Original Reser	Arch Paper Volume - 7 Issue - 8 August - 2017 ISSN - 2249-555X IF : 4.894 IC Value : 79.96 Physiology
1001 * 40100	PLATELET MORPHOLOGY IN DIABETICS & NORMAL HEALTHY ELDERLY INDIVIDUALS
Dr. B. Syamala devi	Associate Professor, Physiology Department, Mahavir institute of medical sciences, Vikarabad.
Dr. P. Jayanth Kumar	Assistant Professor, Physiology Department, Alluri sitarama raju academy of medical sciences (ASRAM), Eluru.
ABSTRACT BACL of the documented several different OBJECTIVE: This study is MATERIALS & METHOD was performed by REES-EC weight were measured with t was done by Microsoft office BESUUTS: We evaluated tot	 KGROUND: Diabetic mellitus is considered to be a prothrombotic state with chronic platelet activation, activation coagulation system & decreased fibrinolytic potential. A number of studies performed during the past decades have platelet function alternations in patients with DM. berformed to assess the platelets morphology in control & diabetics. DS: This study is performed in Alluri Sita Rama Raju academy of medical sciences (ASRAM), Eluru. Platelet count KER method in dept of physiology, ASRAM. Platelet morphology is performed through Thyrocare. Height and he subject in light clothes without shoes and body mass index (BMI) (kg/m2) was calculated. Statistical analysis 2010. Predefined inclusion and exclusion criteria were set for selection of the study group. Blue of Complex performes. The average considered is between 50.60 (wars: We obtained statistically significant results).

RESULTS: We evaluated totally of 60 male persons. The average considered is between 50-60 years. We obtained statistically significant results. There is a significant increase in fasting blood glucose, platelet distribution width, mean platelet volume & platelet count in study group people as compared with control.

CONCLUSION: Factors that contribute directly to greater platelet reactivity include metabolic abnormalities such as hyperglycemia and hyperlipidemia, both insulin resistance (relative insulin deficiency) and absolute insulin deficiency, as well as associated conditions such as oxidative stress, inflammation, and endothelial dysfunction. Change in platelet structure & volume is observed in diabetes mellitus people. Platelet structure & volume should be monitored frequently & proper care should be taken.

KEYWORDS : Fasting Blood Glucose (FBG), Platelet Distribution Width (PDW), Mean Platelet Volume (MPV), Platelet Count (PC), Diabetis mellitus (DM).

INTRODUCTION:

Hyperglycemia is the diagnostic hallmark finding in diabetes mellitus and is associated with macrovascular disease even in the prediabetic stage. Hyperglycemia, particularly postprandial, plays a significant role in the DM-associated development of cardiovascular disease as well as the DM prothrombotic state¹⁻² Exposure of platelets to hyperosmolar solutions also causes increased reactivity, suggesting that hyperglycemia may have a direct osmotic effect³. Both chronic and acute hyperglycemia causes in vivo activation of protein kinase C (PKC), a transduction pathway mediator for many pro-aggregatory platelet agonists⁴.

Advanced glycation end products (AGEs) cause externalization of platelet membrane phosphatidylserine that leads to surface clotting factor activation and so directly enhance the thrombogenic state⁵ Similarly, the platelets of patients with diabetes have increased glycation levels of surface membrane proteins which cause decreased membrane fluidity and increased platelet sensitivity to agonists⁶⁻⁷. Insulin can directly regulate platelet function via a functional insulin receptor (IR) found on human platelets⁸. The effects of hyperinsulinemia on platelets, however, are complex and disparate between normal individuals and patients with insulin resistance. In vitro experiments using platelets from healthy nonobese individuals reveal that binding of insulin to its receptor causes magnesium to translocate into the platelet and is associated with decreased thrombininduced platelet aggregation and reduced production of proaggregatory thromboxane B2°. Binding of insulin to the IR leads to activation of the insulin receptor substrate 1 (IRS-1) through tyrosine phosphorylation which initiates association with the Ga-subunit. The result is reduced Gactivity that impairs tonic cAMP suppression, and thus leads to increased cAMP intraplatelet levels, blunting of P2Y12 signaling and reduced platelet activity¹⁰

MATERIALS & METHODS:

This study was conducted at ASRAM Hospital, Eluru in Central Lab. 30 persons were considered in each group. Totally, study was conducted on 60 males and having age 50–60 years.

In this study we compared serum blood glucose levels at early morning before breakfast in control & study group men. Mean platelet volume, platelet distribution width & platelet count tests were performed in study & control group men. We excluded diabetic, obese and any chronic disordered women. Height and weight were with the subject in light clothes without shoes and Body Mass Index (BMI) was calculated kg/m2.

The considered subjects were interviewed and basic history was taken into consideration. The subjects were given proper precautions and self-consent form was taken before drawing blood.

RESULTS

We evaluated totally of 60 male persons. The average considered is between 50-60 years. We obtained statistically significant results. In control, the fasting blood sugar levels are within normal limits. We considered study group people based on their fasting blood sugar levels. If the fasting blood sugar levels exceed 250mg/dl, we excluded them. There is a significant increase in fasting blood glucose levels, platelet distribution width, mean platelet volume & platelet count in study group people as compared with control.

Table-1

S. NO	VARIABLE	Control MEAN±STD EV	Study Group MEAN±STDE V	"p" VALUE
1	Fasting Blood Glucose Levels	110±5.65*	213.96±41.33*	0.05*
2	Platelet Distribution Width	11.89±0.53*	13.66±1.38*	0.05*
3	Mean Platelet Volume	7.53±0.34*	8.08±0.43*	0.05*
4	Platelet Count in Men	2.16±0.22*	3.21±0.41*	0.05*





108

Volume - 7 | Issue - 8 | August - 2017 | ISSN - 2249-555X | IF : 4.894 | IC Value : 79.96

DISCUSSION

Platelets from subjects with diabetes exhibit increased reactivity. Diabetes is associated with systemic inflammation and oxidative stress that may contribute to increased platelet reactivity. Superoxide has been shown to increase platelet reactivity¹². Insulin antagonizes the effect of platelet agonists such as collagen, ADP, epinephrine, and platelet-activating factor¹³. This antagonism is mediated by activation of an inhibitory G protein by insulin receptor substrate (IRS)-1¹⁴. Hyperglycemia can increase platelet reactivity by inducing nonenzymatic glycation of proteins on the surface of the platelet. Such glycation decreases membrane fluidity and increases the propensity of platelets to activate¹⁵. Osmotic effect of glucose is a second mechanism by which hyperglycemia can increase platelet reactivity¹⁶. We found that brief exposure of platelets in vitro to hyperglycemia or a similar concentration of mannitol increased their reactivity. Activation of protein kinase C is a third mechanism by which hyperglycemia can increase platelet reactivity. Protein kinase C is an essential mediator of platelet activation17

A large body of evidence from animal models and patient studies indicates that redox stress plays a major role in the pathogenesis of vascular complications of diabetes. There is convincing evidence linking decreased vascular NO production coupled with the overproduction of reactive oxygen species (ROS) and the generation of potent oxidants such as peroxynitrite (ONOO) to altered platelet function. Although the regulation of a platelet NOS is currently controversial, insulin-induced cyclic GMP production in platelets from subjects with diabetes is reported to be attenuated, and agonistinduced platelet aggregation becomes insensitive to NOS inhibitors¹⁸. The generation of ROS in diabetes may either lead to or result from platelet activation, suggesting that oxidative stress and platelet activation may be closely interrelated. Indeed, superoxide anions and hydrogen peroxide, which are both reported to play an important role in platelet activation, are continuously produced in these cells, and diabetes is associated with reduced platelet antioxidant levels¹⁹. High concentration of ROS can modify platelet function in different ways; e.g., hydrogen peroxide elevates protein tyrosine phosphorylation by activating Bruton's tyrosine kinase, the Janus kinases, and the SRC family tyrosine kinases²⁰⁻²¹. The results of a recent clinical study revealed that platelet responsiveness to the nitrovasodilator sodium nitroprusside was increased by insulin administration to patients with acute coronary syndrome, which is an insulin-resistant state²². Despite the proven benefits of insulin, optimal protection against the cardiovascular complications of diabetes has been obtained when insulin was used in combination with oral antidiabetes drugs, such as the thiazolidinediones and biguanides²³

CONCLUSION

Diabetes is associated with increased platelet reactivity. Factors that contribute directly to greater platelet reactivity include metabolic abnormalities such as hyperglycemia and hyperlipidemia, both insulin resistance (relative insulin deficiency) and absolute insulin deficiency, as well as associated conditions such as oxidative stress, inflammation, and endothelial dysfunction. Although antiplatelet therapy is necessary to suppress increased platelet reactivity, control of hyperglycemia with regimens that decrease insulin resistance and prevent apoptosis of pancreatic β-cells should decrease platelet reactivity and enhance the efficacy of antiplatelet therapy by addressing root causes of increased reactivity. Change in platelet structure & volume is observed in diabetes mellitus people. Platelet structure & volume should be monitored frequently & proper care should be taken.

REFERENCES:

- M. Tominaga, H. Eguchi et al., Impaired glucose tolerance is a risk factor for cardiovascular disease, but not impaired fasting glucose: the Funagata diabetes study,"Diabetes Care 1999; vol. 22, no. 6, pp. 920–924.
- P. J. Lefebvre. Glucose metabolism and the postprandial state. European Journal of Clinical Investigation, 1999; vol. 29, supplement 2, pp. 1–6. 2
- V. R. Vaidyula, A. K. Rao et al., Effects of hyperglycemia and hyperinsulinemia on circulating tissue factor procoagulant activity and platelet CD40 ligand. Diabetes, 2006; 3. vol. 55, no. 1, pp. 202–208.
- F. K. Keating, B. E. Sobel, D. J. Schneider, Effects of increased concentrations of characteristics of the second seco
- R. Assert, G. Scherk et al., Regulation of protein kinase C by short term hyperglycaemia
- Reference of the second sec 6
- P. D. Winocour, C. Watala et al., Decreased platelet membrane fluidity due to glycation or acetylation of membrane proteins. Thrombosis and Haemostasis, 1992; vol. 68, no. 5,

pp. 577–582

- 8 C. Watala. M. Boncer et al., Platelet membrane lipid fluidity and intraplatelet calcium mobilization in type 2 diabetes mellitus. European Journal of Haematology, 1998; vol. 61, no. 5, pp. 319-326.
- C. Falcon, G. Pfliegler et al., The platelet insulin receptor: detection, partial 9 Characterization, and search for a function," Biochemical and Biophysical Research Communications, 1988; vol. 157, no. 3, pp. 1190–1196.
 D. L. Hwang, C. F. Yen, J. L. Nadler. Insulin increases intracellular magnesium transport
- in human platelets, Journal of Clinical Endocrinology and Metabolism, 1993; vol. 76, no. 3, pp. 549-553.
- M. Trovati, G. Anfossi, P. Massucco et al., Insulin stimulates nitric oxide synthesis in human platelets and, through nitric oxide, increases platelet concentrations of both Juanosine-3',5'-cyclic monophosphate and adenosine-3',5'-cyclic monophosphate. Diabetes, 1997; vol. 46, no. 5, pp. 742–749. I. A. Ferreira, K. L. Eybrechts et al., IRS-1 Mediates Inhibition of Ca2+ Mobilization by
- Insulin via the Inhibitory G-protein Gi. Journal of Biological Chemistry, 2004; vol. 279, no. 5, pp. 3254–3264. Handin RI, Karabin R, Boxer GJ. Enhancement of platelet function by superoxide anion. J Clin Invest. 59: 959-965, 1977
- Westerbacka J, Yki-Järvinen H et al., Inhibition of platelet-collagen interaction: an in vivo action of insulin abolished by insulin resistance in obesity. Arterioscler Thromb 13 Vasc Biol. 22: 167-172. 2002.
- Ferreira IA, Eybrechts KL et al., IRS-1 mediates inhibition of Ca2+s mobilization by 14.
- Vision and the protein RD et al. (Normality in the relation in the relation of 15. platelet membranes from diabetic and control subjects. Thromb Haemost 67: 567-571, 1992
- Keating FK, Sobel BE et al., Effects of increased concentrations of glucose on platelet 16. reactivity in healthy subjects and in patients with and without diabetes. Am J Cardiol. 92: 1362-1365, 2003.
- 17. Assert R, Scherk G et al., Regulation of protein kinase C by short term hyperglycaemia in human platelets in vivo and in vitro. Diabetologia. 44: 188–195, 2001. 18
- Trovati M, Anfossi G. Influence of insulin and of insulin resistance on platelet and vascular smooth muscle cell function. J Diabetes Complications. 16: 35–40, 2002. 19
- Seghieri G, Di Simplicio P et al., Platelet antioxidant enzymes in insulin-dependent diabetes mellitus. Clin Chim Acta 309. 19–23, 2001. Abe J, Takahashi M et al., c-Src is required for oxidative stress-mediated activation of big mitogen-activated protein kinase 1 (BMK1). J Biol Chem. 272: 20389-20394,
- Redondo PC, Ben-Amor N et al., Ca2+-independent activation of Bruton's tyrosine 21. kinase is required for store-mediated Ca2+ entry in human platelets. Cell Signal. 17: 1011-1021, 2005.
- Worthley MI, Holmes AS et al., The deleterious effects of hyperglycemia on platelet function in diabetic patients with acute coronary syndromes: mediation by superoxide 22. production, resolution with intensive insulin administration. J Am Coll Cardiol. 49: 304–310, 2007.
- Randriamboavonjy V, Pistrosch F et al., : Platelet sarcoplasmic endoplasmic reticulum Ca2+-ATPase and μ -calpain activity are altered in type 2 diabetes mellitus and restored by rosiglitazone. Circulation 117: 52–60, 2008.