Side and Distal among studied groups

Discussion
It has been proved that the rate of OTM varies from person to another. However, clinical experiments combined with scientific evidence points an average rate of OTM of 1 mm per month [12,56].

As an alternative to surgically derived regional accelerations of OTM [14,57], several studies have attempted to expedite orthodontic treatment by increasing the velocity of OTM via pharmacological and electromagnetic modulations of the biological processes involved in bonemetabolism.

There have been several publications describing a positive effect on bone metabolism and OTM with the delivery of vitamin D [89], prostaglandin E 1 and 2 [90,91], osteocalcin [92], parathyroid hormone [93], long-term or high dose corticosteroids [93, 1992], thyroxin [94], pulsed electromagnetic fields [95], low intensity ultrasound [96] and low level laser therapy (LLLT) [51,71,72,73,75]. Although successful, the major drawback to most of these treatment interventions is the necessity for systemic delivery and the ensuing systemic side effects of pharmacological agents. Alternatively, local delivery of the pharmacological agent requires repeated painful injections while delivery of phototherapy using LLLT necessitates demanding scheduling challenges with the need for repeated and frequent treatment. Great-

est appeals of LLLT use to increase tooth movement is its avoidance of many of these risks and disadvantages associated with more invasive techniques. Clinical studies show that there are different Phases in tooth movement. The application of force during orthodontic tooth movement results in bone resorption by osteoclasts and deposition by osteoblasts on the pressure and tension sides of the periodontal ligament.

Recent studies in mice demonstrate that preosteoclasts, and not monocytes, may be recruited to the periodontal ligament during orthodontic tooth movement, and these cells may be targeted for acceleration of tooth movement. For the present study the

descriptive statistics for percentages of movement (P.M.) for each contrast's periods at different groups and Comparisons significant was recorded in table (5). High percentage recorded in 1w. & 3w. and specifically to group VEGF (261.76). The energy density is directly proportional with the exposure time, depending on the equation (Energy Density = Power Density X Exposure Time) Energy Density is a measure indicating the amount of energy received by a given tissue. It is calculated by dividing the energy output, measured in Joules, by the radius, or size, of the tissue receiving the laser application in centimeters² (cm²). This parameter is reported as J/cm². If a patient received 1 J application dose through the stationary placement of a diode fiber tip with the diameter of 1 cm, he/she would receive 1.27 J/cm². Power density can be used to distinguish between low and high power lasers. The absorption coefficient depends on the specific chromophore and wavelength of incident laser beam the meaning of that is photoreactions occur when a photon from light excites an absorbing molecule, result in photoreceptors or photoacceptors. [135 40]

This Photon absorption increases the energy state of the electrons of that molecule. If the increased energy is not emitted from the molecule as a photon of a longer wavelength or lost as heat, researchers propose the molecule undergo a chemical or morphogenic change which signals or allows for the initiation of a biochemical event. [135]

While it is not entirely clear which cellular molecules absorb light and have regulatory control, many studies have implicated components within the mitochondria. [144, 146, 23]

Resulting in unique characteristics of laser-tissue interactions and biological effects that is why it has been reported that multiple irradiation would be more effective for acceleration of cellular proliferation and bone formation with ideal wavelength.

Low-level laser therapy (LLLT)

Photobiomodulation or low-level laser therapy (LLLT) is one of the most promising approaches today. Laser has a biostimulatory effect on bone regeneration, which has been shown in the midpalatal suture during rapid palatal expansion [5], and also stimulates bone regeneration after bone fractures and extraction site. It has been found that laser light stimulates the proliferation of osteoclast, osteoblast, and fibroblasts, and thereby affects bone remodeling and accelerates tooth movement. The mechanism involved in the acceleration of tooth movement is by the production of ATP and activation of cytochrome C, as shown in [6] that low-energy laser irradiation enhanced the velocity of tooth movement via RANK/RANKL and the macrophage colony-stimulating factor and its receptor expression.

Animal experiments have shown that low-level laser can accelerate tooth movement. Furthermore, clinical trial attempts were made in which different intensities of laser were used and different results were obtained [5]. Low-level laser therapy can be a very useful technique for acceleration of tooth movement since it increases bone remodeling without side effects to the periodontium. Laser wavelength of 809 nm and output power 25 mW have indicated significant stimulation of bone metabolism, rapid ossification [7], and also acceleration of tooth movement to (1.5) fold in rat experiments. Lately in a clinical trial study, the laser wavelength they have used in a continuous wave mode at 800 nm, with an output of 25 mW, and exposure of 10 s was found to accelerate tooth movement at 1.3-fold higher than the control [8]. In another study done by Kau [9] on 90 subjects (73 test subjects and 17 controls), there was (1.12) mm change per week in the test subjects versus (0.49) mm in the control group. Having said this, there are a lot of contradictory results related to the LLLT. Therefore, more experiments are needed to differentiate the optimum energy, wavelength, and the optimum duration for usage. The present study shows 2w. & 3w. high significant about

-125.77 0.001 HS p<0.01. While it -114.7 0.604
NS 1w. & 3w. non-significant p>0.05

References

1. Z. Tooth movement. Crit Rev Oral Biol Med. Davidovitch 1991; 2(4) : 411-50.
2. Virinder Singh Kohli Role of RANKL-RANK/ osteoprotegerin molecular complex in bone remodeling and its immunopathologic implications Sarvraj Singh Kohli RANK pathway in giant cell tumor of bone: pathogenesis and therapeutic aspects Pan-Feng Wu, Ju-yu Tang, Kang-hua Li Tumor Biology. 2015; [PubMed].
3. STC1 interference on calcitonin family of receptors signaling during osteoblastogenesis via adenylate cyclase inhibition Silvia R. Terra, João Carlos d.R. Cardoso, Rute C. Félix, Leo Anderson M Martins, Diogo Onofre G.d Souza, Fatima C.R. Guma, Adelino Vicente M. Canário, Vanessa Schein Molecular and Cellular Endocrinology. 2015; [PubMed].
4. Role of Wnt/ β -catenin and RANKL/OPG in bone healing of diabetic Charcotarthropathy patients Agnetha Folestad, Martin Ålund, Susanne Asteberg, Jesper Fowelin, Ylva Aurell, Jan Göthlin, Jean Cassuto Acta Orthopaedica. 2015; : 1 [PubMed]
5. Osteoprotegerin and kidney disease Alejandra Montañez-Barragán, Isaías Gómez-Barrera, María D. Sanchez-Niño, Alvaro C. Ucero, Liliana González-Espinoza, Alberto Ortiz Journal of Nephrology. 2014; [PubMed].
6. Effect of low-level laser therapy irradiation and Bio-Oss graft material on the osteogenesis process in rabbit calvarium defects: a double blind experimental study Amir Alireza Rasouli Ghahroudi, Amir Reza Rokn, Katayoun A. M. Kalhori, Afshin Khorsand, Alireza Pournabi, A. L. B. Pinheiro, Reza Fekrazad Lasers in Medical Science. 2013; [PubMed].
7. Involvement of p38MAPK/NF- κ B Signaling Pathways in Osteoblasts Differentiation in Response to Mechanical Stretch Liang Wang, Jianyu Li, Xizheng Zhang, Lu Liu, Zong-ming Wan, Rui-xin Li, Yong Guo Annals of Biomedical Engineering. 2012; 40(9): 1884 [PubMed].
8. Kohli SS, Kohli VS. Role of RANKL-RANK/osteoprotegerin molecular complex in bone remodeling and its immunopathologic implications. Indian J Endocr Metab [serial online] 2011 [cited 2015 Jun 24]; 15:175-81. Available.
9. Desmet, K. D., Paz, D. A., Corry, J. J., Eells, J. T., Wong-Riley, M. T., Henry, M. M., . . . Whelan, H. T. Clinical and experimental application photobiomodulation. Photomed Laser Surgery, 24(2), 121-128. doi: 10.1089/pho.2006.24.121 (2006).
10. Karu, T. I., Pyatibrat, L. V., Kolyakov, S. F., & Afanasyeva, N. I. (2005). Absorption measurements of a cell monolayer relevant to phototherapy: Reduction of cytochrome c oxidase under near IR radiation. Journal of Photochemistry and Photobiology, B, Biology, 81(2), 98-106. doi: 10.1016/j.jphotobiol.2005.07.002

11. Kocoglu-Altan, S. The effects of Nd:YAG laser on maxillary canine distalization rate. *Turkish Journal of Orthodontics*, 22, 16-17-25. (2009).
12. Eells, J. T., Wong-Riley, M. T., VerHoeve, J., Henry, M., Buchman, E. V., Kane, M. P., . . . Whelan, H. T. Mitochondrial signal transduction in accelerated wound and retinal healing by near-infrared light therapy. *Mitochondrion*, 4(5-6), 559-567. doi: 10.1016/j.mito.2004.07.033(2004).
13. Schieke, S. M., Schroeder, P., & Krutmann, J. Cutaneous effects of infrared radiation: From clinical observations to molecular response mechanisms. *Photodermatology, Photoimmunology & Photomedicine*, 19(5), 228-234. (2003).
14. Stolik, S., Delgado, J. A., Anasagasti, L., & Perez, A. Effective thermal penetration depth in photo-irradiated ex vivo human tissues. *and Laser Surgery*, top, -100px; " width="0" height="0" type="application/x-vnd.skype.toolbars.npplugin.4.2. doi: 10.1089/pho.2010.2948 . (2011)
15. Saito, S., & Shimizu, N. Stimulatory effects of low-power laser irradiation on bone regeneration in midpalatal suture during expansion in the rat. *American Journal of Orthodontics and Dentofacial Orthopedics : Official Publication of the American Association of Orthodontists, its Constituent Societies, and the American Board of Orthodontics*, 111(5), 525-532. (1997)
16. Niemcz, M. H. *Laser-tissue interactions: Fundamentals and applications*. (3rd ed.). Germany: Springer Science and Business Media. Desmet, K. D., Paz, D. A., Corry, J. J.
17. Eells, J. T., Wong-Riley, M. T., Henry, M. M., . . . Whelan, H. T. (2006). Clinical and experimental applications of NIR-LED photobiomodulation. *Photomedicine and Laser Surgery*, 24(2), 121-128. doi: 10.1089/pho.2006.24.121. (2007).
18. Eells, J. T., Wong-Riley, M. T., VerHoeve, J., Henry, M., Buchman, E. V., Kane, M. P., . . . Whelan, H. T. Mitochondrial signal transduction in accelerated wound and retinal healing by near-infrared light therapy. *Mitochondrion*, 4(5-6), 559-567. doi: 10.1016/j.mito.2004.07.033 (2004).
19. Karu, T. I., Pyatibrat, L. V., Kolyakov, S. F., & Afanasyeva, N. I. Absorption measurements of a cell monolayer relevant to phototherapy: Reduction of cytochrome c oxidase under near IR radiation. *Journal of Photochemistry and Photobiology, B, Biology*, 81(2), 98-106. doi: 10.1016/j.jphoto.2005.07.002(2005).
20. Desmet, K. D., Paz, D. A., Corry, J. J., Eells, J. T., Wong-Riley, M. T., Henry, M. M., . . . Whelan, H. T. Clinical and experimental applications of NIR-LED photobiomodulation. *Photomedicine and Laser Surgery*, 24(2), 121-128. doi: 10.1089/pho.2006.24.121
21. Bicakci, A. A., Kocoglu-Altan, B., Toker, H., Mutaf, I., & Sumer, Z. (2012). Efficiency of low-level laser therapy in reducing pain induced by orthodontic forces. *Photomedicine and Laser Surgery*, 30(8), 460-465. doi: 10.1089/pho.2012.3245; 10.1089/pho.2012.3245 . (2006).
22. Doshi-Mehta, G., & Bhad-Patil, W. A. Efficacy of low-intensity laser therapy in reducing treatment time and orthodontic pain: A clinical investigation. *American Journal of Orthodontics and Dentofacial Orthopedics : Official Publication of the American Association of Orthodontists, its Constituent Societies, and the American Board of Orthodontics*, 141(3), 289-297. doi: 10.1016/j.ajodo.2011.09.009; 10.1016/j.ajodo.2011.09.009 . (2012)
23. Esper, M. A., Nicolau, R. A., & Arisawa, E. A. The effect of two phototherapy protocols on pain control in orthodontic procedure—a preliminary clinical study. *Lasers in Medical Science*, 26(5), 657-663. doi: 10.1007/s10103-011-0938-6; 10.1007/s10103-011-0938-6 .(2011)
24. Bicakci, A. A., Kocoglu-Altan, B., Toker, H., Mutaf, I., & Sumer, Z. Efficiency of low level laser therapy in reducing pain induced by orthodontic forces. *Photomedicine and Laser Surgery*, 30(8), 460-465. doi: 10.1089/pho.2012.3245; 10.1089/pho.2012.3245 .
25. Esper, M. A., Nicolau, R. A., & Arisawa, E. A. (2011). The effect of two phototherapy protocols on pain control in orthodontic procedure—a preliminary clinical study. *Lasers in Medical Science*, 26(5), 657-663. doi: 10.1007/s10103-011-0938-6; 10.1007/s10103-011-0938-6(2012)
26. Seifi, M., Shafeei, H. A., Daneshdoost, S., & Mir M. (2007). Effects of two types of low-level laser wave lengths (850 and 630 nm) on the orthodontic tooth movements in rabbits. *Lasers in Medical Science*, 22(4), 261-264. doi: 10.1007/s10103-007-0447-9
27. Turhani, D., Scheriau, M., Kapral, D., Benesch, T., Jonke, E., & Bantleon, H. Pain relief by single low-level laser irradiation in orthodontic patients undergoing fixed appliance therapy. *American Journal of Orthodontics and Dentofacial Orthopedics : Official Publication of the American Association of Orthodontists, its Constituent Societies, and the American Board of Orthodontics*, 130(3), 371-377. doi: 10.1016/j.ajodo.2005.04.036 . (2006).
28. Xiaoting, L., Yin, T., & Yangxi, C. (2010). Interventions for pain during fixed orthodontic appliance therapy. A systematic review. *The Angle Orthodontist*, 80(5), 925-932. doi: 10.2319/010410-10.1; 10.2319/010410-10.1 .
29. Yamaguchi, M., Hayashi, M., Fujita, S., Yoshida, T., Utsunomiya, T., Yamamoto, H., & Kasai, K. (2010). Low-energy laser irradiation facilitates the velocity of tooth movement and the expressions of matrix metalloproteinase-9, cathepsin K, and alpha(v) beta(3) integrin in rats. *European Journal of Orthodontics*, 32(2), 131-139. doi: 10.1093/ejo/cjp078; 10.1093/ejo/cjp078
30. Yamaguchi, M., Fujita, S., Yoshida, T., Oikawa, K., Utsunomiya, T., Yamamoto, H., & Kasai, K. (2007). Low-energy laser irradiation stimulates the tooth movement velocity via expression of M-CSF and c-fms. *Orthodontic Waves*, 66(4), 139-148.
31. Aihara, N., Yamaguchi, M., & Kasai, K. (2006). Low-energy irradiation stimulates formation of osteoclast-like cells via RANK expression in vitro. *Lasers in Medical Science*, 21(1), 24-33. doi: 10.1007/s10103-005-0368-4
32. Fujiyama, K., Deguchi, T., Murakami, T., Fujii, A., Kushima, K., & Takano-Yamamoto, T. (2008). Clinical effect of CO(2) laser in reducing pain in orthodontics. *The Angle Orthodontist*, 78(2), 299-303. doi: 10.2319/033007-153.1 .
33. Genc, G., Kocadereli, I., Tasar, F., Kilinc, K., El, S., & Sarkarati, B. (2013). Effect of low-level laser therapy (LLLT) on orthodontic tooth movement. *Lasers in Medical Science*, 28(1), 41-47. doi: 10.1007/s10103-012-1059-6; 10.1007/s10103-012-1059-6.
34. Kocoglu-Altan, S. (2009). The effects of Nd:YAG laser on maxillary canine distalization rate. *Turkish Journal of Orthodontics*, 22, 16-17-25.
35. Doshi-Mehta, G., & Bhad-Patil, W. A. (2012). Efficacy of low-intensity laser therapy in reducing treatment time and orthodontic pain: A clinical investigation. *American Journal of Orthodontics and Dentofacial Orthopedics : Official Publication of the American Association of Orthodontists, its Constituent Societies, and the American Board of Orthodontics*, 141(3), 289-297. doi: 10.1016/j.ajodo.2011.09.009; 10.1016/j.ajodo.2011.09.009 .
36. Habib, F. A., Gama, S. K., Ramalho, L. M., Cangussu, M. C., dos Santos Neto, F. P., Lacerda, J. A., . . . Pinheiro, A. L. (2012). Effect of laser phototherapy on the hyalinization following orthodontic tooth movement in rats. *Photomedicine and Laser Surgery*, 30(3), 179-185. doi: 10.1089/pho.2011.3085; 10.1089/pho.2011.3085.
37. Meikle, M. C. (2006). The tissue, cellular, and molecular regulation of orthodontic tooth movement: 100 years after Carl Sandstedt. *European Journal of Orthodontics*, 28(3), 221-240. doi: 10.1093/ejo/cjl001 .
38. Uysal, T., Ekizer, A., Akcay, H., Etoz, O., & Guray, E. (2010). Resonance frequency analysis of orthodontic miniscrews subjected to light-emitting diode photobiomodulation therapy. *European Journal of Orthodontics*, doi: 10.1093/ejo/cjq166
39. Limpanichkul, W., Godfrey, K., Srisuk, N., & Rattanayatikul, C. (2006). Effects of low-level laser therapy on the rate of orthodontic tooth movement. *Orthodontics & Craniofacial Research*, 9(1), 38-43. doi: 10.1111/j.1601-6343.2006.00338.x.
40. Cruz DR, Kohara EK, Ribeiro MS, Wetter NU. Effects of low-intensity laser therapy on the orthodontic movement velocity of human teeth: a preliminary study. *Lasers Surg Med*. 2004;35:117-20.
41. Goulart CS, Nouer PRA, Mouramartins L, Garbin IU, de Fátima Zanirato Lizarelli R. Photoradiation and orthodontic movement: experimental study with canines. *Photomed Laser Surg*. 2006;24:192-6.
42. T, Yamamoto H, Kasai K. Low-energy laser irradiation stimulates the tooth movement velocity via expression of M-CSF and c-fms. *Ortho Waves*. 2007;66:139-48.
43. Fujita S, Yamaguchi M, Utsunomiya T, Yamamoto H, Kasai K. Low-energy laser stimulates tooth movement velocity via expression of RANK and RANKL. *Orthod Craniofac Res*. 2008;11:143-55.
44. Kim S, Moon S, Kang S, Park Y. Effects of low-level laser therapy after Corticision on tooth movement and paradental remodeling. *Lasers Surg Med*. 2009;41:524-33.

45. Yoshida T, Yamaguchi M, Utsunomiya T, Kato M, Arai Y, Kaneda T, Yamamoto H, Kasai K. Low-energy laser irradiation accelerates the velocity of tooth movement via stimulation laser of the alveolar bone remodeling. *OrthodCraniofac Res.* 2009;12:289-98.
46. Mester E, Mester AF, Mester A. The biomedical effects of laser application. *Lasers Surg Med.* 1985;5:31-9.
47. Sommer AP, Pinheiro AL, Mester AR, Franke RP, Whelan HT. Biostimulatory windows in low-intensity laser activation: lasers, scanners, and NASA's light-emitting diode array system. *J Clin Laser Med Surg.* 2001;19:29-33.
48. Parker S. Low-level laser use in dentistry. *Br Dent J.* 2007;202:131-8.
49. Avery J, Steele P, Avery N. *Oral Development and Histology.* 3rd ed. New York: Thieme; 2002:74-84.
50. Nanci A. *Ten Cate's Oral Histology.* 7th ed. St. Louis: Mosby; 2007:89-95.
51. Marks SC, Schroeder HE. Tooth eruption: theories and facts. *Anat Rec.* 1996;245:374-93
52. Journal of Oral Sciences, vol. 115, no. 5, pp.355–362, 2007., T. P. Garlet, U. Coelho, J. S. Silva, and G. P. Garlet, "Cytokine expression pattern in compression and tension sides of the periodontal ligament during orthodontic tooth movement in humans," *European*
53. M. Yamaguchi, "RANK/RANKL/OPG during orthodontic tooth movement," *Orthodontics and Craniofacial Research*, vol.12, no. 2, pp. 113–119, 2009.
54. P. J. Brooks, D. Nilfroushan, M. F. Manolson, C. A. Simmons, and S. G. Gong, "Molecular markers of early orthodontic tooth movement," *Angle Orthodontist*, vol. 79, no. 6, pp. 1108–1113, 2009.
55. F. Uribe, Z. Kalajic, J. Bibko et al., "Early effects of orthodontic forces on osteoblast differentiation in a novel mouse organ culture model," *Angle Orthodontist*, vol. 81, no. 2, pp. 284–291, 2011.
56. C. Olson, F. Uribe, Z. Kalajic et al., "Orthodontic tooth movement causes decreased promoter expression of collagen type-1, bone sialoprotein and alpha-smooth muscle actin in the periodontal ligament," *Orthodontic Craniofacial Research*, vol.15, pp. 52–61, 2012.
57. P. J. Brooks, A. F. Heckler, K. Wei, and S. G. Gong, "M-CSF accelerates orthodontic tooth movement by targeting pre-osteoclasts in mice," *Angle Orthodontist*, vol. 81, no. 2, pp. 277–283, 2011.
58. Y. Ren, J. C. Maltha, and A. M. Kuijpers-Jagtman, "The rat as a model for orthodontic tooth movement—a critical review and a proposed solution," *European Journal of Orthodontics* vol. 26, no. 5, pp. 483–490, 2004.
59. C.C.Teixeira, E. Khoo, J. Tran et al., "Cytokine expression and accelerated tooth movement," *Journal of Dental Research*, vol.89, no. 10, pp. 1135–1141, 2010.
60. L. R. Iwasaki, J. R. Chandler, D. B. Marx, J. P. Pandey, and J.C. Nickel, "IL-1 gene polymorphisms, secretion in GCF, and speed of human tooth orthodontic movement," *Orthodontics and Craniofacial Research*, vol. 12, no. 2, pp. 129–140, 2009.
61. W. R. Proffit, "Biologic basis of orthodontic therapy," in *Contemporary Orthodontics*, W.R. Proffit and H. W. Fields, Eds., Mosby, St. Louis, Mo, USA, 3rd edition, 2000.
62. A. Bletsa, E. Berggreen, and P. Brudvik, "Interleukin-1 and tumor necrosis factor- expression during the early phases of orthodontic tooth movement in rats," *European Journal of Oral Sciences*, vol. 114, no. 5, pp. 423–429, 2006.
63. C.C.Teixeira, E. Khoo, J. Tran et al., "Cytokine expression and accelerated tooth movement," *Journal of Dental Research* vol.89, no. 10, pp. 1135–1141, 2001.
64. Masaru. et al. Low-energy laser irradiation stimulates the tooth movement velocity via expression of M-CSF and c-fms, *Orthodontic Waves* December 2007, Vol.66(4):139–148, doi:10.1016/j.odw.2007.09.002
65. Kim YD, Kim SS, Hwang DS, Kim SG, Kwon YH, Shin SH, et al. Effect of low-level laser treatment after installation of dental titanium implant-immunohistochemical study of RANKL, RANK, OPG: an experimental study in rats. *Lasers Surg Med.* 2007;39:441–450. [PubMed]
66. Mester E, Mester AF, Mester A. The biomedical effects of laser application. *Lasers Surg Med.* 1985;5:31–39.[PubMed]
67. Hahm E, Kulhari S, Arany PR. Targeting the pain, inflammation and immune (PII) axis: plausible rationale for LLLT. *Photonics Lasers Med.* 2012;1:241–254.
68. Arany PR, Nayak RS, Hallikerimath S, Limaye AM, Kale AD, Kondaiah P. Activation of latent TGF-beta1 by low-power laser in vitro correlates with increased TGF-beta1 levels in laser-enhanced oral wound healing. *Wound Repair Regen.* 2007;15:866–874. [PubMed]
69. Thalji G, Cooper LF. Molecular assessment of osseointegration in vivo: a review of the current literature. *Int J Oral Maxillofac Implants.* 2013;28:e521–e534.
70. Bengtson AL, Bengtson NG, Bengtson CR, Mendes FM, Pinheiro SR. Histological and radiographic evaluation of the muscle tissue of rats after implantation of bone morphogenetic protein (rhBMP-2) in a scaffold of inorganic bone and after stimulation with low-power laser light. *Indian J Dent Res.* 2010;21:420–424. [PubMed]
71. Pinto MR, dos Santos RL, Pithon MM, Araujo MT, Braga JP, Nojima LI. Influence of low-intensity laser therapy on the stability of orthodontic mini-implants: a study in rabbits. *Oral Surg Oral Med Oral Pathol Oral Radiol.* 2013;115:e26–e30. [PubMed]
72. Favero LG, Pisoni A, Paganelli C. Removal torque of osseointegrated mini-implants: an in vivo evaluation. *Eur J Orthod.* 2007;29:443–448. [PubMed]
73. Dortbudak O, Haas R, Mailath-Pokorny G. Effect of low-power laser irradiation on bony implant sites. *Clin Oral Implants Res.* 2002;13:288–292. [PubMed]
74. Pinto MR, dos Santos RL, Pithon MM, Araujo MT, Braga JP, Nojima LI. Influence of low-intensity laser therapy on the stability of orthodontic mini-implants: a study in rabbits. *Oral Surg Oral Med Oral Pathol Oral Radiol.* 2013;115:e26–e30. [PubMed]
75. Favero LG, Pisoni A, Paganelli C. Removal torque of osseointegrated mini-implants: an in vivo evaluation. *Eur J Orthod.* 2007;29:443–448. [PubMed]
76. Dortbudak O, Haas R, Mailath-Pokorny G. Effect of low-power laser irradiation on bony implant sites. *Clin Oral Implants Res.* 2002;13:288–292. [PubMed]
77. Maluf AP, Maluf RP, Brito Cda R, Franca FM, de Brito RB, Jr. Mechanical evaluation of the influence of low-level laser therapy in secondary stability of implants in mice shinbones. *Lasers Med Sci.* 2010;25:693–698. [PubMed]
78. Simmons D. The use of animal models in studying genetic disease: transgenesis and induced mutation. *Nature Educ.* 2008;1:70.
79. Mapara M, Thomas BS, Bhat KM. Rabbit as an animal model for experimental research. *Dent Res J (Isfahan)* 2012;9:111–118. [PMC free article] [PubMed]
80. Wang X, Mabrey JD, Agrawal CM. An interspecies comparison of bone fracture properties. *Biomed Mater Eng.* 1998;8:1–9. [PubMed]
81. VandeBerg JL, Williams-Blangero S. Advantages and limitations of non-human primates as animal models in genetic research on complex diseases. *J Med Primatol.* 1997;26:113–119. [PubMed]
82. Cruz, D. R., Kohara, E. K., Ribeiro, M. S., & Wetter, N. U. (2004). Effects of low-intensity laser therapy on the orthodontic movement velocity of human teeth: A preliminary study. *Lasers in Surgery and Medicine*, 35 (2), 117-120. doi: 10.1002/lsm.20076
83. Genc, G., Kocadereli, I., Tasar, F., Kilinc, K., El, S., & Sarkerati, B. Effect of low-level laser therapy (LLLT) on orthodontic tooth movement. *Lasers in Medical Science*, 28(1), 41-47. doi: 10.1007/s10103-012-1059-6; (2013).
84. 10.1007/s10103-012-1059-6 Gonzales, C., Hotokezaka, H., Matsuo, K., Shibazaki, T., Kalia, S., Melsen, B., & Verna, CTissue reaction to orthodontic tooth movement in acute and chronic corticosteroid treatment. *Orthodontics & Craniofacial Research*, 7(1), 26-34. doi: 10.1111/j.1601-6343.2004.00278. (2004)
85. Kocoglu-Altan, S. The effects of Nd:YAG laser on maxillary canine distalization rate. *Turkish Journal of Orthodontics*, 22, 16-17-25(2009).
86. Hadjiargyrou, M., McLeod, K., Ryaby, J. P., & Rubin, C. Enhancement of fracture healing by low intensity ultrasound. *Clinical Orthopaedics and Related Research*, (355 Suppl) (355 Suppl), S216-29(1998).
87. Li, W. T., Leu, Y. C., & Wu, J. L. Red-light light-emitting diode irradiation increases the proliferation and osteogenic differentiation of rat bone marrow mesenchymal stem cells. *Photomedicine and Laser Surgery*, 28 Suppl 1, S157-65. doi: 10.1089/pho.2009.2540; 10.1089/pho.2009.2540 (2010).

88. Hashimoto, F., Kobayashi, Y., Mataka, S., Kobayashi, K., Kato, Y., & Sakai, H. Administration of osteocalcin accelerates orthodontic tooth movement induced by a closed coil spring in rats. *European Journal of Orthodontics*, 23(5), 535-545(2001).
89. Sommer, A. P., Pinheiro, A.L., Mester, A. R., Franke, R. P., & Whelan, H. T. Biostimulatory windows in low-intensity laser activation: Lasers, scanners, and NASA's light-emitting diode array system. *Journal of Clinical Laser Medicine & Surgery*, 19(1), 29-33. doi: 10.1089/104454701750066910 (2001)
90. Wilcko, M. T., Wilcko, W. M., Pulver, J. J., Bissada, N. F., & Bouquet, J. E. Accelerated osteogenic orthodontics technique: A 1-stage surgically facilitated rapid orthodontic technique with alveolar augmentation. *Journal of Oral and Maxillofacial Surgery : Official Journal of the American Association of Oral and Maxillofacial Surgeons*, 67(10), 2149-2159. doi: 10.1016/j.joms.2009.04.095; 10.1016/j.joms.2009.04.095. (2009).
91. Yamaguchi, M., Fujita, S., Yoshida, T., Oikawa, K., Utsunomiya, T., Yamamoto, H., & Kasai, K. Low-energy laser irradiation stimulates the tooth movement velocity via expression of M-CSF and c-fms. *Orthodontic Waves*, 66(4), 139-148(2007)
92. Youssef, M., Ashkar, S., Hamade, E., Gutknecht, N., Lampert, F., & Mir, M. The effect of low-level laser therapy during orthodontic movement: A preliminary study. *Lasers in Medical Science*, 23(1), 27-33. doi: 10.1007/s10103-007-0449-7 (2008).
93. Seifi M, Doshid-Dastjerdi E.E efficacy of low- intensity in reducing treatment timed orthodontic pain; a clinical investigation. *Am. j Orthod-Dentofacial Orthop.* 2012 Mar;141(3);289-97.
94. Virinder Singh Koh Role of RANKL-RANK/ osteoprotegerin molecular complex in bone remodeling and its immunopathologic implications Sarvraj Singh Kohli RANK pathway in giant cell tumor of bone: pathogenesis and therapeutic aspects Pan-Feng Wu, Ju-yu Tang, Kang-hua Li *Tumor Biology*. 2015; [PubMed].
95. STC1 interference on calcitonin family of receptors signaling during osteoblastogenesis via adenylate cyclase inhibition Silvia R Terra, João Carlos d.R. Cardoso, Rute C. Félix, Leo Anderson M Martins, Diogo Onofre G.d Souza, Fatima C.R. Guma, Adelino Vicente M. Canário, Vanessa Schein *Molecular and Cellular Endocrinology*. 2015; [PubMed].
96. Role of Wnt/ β -catenin and RANKL/OPG in bone healing of diabetic Charcotarthropathy patients Agnetha Folestad, Martin Ålund, Susanne Astenberg, Jesper Fowelin, Ylva Aurell, Jan Göthlin, Jean Cassuto *Acta Orthopaedica*. 2015; : 1 [PubMed]
97. Osteoprotegerin and kidney disease Alejandra Montañez-Barragán, Isaías Gómez-Barrera, María D. Sánchez-Niño, Alvaro C. Ucero, Liliana González-Espinoza, Alberto Ortiz *Journal of Nephrology*. 2014; [PubMed].
98. Effect of low-level laser therapy irradiation and Bio-Oss graft material on the osteogenesis process in rabbit calvarium defects: a double blind experimental study Amir Alireza Rasouli Ghahroudi, Amir Reza Rokn, Katayoun A. M. Kalhori, Afshin Khorsand, Alireza Pourmabi, A. L. B. Pinheiro, Reza Fekrazad *Lasers in Medical Science*. 2013; [PubMed].
99. Involvement of p38MAPK/NF- κ B Signaling Pathways in Osteoblasts Differentiation in Response to Mechanical Stretch Liang Wang, Jianyu Li, Xizheng Zhang, Lu Liu, Zong-ming Wan, Rui-xin Li, Yong Guo *Annals of Biomedical Engineering*. 2012; 40(9): 1884 [PubMed].
100. Kohli SS, Kohli VS. Role of RANKL-RANK/osteoprotegerin molecular complex in bone remodeling and its immunopathologic implications. *Indian J Endocr Metab [serial online]* 2011 [cited 2015 Jun 24];15:175-81. Available.
101. Desmet, K. D., Paz, D. A., Corry, J. J., Eells, J. T., Wong-Riley, M. T., Henry, M. M., . . . Whelan, H. T. Clinical and experimental application photobiomodulation. *Photomed and Laser Surgery*, 24(2), 121-128. doi: 10.1089/pho.2006.24.121 (2006).
102. Karu, T. I., Pyatibrat, L. V., Kolyakov, S. F., & Afanasyeva, N. I. (2005). Absorption measurements of a cell monolayer relevant to phototherapy: Reduction of cytochrome c oxidase under near IR radiation. *Journal of Photochemistry and Photobiology, B, Biology*, 81(2), 98-106. doi: 10.1016/j.jphotobiol.2005.07.002
103. Kocoglu-Altan, S. The effects of Nd:YAG laser on maxillary canine distalization rate. *Turkish Journal of Orthodontics*, 22, 16-17-25. (2009).
104. Eells, J. T., Wong-Riley, M. T., VerHoeve, J., Henry, M., Buchman, E. V., Kane, M. P., . . . Whelan, H. T. Mitochondrial signal transduction in accelerated wound and retinal healing by near-infrared light therapy. *Mitochondrion*, 4(5-6), 559-567. doi: 10.1016/j.mito.2004.07.033(2004).
105. Schieke, S. M., Schroeder, P., & Krutmann, J. Cutaneous effects of infrared radiation: From clinical observations to molecular response mechanisms. *Photodermatology, Photoimmunology & Photomedicine*, 19(5), 228-234. (2003).
106. Stolik, S., Delgado, J. A., Anasagasti, L., & Perez, A. M. Effective thermal penetration depth in photo-irradiated ex vivo human tissues. *Laser Surgery, top*, -100px; "width="0" height="0" type="application/x-vnd.skype.toolbars.npplugin.4.2. doi: 10.1089/pho.2010.2948 . (2011)
107. Saito, S., & Shimizu, N. Stimulatory effects of low-power laser irradiation on bone regeneration in midpalatal suture during expansion in the rat. *American Journal of Orthodontics and Dentofacial Orthopedics : Official Publication of the American Association of Orthodontists, its Constituent Societies, and the American Board of Orthodontics*, 111(5), 525-532. (1997)
108. Niemz, M. H. *Laser-tissue interactions: Fundamentals and applications*. (3rd ed.). Germany: Springer Science and Business Media. Desmet, K. D., Paz, D. A., Corry, J. J.
109. Eells, J. T., Wong-Riley, M. T., Henry, M. M., . . . Whelan, H. T. (2006). Clinical and experimental applications of NIR-LED photobiomodulation. *Photomedicine and Laser Surgery*, 24(2), 121-128. doi: 10.1089/pho.2006.24.121. (2007).
110. Eells, J. T., Wong-Riley, M. T., VerHoeve, J., Henry, M., Buchman, E. V., Kane, M. P., . . . Whelan, H. T. Mitochondrial signal transduction in accelerated wound and retinal healing by near-infrared light therapy. *Mitochondrion*, 4(5-6), 559-567. doi: 10.1016/j.mito.2004.07.033 (2004).
111. Karu, T. I., Pyatibrat, L. V., Kolyakov, S. F., & Afanasyeva, N. I. Absorption measurements of a cell monolayer relevant to phototherapy: Reduction of cytochrome c oxidase under near IR radiation. *Journal of Photochemistry and Photobiology, B, Biology*, 81(2), 98-106. doi: 10.1016/j.jphotobiol.2005.07.002(2005).
112. Desmet, K. D., Paz, D. A., Corry, J. J., Eells, J. T., Wong-Riley, M. T., Henry, M. M., . . . Whelan, H. T. Clinical and experimental applications of NIR-LED photobiomodulation. *Photomedicine and Laser Surgery*, 24(2), 121-128. doi: 10.1089/pho.2006.24.121
113. Bicakci, A. A., Kocoglu-Altan, B., Toker, H., Mutaf, I., & Sumer, Z. (2012). Efficiency of low-level laser therapy in reducing pain induced by orthodontic forces. *Photomedicine and Laser Surgery*, 30(8), 460-465. doi: 10.1089/pho.2012.3245; 10.1089/pho.2012.3245 . (2006).
114. Doshi-Mehta, G., & Bhad-Patil, W. A. Efficacy of low-intensity laser therapy in reducing treatment time and orthodontic pain: A clinical investigation. *American Journal of Orthodontics and Dentofacial Orthopedics : Official Publication of the American Association of Orthodontists, its Constituent Societies, and the American Board of Orthodontics*, 141(3), 289-297. doi: 10.1016/j.ajodo.2011.09.009; 10.1016/j.ajodo.2011.09.009 . (2012)
115. Esper, M. A., Nicolau, R. A., & Arisawa, E. A. The effect of two phototherapy protocols on pain control in orthodontic procedure—a preliminary clinical study. *Lasers in Medical Science*, 26(5), 657-663. doi: 10.1007/s10103-011-0938-6; 10.1007/s10103-011-0938-6 . (2011)
116. Bicakci, A. A., Kocoglu-Altan, B., Toker, H., Mutaf, I., & Sumer, Z. Efficiency of low level laser therapy in reducing pain induced by orthodontic forces. *Photomedicine and Laser Surgery*, 30(8), 460-465. doi: 10.1089/pho.2012.3245; 10.1089/pho.2012.3245 .
117. Esper, M. A., Nicolau, R. A., & Arisawa, E. A. (2011). The effect of two phototherapy protocols on pain control in orthodontic procedure—a preliminary clinical study. *Lasers in Medical Science*, 26(5), 657-663. doi: 10.1007/s10103-011-0938-6; 10.1007/s10103-011-0938-6(2012)
118. Seifi, M., Shafeei, H. A., Daneshdoost, S., & Mir M. (2007). Effects of two types of low-level laser wave lengths (850 and 630 nm) on the orthodontic tooth movements in rabbits. *Lasers in Medical Science*, 22(4), 261-264. doi: 10.1007/s10103-007-0447-9
119. Turhani, D., Scheriau, M., Kapral, D., Benesch, T., Jonke, E., & Bantleon, H. P. Pain relief by single low-level laser irradiation in orthodontic patients undergoing fixed appliance therapy. *American Journal of Orthodontics and Dentofacial Orthopedics : Official Publication of the American Association of Orthodontists, its Constituent Societies, and the American Board of Orthodontics*, 130(3), 371-377. doi: 10.1016/j.

- ajodo.2005.04.036 . (2006).
120. Xiaoting, L., Yin, T., &Yangxi, C. (2010). Interventions for pain during fixed orthodontic appliance therapy. A systematic review. *The Angle Orthodontist*, 80(5), 925-932. doi: 10.2319/010410-10.1; 10.2319/010410-10.1 .
 121. Yamaguchi, M., Hayashi, M., Fujita, S., Yoshida, T., Utsunomiya, T., Yamamoto, H., & Kasai, K. (2010). Low-energy laser irradiation facilitates the velocity of tooth movement and the expressions of matrix metalloproteinase-9, cathepsin K, and alpha(v) beta(3) integrin in rats. *European Journal of Orthodontics*, 32(2), 131-139. doi: 10.1093/ejo/cjp078; 10.1093/ejo/cjp078
 122. Yamaguchi, M., Fujita, S., Yoshida, T., Oikawa, K., Utsunomiya, T., Yamamoto, H., & Kasai, K. (2007). Low-energy laser irradiation stimulates the tooth movement velocity via expression of M-CSF and c-fms. *Orthodontic Waves*, 66(4), 139-148.
 123. Aihara, N., Yamaguchi, M., & Kasai, K. (2006). Low-energy irradiation stimulates formation of osteoclast-like cells via RANK expression in vitro. *Lasers in Medical Science*, 21(1), 24-33. doi: 10.1007/s10103-005-0368-4
 124. Fujiyama, K., Deguchi, T., Murakami, T., Fujii, A., Kushima, K., & Takano-Yamamoto, T. (2008). Clinical effect of CO(2) laser in reducing pain in orthodontics . *The Angle Orthodontist*, 78(2), 299-303. doi: 10.2319/033007-153.1 .
 125. Genc, G., Kocadereli, I., Tasar, F., Kilinc, K., El, S., &Sarkarati, B. (2013). Effect of low-level laser therapy (LLLT) on orthodontic tooth movement. *Lasers in Medical Science*, 28(1), 41-47. doi: 10.1007/s10103-012-1059-6; 10.1007/s10103-012-1059-6.
 126. Kocoglu-Altan, S. (2009). The effects of nd:YAG laser on maxillary canine distalization rate. *Turkish Journal of Orthodontics*, 22, 16-17-25.
 127. Doshi-Mehta, G., &Bhad-Patil, W. A. (2012). Efficacy of low-intensity laser therapy in reducing treatment time and orthodonticpain: A clinicalinvestigation. *American Journal of Orthodontics and Dentofacial Orthopedics : Official Publication of the American Association of Orthodontists, its Constituent Societies, and the American Board of Orthodontics*, 141(3), 289-297. doi: 10.1016/j.ajodo.2011.09.009; 10.1016/j.ajodo.2011.09.009 .
 128. Habib, F. A., Gama, S. K., Ramalho, L. M., Cangussu, M. C., dos Santos Neto, F. P., Lacerda, J. A., . . . Pinheiro, A. L. (2012). Effect of laser phototherapy on the hyalinization following orthodontic tooth movement in rats. *Photomedicine and Laser Surgery*, 30(3), 179-185. doi: 10.1089/pho.2011.3085; 10.1089/pho.2011.3085.
 129. Meikle, M. C. (2006). The tissue, cellular, and molecular regulation of orthodontic tooth movement: 100 years after carlsandstedt. *European Journal of Orthodontics*, 28(3), 221-240. doi: 10.1093/ejo/cj001 .
 130. Uysal, T., Ekizer, A., Akcay, H., Etoz, O., &Guray, E. (2010). Resonance frequency analysis of orthodontic miniscrews subjected to light-emitting diode photobiomodulation therapy. *European Journal of Orthodontics*, doi: 10.1093/ejo/cjq166
 131. Limpanichkul, W., Godfrey, K., Srisuk, N., &Rattanayatikul, C. (2006). Effects of low-level laser therapy on the rate of orthodontic tooth movement. *Orthodontics & Craniofacial Research*, 9(1), 38-43. doi: 10.1111/j.1601-6343.2006.00338.x.
 132. Cruz DR, Kohara EK, Ribeiro MS, Wetter NU. Effects of low-intensity laser therapy on the orthodontic movement velocity of human teeth: a preliminary study. *Lasers Surg Med*. 2004; 35:117-20.
 133. Goulart CS, Nouer PRA, Mouramartins L, Garbin IU, de Fátima Zanirato Lizarelli R. Photoradiation and orthodontic movement: experimental study with canines. *Photomed Laser Surg*. 2006;24:192-6.
 134. T, Yamamoto H, Kasai K. Low-energy laser irradiation stimulates the tooth movement velocity via expression of M-CSF and c-fms. *Ortho Waves*. 2007;66:139-48.
 135. Fujita S, Yamaguchi M, Utsunomiya T, Yamamoto H, Kasai K. Low-energy laser stimulates tooth movement velocity via expression of RANK and RANKL. *Orthod Craniofac Res*. 2008;11:143-55.
 136. Kim S, Moon S, Kang S, Park Y. Effects of low-level laser therapy after Corticision on tooth movement and paradental remodeling. *Lasers Surg Med*. 2009;41:524-33.
 137. Yoshida T, Yamaguchi M, Utsunomiya T, Kato M, Arai Y, Kameda T, Yamamoto H, Kasai K. Low-energy laser irradiation accelerates the velocity of tooth movement via stimulation of the alveolar bone remodeling. *Orthod Craniofac Res*. 2009;12:289-98.
 138. Mester E, Mester AF, Mester A. The biomedical effects of laser application. *Lasers Surg Med*. 1985;5:31-9.
 139. Sommer AP, Pinheiro AL, Mester AR, Franke RP, Whelan HT. Biostimulatory windows in low-intensity laser activation: lasers, scanners, and NASA's light-emitting diode array system. *J Clin Laser Med Surg*. 2001; 19:29-33.
 140. Parker S. Low-level laser use in dentistry. *Br Dent J*. 2007;202:131-8.
 141. Avery J, Steele P, Avery N. *Oral Development and Histology*. 3rd ed. New York: Thieme; 2002:74-84.
 142. Nanci A. *Ten Cate's Oral Histology*. 7th ed. St. Louis: Mosby; 2007:89-95.
 143. Marks SC, Schroeder HE. Tooth eruption: theories and facts. *Anat Rec*. 1996;245:374-93