RESEARCH PAPER

Botany



Antioxidant Activity of One Year Old Aloe Vera Plants Extract After Applying of Organic Manure

KEYWORDS	Superoxide dismutase, glutathione reductase, catalase, ascorbic acid and total $\beta-$ carotene							
Rakesh Me	hta	* Saroj Mawase	Zea–ul–Hasan					
Department of Botar Science College, Itarsi	ny, M. G. M. (M. P.), India.	Department of Botany, M. G. M. Science College, Itarsi (M. P.), India. * corresponding author	Department of Botany, Sofea College, Bhopal (M.P.), India.					

ABSTRACT Aloe vera is a stemless or very short-stemmed succulent plant growing to 60–100 cm (24–39 inch), spreading by offsets. The leaves are thick and fleshy, green to grey-green, with some varieties showing white flecks on their upper and lower stem surfaces like other Aloe species. Aloe vera forms a symbiosis that allows the plant better access to mineral nutrients in soil with applied of farm yard manure. Antioxidants are the substances in Aloe vera which can prevent or slow oxidative damage to cells. These agents are able to remove the deleterious effects of free radicals within cells. Solvent extraction methods are widely used for extracting antioxidants in food and other sources.

Introduction:

Plants, vegetables and herbs used in the folk and traditional medicine have been accepted currently as one of the main source of drug discovery and development.⁽¹⁾ There is a growing interest in the pharmacological evaluation of various plants used in Indian traditional system of medicine. Aloe vera (Sottrukattalai, Tamil) is a perennial succulent belonging to the Liliaceae family and is called the healing plant or the silent healer. Aloe vera contains 75 potentially active constituents: vitamins, enzymes, minerals, sugars, lignin, saponins, salicylic acids, and amino acids.⁽²⁾ Aloe gel has demonstrated wound healing⁽³⁾, antiinflammatory⁽⁴⁾, antiviral⁽⁵⁾, spermicidal⁽⁶⁾, gastroprotective⁽⁷⁾ and immune stimulating⁽⁸⁾ properties. Plant derived natural products such as flavonoids, terpenoids and steroids etc have received considerable attention in recent years due to their diverse pharmacological properties including antioxidant and antitumor activity.^(9, 10) Antioxidants play an important role in inhibiting and scavenging free radicals, thus providing protection against infection and degenerative diseases. From this viewpoint, the present study was carried out to evaluate the antioxidant status of ethanol extract of one year Aloe vera plants.

Material and methods:

A study undertaken to evaluated the "Effect of graded levels of fertilizer application on antioxidants dynamics of *Aloe vera*". The study was conducted in the Department of Botany, Sofea College, Bhopal (M.P.), India (23°16 0 North, 77°24 0 East). Experimental *Aloe vera* plants were completely cultivated in four pots from 2010 to 2014.

Field of proposed works:

The present study is performed to evaluate the antioxidants effect of aqueous extract of one year *Aloe vera* plants.

(i) Climate:

The climate of the area is semi-arid subtropical monsoon type with an average annual rainfall received during the monsoon month (June–September). The mean maximum and minimum temperature ranged from 29–44°C and 9–23°C respectively. The area is characterized by hot sum-

mer and mild winter. The mean date of commencement of monsoon is around June-19 whereas the mean date of withdrawal of monsoon is September-21.

(ii) Preparation of experimental soils:

A field experiment is laid out at **Sofea College**, **Bhopal** (**M.P.**). The experiment is conducted in fixed pots for cropping of *Aloe vera*. The soil for experimental is sandy coastal and heavy black cotton soils.

(iii) Pot's soil preparation (Treatment):

T-1. Sandy coastal soil: Golden sand: Farm yard manure (1:1:1), (4 Pots).

T–3. Heavy black cotton soil: Golden sand: Farm yard manure (1:1:1), (4 Pots).

T–5. Control (Sandy coastal soil: Golden sand – 1: 1), (4 Pots).

T-6. Control (Heavy black cotton soil: Golden sand - 1: 1), (4 Pots).

(iv) Planting time:

Suckers should be planted in July-August (2010) during monsoon season to get better field survival and subsequent growth of the plants.

Sample collection for investigation of antioxidants (Aloe vera):

The thick fleshy leaves are ready for harvest from the first after planting. It is carried out in the morning. Leaves were weighed and expressed in mg per leaf. Leaf extracts were prepared from 2 g fresh weight. Fresh clean whole *Aloe vera* leaves were cut and the outer green rind was discarded. The tip and basal portions of *Aloe vera* leaves are trimmed off and washed in clean water to remove soil and other dirty materials. Finally the leaves were soaked in clean distilled water. After removing the rinds from the leaves, the inner gel was collected. The mucilaginous inner pulp was minced and thoroughly homogenised with a hand held blender. Each leaf produced approximately 120 ml of gel. The homogenised gel was lyophilised at 22°C

RESEARCH PAPER

Preparation of Aloe vera leaf extract:

Freshly collected lyophilised material of *Aloe vera* homogenized with 800 ml PBS (phosphate-buffered saline: 0.06 mM sodium phosphate buffer containing 0.15 M NaCl, pH 7.4) in a small blender, extracted with PBS, at room temperature overnight, filtered through cloth and then centrifuged at 6000 rpm for 30 min. The precipitate was discarded and the clear yellow supernatant (45 ml) was named *Aloe vera* leaf gel (**AVLG**).

Chemicals – All chemicals and reagents used in the study were of analytical grade and mostly purchased from Sigma chemicals, (India). Temperatures during the experiment were 25±3°C during the day. Investigations for antioxidants performed for various parameters by using their methodology, described principle in following manner:

Assay of Superoxide dismutase:

The rate of autoxidation of epinephrine or sensivity of autoxidation which inhibited by superoxide dismutase. The avaibility of superoxide dismutase enzyme, capable to removing superoxide radicals from reactant mixture by catalyzing its dismutation of O^{2-} to H_2O_2 . Supeoxide ione (O^2) generated by xanthin oxidase reaction which oxidized epinephrine to adrenochrome. The production of adrenochrome increased with increasing concentration of epinephrine which measured by using colorimeter at 480

nm.(11)

Assay of glutathione reductase:

Glutathione reductase catalyses the reduction of glutathione (GSSG) in the presence of NADPH which is oxidised to NADP⁺. The decrease in absorbance at 340 nm is measured (by using colorimeter) and it is directly proportional to the glutathione reductase activity in sample.⁽¹²⁾

Assay of catalase:

The method based on the fact that dichromate in acetic acid is reduced to chromic acid when heated in the presence of hydrogen peroxide with the formation of perchromate (acid) as an unstable intermediate. The chromic acetate thus produced is measured colorimetrically at 570 nm. The reaction stopped at a particular time by the adding dichromate acetic acid mixture and the remaining hydrogen peroxide is determined by measuring chromic acetate colorimetrically after heating the reaction mixture.⁽¹³⁾

Assay of ascorbic acid:

Ascorbic acid was giving coloured complex with presence of dye 2, 4-dichlorophenol indophenols in an alcoholic acidic medium (n-amyl alcohol, m-Phosphoric acid) which measured at wavelength of 546 nm by using colorimeter.⁽¹⁴⁾

Assay of total -carotene:

Oxidation of linoleic acid occur when react with oxygenated water. The oxidative losses of -carotene were used to assess the antioxidation ability of the <u>Aloe vera leaf gel</u>

(AVLG) fractions. An absorbance at 470 nm wavelength was recorded by using colorimeter.^(15, 16)

Observation:

Table: 1. Comparative study of antioxidants values of *Aloe vera* between (T–5) Control (Sandy coastal soil: Golden sand₍₁₋₁₎ and (T–1) Sandy coastal soil: Golden sand: Farm yard manure₍₁₋₁₎ (One year plant).

S. No.	Aloe vera gel extract parameters	(T–5) Con Golden sa No. of	-5) Control (Sandy coastal soil: olden sand _{4.11} o. of Antioxidant value		(T–1) Sandy coastal soil: Golden sand: Farm yard manure No. of Antioxidant value			t–test	P-value
1.	SOD (Unit/ma.protein/ml)	leaves	38–42	39.89±1.55	leaves	40–55	47.80±5.33	8.62	P<0.0001
2.	GSH-R (Unit/mg protein/ml)	36	13.04– 14.03	13.46±0.41	46	14.08– 14.50	14.23±0.14	11.8	P<0.0001
3.	Catalase (Unit/mg protein/ml)		9.60– 10.10	9.70±0.16		10.10– 11.50	10.83±0.44	13.3	P<0.0001
4.	Ascorbic acid (mg/ml)		0.41– 0.45	0.43±0.01		0.41– 0.44	0.42±0.01	1.3	0.19 _(NS)
5.	Total –carotene (mg/ml)		0.031– 0.035	0.033±0.001		0.034– 0.037	0.035±0.001	9.13	P<0.0001

Note: P<0.0001(Extremely statistically significant). SOD; Superoxide dismutase, GSH–R; Glutathione reductase.

Table: 2.	Comparative study of	of antioxidants	values of	Aloe vera	between (T–6)	Control	(Heavy black	cotton s	oil: Gold-
en sand _{(1:}	₁₎ and (T–3) Heavy bl	ack cotton soil	: Golden s	and: Farm	yard manure _{(1:}	1:1) (One	year plant).		

S. No.	Aloe vera gel extract parameters	(T–6) Control (Heavy black cotton soil: Golden sand _(1:1)			(T-3) Heavy black cotton soil: Golden sand: Farm yard ma- nure			t-test	P-value
		No. of	Antioxidant value		No. of	Io. of Antioxidant value			
		leaves	Range	Mean±SD	leaves	Range	Mean±SD		
1.	SOD (Unit/mg protein/ml)		39–43	40.62±1.48		42–58	50.49±4.94	12.42	P<0.0001
2.	GSH-R (Unit/mg protein/ml)		13.08– 14.60	13.78±0.51		14.30– 14.80	14.55±0.17	9.61	P<0.0001
3.	Catalase (Unit/mg protein/ml)	42	9.70– 10.30	10.00±0.20	45	11.60– 12.40	12.02±0.24	41.77	P<0.0001
4.	Ascorbic acid (mg/ml)		0.44– 0.47	0.45±0.012		0.43– 0.46	0.44±0.011	2.94	P<0.001
5.	Total –carotene (mg/ml)		0.032– 0.036	0.034±0.001		0.036– 0.039	0.037±0.001	12.61	P<0.0001

Note: P<0.0001(Extremely statistically significant), P<0.001 (Very statistically significant).

Volume : 6 | Issue : 3 | March 2016 | ISSN - 2249-555X | IF : 3.919 | IC Value : 74.50

SOD; Superoxide dismutase, GSH–R; Glutathione reductase.

Results and Discussion:

The Aloe vera plant contains many various bioactive compounds including antioxidants (SOD, glutathione reductase, catalase, ascorbic acid and - carotene) existed in different parts of the plant.⁽¹⁷⁾ Antioxidant is a bioactive molecule which neutralizes harmful Reactive Oxygen Species (ROS), Reactive Nitrogen Species (RNS) and Reactive Chlorine Species (RCS) that cause damage to living cells.⁽¹⁸⁾

Farm yard manure did affect all measured qualitative characteristics while previous research indicated that nutrient solution with the highest level of nitrogen increased *Aloe vera* vegetative growth. Therefore, without any negative effect on qualitative indices we are able to produce the highest level of vegetative growth in *Aloe vera*. In other hand we may be possible to optimize for higher yields of target organs as mentioned previously.⁽¹⁹⁾

The major groups of phytochemicals that have been suggested as a natural source of antioxidants may contribute to the total antioxidant activity of plant materials including polyphenols, carotenoid and traditional antioxidant vitamins such as vitamin C and E. Antioxidant is any substance that when present at low concentration compared to those of an oxidisable substrate significantly delays or prevents oxidation of that substrate.⁽²⁰⁾ In fact, many authors have reported a direct relationship between total phenolic content and antioxidant activity in numerous seeds, fruits and vegetables.⁽²¹⁾ Previous study reported that antioxidant activity of plant material is very well correlated with the content of phenolic compounds.⁽²²⁾ Contribution of phenolic compounds is one of the mechanisms of the overall antioxidant activities. This mainly due to their redox properties involve in the plant material. Generally, the mechanisms of phenolic compounds for antioxidant activity are neutralizing lipid free radicals and preventing decomposition of hydroperoxides into free radicals.(23) The study clearly indicates that it is vital to measure the antioxidant activity using various radicals and oxidation systems and to take both phenolic content and antioxidant activity into account while evaluating the antioxidant potential of plant extracts. The results obtained in this work have considerable value with respect to the antioxidant activities of the selected parts of the Aloe vera. The author previously reported that D. salina (algae) accumulated large amount of carotenoids, -tocopherol and ascorbic acid. Enhanced the activities of the antioxidant enzymes when grown under high light intensity, in media containing high salt concentration and/or limiting nitrogen.(24)

In this study exposed of Aloe vera from high Sun light field caused significant increase in the amount of carotenoids, as compared with unexposed cells. The results revealed that significant higher growth of Aloe vera leave antioxidants (superoxide dismutase, glutathione reductase, catalase and total -carotene) in (T-1) one year (P<0.0001) plants when compared to (T-5) control one year plants (Table: 1). Thus, a positive relationship between carotenoids content and relief from high Sun light field was observed. Also, the results showed that in Sun light field exposed cells, the carotenoids biosynthesis is shifted toward prevents formation of peroxidation processes in liposomes much better than -carotene.⁽²⁵⁾ Furthermore, these results are evidence for a mechanism for protecting the cells against irradiation damage. The Aloe vera has developed defense system against photo oxidative damage by antioxidative mechanisms to detoxify and eliminate these highly reactive oxygen species. These antioxidant defense system includes hydrophobic (carotenoids & -tocopherol) and hydrophilic antioxidant (ascorbic acid & glutathione) and antioxidant enzymes likes superoxide dismutase (SOD), catalsae (CAT), glutathione reductase.⁽²⁶⁻²⁹⁾

In this study exposed of farm yard manure in black soil caused significant (P<0.0001) increase in the amount of Superoxide dismutase, Catalase, Glutathione reductase and total -carotene (T-3) one year plants as compared with (T-6) control one year plants. Significant (P<0.001) increased ascorbic acid was found in (T-3) one year plants as compared with (T-6) control one year plants (Table: 2). Evidence suggests that drought causes oxidative damage through generation of oxygen radicals or inhibition of antioxidant systems in plant.⁽³⁰⁻³²⁾ Drought related physiological changes such as a decrease in leaf water result in limited CO₂ availability to the channeling of reducing equivalents to the production of active oxygen species rather CO, fixation. Among the four major active oxygen species [superoxide radical (O⁻²), hydrogen peroxide (H₂O₂), hydroxyl radical (OH) and singlet oxygen (O₂)] are most active toxic and destructive. Hydrogen peroxide can be produced by either dismutation of O⁻² by SOD or photorespiration and catalase hydrolysed H₂O₂ to water molecule.⁽³³⁾

Conclusion:

The study of the antioxidant and antioxidant enzymes revealed that activities of these molecules were affected by foliar application of farm yard manure. Antioxidants enzymes are implicated in a variety of physiological processes including ethylene biogenesis, cell development, membrane integrity, response to injury, disease resistance. Antioxidants enzymes have multiple roles in different aspects of plant metabolism and are known to be implicated in plant differentiation and in the response against environmental stress.

References:

- Abdullaev F. I., R. R. Luna, B. V. Roitenburd, A. J. Espinosa. (2000). Pattern of childhood cancer mortality in Mexico. Arch. Med. Res. 31; 526– 531.
- Reynolds T. and A. C. Dweck. (1999). Aloe vera leaf gel: a review update. J. Ethnopharmacol. 68; 3–37.
- Heggers J., et al. (2006). Beneficial effects of aloe in wound healing. *Phytother Res.* 7; S48–S52.
- Vazquez B., et al. (1996). Anti inflammatory activity of extracts from aloe vera gel. J Ethnopharmacol. 55; 69–75.
- Saoo K., et al. (1996). Antiviral activity of aloe extracts against cytomegalovirus. *Phytother Res.* 10; 348–350.
- Fahim M. S., Wang M. (1996). Zinc acetate and lyophilized Aloe barbadensis as vaginal contraceptive. Contraception. 53; 231–236.
- Danhof I. (1991). Potential benefits from orally injested internal aloe vera gel. International Aloe Science Council Tenth Annual Aloe Scientific Semina; Irving, Texas. Aliment Pharmacol Ther. 14 (1); 18–25.
- Zhang L., Tizard I. R. (1996). Activation of a mouse macrophage cell line by acemannan: the major carbohydrate fraction from Aloe vera gel. *Immunopharmacology*. 35; 119–128.
- DeFeudis F. V., Papadopoulos V., Drieu K. (2003). *Ginkgo biloba* extracts and cancer: a research area in its infancy. *Fundam Clin Pharmacol.* 17; 405–417.
- Takeoka G. R., Dao L. T. (2003). Antioxidant constituent of almond [Prunus dulcis (Mill.) D.A. Webb.] Hulls. J Agric Food Chem. 51; 496–501.
- Misra H. P. and Fridovich I. (1972). The role of superoxide anion in the autooxidation of epinephrine and a simple assay for SOD. J Biol Chem. 247; 3170–3175.
- Bergmayer H. U. (1963). 'Methods of Enzymatic Analysis'. New York Academic Press. pp875–879.
- 13. Sinha K. A. (1972). Colorimetric assay of catalase. Analytical Biochemis-

RESEARCH PAPER

try. 47; 389–394.

- Chinoy J. J. (1962). Formation and utilization of ascorbic acid in the shoot apex of Wheat. As factor of growth and development. *Ind. J. Plant Physiol.* 5; 172–201.
- 15. S. Chevolleau, A. Debal, E. Ucciani. (1992). Rev. Fr. Corps Gras. 39 (1-2); 3–5.
- A. Moure, D. Franco, J. Sineiro, H. Dominguez, M. J. Nunez, J. M. Lema. (2000). J. Agr. Food Chem. 48; 3890–3895.
- Joseph B., Raj S. J. (2010). Pharmacognostic and phytochemical properties of Aloe Vera Linn. - an overview. International Journal of Pharmaceutical Sciences Review and Research. 4 (2); 106–110.
- Zaveri N. T. (2006). Green tea and its polyphenolic catechins: Medicinal uses in cancer and non cancer applications. *Life Sciences*. 78; 2073– 2080.
- Salighehdar F., Sedaqat-Hoor S., Olfati J. (2013). Effects of four nutrient solutions on vegetative traits of *Aloe vera* L. cv. Austin at six harvest periods. *EJGCST*. 4; 15–27.
- Yang J., Liu R. and Halim L. (2009). Antioxidant and anti proliferative activities of common edible nut seeds. *Food Science and Technology*. 42; 1–8.
- Velioglu Y. S., Mazza G. and Oomah B. D. (1998). Antioxidant activity and total phenolics in selected fruits, vegetables, and grain products. *Journal of Agricultural and Food Chemistry*. 46; 4113–4117.
- Li B. B., Smith B. and Hossain Md. M. (2006). Extraction of phenolics from citrus peels: I. Solvent extraction method. Separation and Purification Technology. 48; 182–188.
- El-Baz F. K., M. A Aboul-Enein, G. S. El-Baroty, A. M. Youssef and H. H. Abd El-Baky. (2002). Accumulation of antioxidant vitamins in Dunaliella salina. Online J. Biol. Sci. 2; 220–223.
- Gotz T., U. Windhovel, P. Boger and G. Sandman. (1999). Protection of photosynthesis against ultraviolet-B radiation by carotenoids in transformants of the cyanobacterium Synechococcus PCC79421. *Plant Physiol.* 120; 599–604.
- Rao M. V., G. Paliyath and D. P. Ormrod. (1996). Ultraviolet-B and ozoneinduced biochemical changes in antioxidant enzymes of *Arabidopsis thaliana*. *Plant Physiol*. 110; 125–136.
- Malanga G., G. Calmanovici and S. Puntarulo. (1997). Oxidative damage to chloroplasts from *Chlorella vulgaris* exposed to Ultraviolet -radiation. *Physiol. Plant.* 101; 455–62.
- Malanga G. and S. Puntarulo. (1995). Oxidative stress and antioxidant content in *Chlorella vulgaris* after exposure to ultraviolet -radiation. *Physiol. Plant.* 94; 672–679.
- Rijstenbil J. W. (2002). Assessment of oxidative stress in the planktonic diatom *Thalassiosira pseudonana* in response to UV-A and UV-B radiation. J. Plankton Res. 12; 1277–1288.
- Hounsome N., Hounsome B., Tomos D., Edwards–Jones G. (2008). Plant metabolites and nutritional quality of vegetables. J Food Sci. 73(4); 48–65.
- Jagtap V. and Bhargava S. (1995). Variation in the antioxidant metabolism of drought tolerant and drought susceptible varieties of Sorghum bicolor (L.) Moench. exposed to high light, low water and high temperature stress. J. Plant Physiol. 145; 195–197.
- Zhang J. and Kirkham M. B. (1994). Drought-stress induced changes in activities of superoxide dismutase, catalase and peroxidase in wheat species. *Plant Cell Physiol.* 35; 785–791.
- Zhang J. and Kirkham M. B. (1996). Enzymatic responses of the ascorbate glutathione cycle to drought in sorghum and sunflower plants. *Plant Sci.* 113; 139–147.
- Mittler R. and Zilinskas B. A. (1994). Regulation of pea cytosolic ascorbate peroxidise and other antioxidant enzymes during the progression of drought stress and following recovery from drought. *The Plant Journal.* 5; 397–405.