



## Efficacy of herbal growth promoters in augmenting the performance of broilers.

### KEYWORDS

Herbal growth promoter, digestibility co-efficient, performance parameters.

**H.F. Ahmed**

College of Veterinary Science, Khanapara, A.A.U

**K. Ravikanth**

R&D Deptt. Ayurved, Baddi, Himachal Pradesh

**S.Maini**

R&D Deptt. Ayurved, Baddi, Himachal Pradesh

**A.Borthakur**

R&D Deptt. Ayurved, Baddi, Himachal Pradesh

### ABSTRACT

A total of 240 day old commercial broiler chicks were procured and divided randomly into 4 treatment groups (T0 to T3) of 60 chicks each. Group T0 (n=60) was kept as negative control. Group T1 (n=60) was treated with basal ration supplemented with AV/AGP/10 @250g/ton (M/S Ayurved Ltd.). Group T2 (n=60) was treated with basal diet supplemented with Herbiotic @500g/ton. Group T3 (n=60) was treated with basal ration supplemented with Salinomycin @500g/ton. Parameters viz. feed trials and metabolic trials were evaluated. Results revealed improved body weight and better FCR in group T1. The mean digestibility coefficients for dry organic nutrients were improved in all supplemented groups. Nitrogen retention % was significantly better in group T1. Carcass characteristics were improved in the supplemented groups. Duodenal villous height: crypt depth ratio was higher in herbal supplement groups. Mortality rate percentage was decreased after product supplementation. Thus, it can be inferred that herbal growth promoters can significantly enhance broiler performance.

### Introduction

Broiler rearing has become one of the most lucrative ventures given the profitability, short time period of rearing and increasing social acceptability. Use of growth promoters has now become routine to help attain the genetic potential for the faster growth rate imbibed in the present day broiler chickens. They help in efficient utilization of dietary nutrients to optimal level for growth, which otherwise is not achieved by inherent digestive capacity of the birds. Growth promoters are chemical and biological substances which are added to livestock food with the aim to improve the growth of chickens in fattening, improve the utilization of food and in this way realize better production and financial results (Peric, 2009). Growth promoters are getting popularity as feed additives due to their beneficial effect on gut health and immunity, and growth performance (Panda et al., 2009). Growth stimulants as feed additives are added to poultry diet to enhance growth rate and the economic meat production (Bunyan et al., 1997). The animal feed manufacturers are exposed to increasing consumer pressure to reduce the use of antibiotic growth promoters as feed additive and find alternatives to antibiotic growth promoters in poultry diets (Newman, 1997; Hertrampf, 2001; Humphrey et al., 2002). In Europe, research on plant extracts as alternatives to the use of antibiotics as growth promoters has significantly increased. Recently, a number of scientific studies has concentrated on the bactericidal and bacteriostatic effects of various herbs and plant extracts (Dorman and Deans, 2000; Tucker, 2002). It has also been demonstrated that herbs and herbal products have a positive effect on broiler growth performance (Guo et al., 2000). Plant active principles are chemical compounds present in the entire plant or in specific parts of the plant that confers them therapeutic activity or beneficial effects (Martins et al., 2000). As more and more people are beginning to repose their faith in herbal medicine, it is pertinent to carry out further research towards that end and back up our assertions with strong scientific finding. There has been a paradigm shift in the approach towards herbal medicines and their acceptance and penetration into various treatment regimes has increased manifold. It is only obvious that this fervent switch to herbal medicine is only on account of its high safety margin and convincing performance. In light of the tearing need for stronger validation of these natural products, the present study has been carried out to evaluate the supplementation of natural growth promoter as a replacement of antibiotic growth promoter in improving growth performance, carcass traits and intestinal morphometry in broilers.

### Materials and Methods

A total of 240 day old commercial broiler chicks were procured, wing banded and divided randomly into 4 treatment groups (T0 to T3) of 60 chicks each. Group T0 (n=60) was kept as negative control. Group T1 (n=60) was supplemented with AV/AGP/10 @ 250g/ton (M/S Ayurved Ltd) along with basal ration. Group T2 (n=60) was supplemented with Hebiotic @500g/ ton along with basal ration. Group T3 (n=60) was supplemented with Salinomycin @500g/ton along with basal ration. Basal rations (BIS, 1992) were offered ad libitum to experimental chicks under different treatment groups during starter (1-28 days) and finisher (29-42 days) phases along with clean drinking water throughout the feeding trial; period of 42 days. Feeding trials viz. Body weight and Feed consumption were monitored during 42 days of study. Metabolic trial was also carried out. Several parameters viz. growth performance, nutrient utilization, carcass study, intestinal micrometry, incidence of diseases, mortality, economics of the experiment were evaluated during the period of study. All the data obtained were analyzed as per the standard statistical procedure (Snedecor and Cochran, 1980).

### Results

#### Body Weight and Body Weight Gain

The mean body weight of broilers in group T0 (control group) increased from 42.25 g initially to 2340.72 g (Table 1) at the end of 6<sup>th</sup> week with a total gain in body weight of 2298.38 g (Table 2). In group T1 (AV/AGP/10 treated group) the mean body weight increased from 42.08 g initially to 2527.02 g at the end of 6<sup>th</sup> week (Table 1) with a total gain in body weight of 2484.81g (Table 2). In group T2 (Herbiotic treated group), the mean body weight increased from 41.83 g initially to 2391.56 g at the end of 6<sup>th</sup> week with a total gain in body weight of 2349.66 g. In group T3 (Salinomycin treated group) the mean body weight increased from 42.08 g initially to 2439.84 g at the end of 6<sup>th</sup> week with a total gain in body weight of 2397.71g (Table 1 and Table 2).

**Table 1: Mean initial, weekly and final body weights (gm) of experimental broilers**

Tr.	Initial	1 <sup>st</sup> week	2 <sup>nd</sup> week	3 <sup>rd</sup> week	4 <sup>th</sup> week	5 <sup>th</sup> week	6 <sup>th</sup> week
T <sub>0</sub>	42.25±0.29 <sup>a</sup>	165.24±1.13 <sup>b</sup>	390.00±2.74 <sup>a</sup>	776.53±2.72 <sup>a</sup>	1244.60±3.67 <sup>a</sup>	1780.44±11.92 <sup>a</sup>	2340.72±3.41 <sup>a</sup>
T <sub>1</sub>	42.08±0.08 <sup>a</sup>	176.67±2.96 <sup>b</sup>	410.50±5.26 <sup>b</sup>	805.00±7.67 <sup>a</sup>	1310.51±8.82 <sup>b</sup>	1900.09±9.96 <sup>c</sup>	2527.02±4.08 <sup>c</sup>

T <sub>2</sub>	41.83±0.22 <sup>a</sup>	172.58±3.17 <sup>ab</sup>	396.46±0.71 <sup>ab</sup>	788.25±9.15 <sup>a</sup>	1271.92±14.14 <sup>ab</sup>	1810.25±20.21 <sup>ab</sup>	2391.56±35.02 <sup>ab</sup>
T <sub>3</sub>	42.08±0.17 <sup>a</sup>	169.33±2.32 <sup>ab</sup>	397.79±5.48 <sup>ab</sup>	771.31±9.52 <sup>a</sup>	1272.83±10.43 <sup>ab</sup>	1853.42±8.57 <sup>bc</sup>	2439.84±10.06 <sup>b</sup>

Means with different superscripts differ significantly (P≤0.05)

**Table 2: Mean weekly and total gain in body weight (gm) of experimental broilers**

Tr.	1 <sup>st</sup> week	2 <sup>nd</sup> week	3 <sup>rd</sup> week	4 <sup>th</sup> week	5 <sup>th</sup> week	6 <sup>th</sup> week	Total
T <sub>0</sub>	122.99±1.18 <sup>a</sup>	224.76±3.79 <sup>a</sup>	386.53±5.07 <sup>a</sup>	468.07±5.87 <sup>a</sup>	535.83±8.46 <sup>a</sup>	560.29±14.36 <sup>a</sup>	2298.38±7.94 <sup>a</sup>
T <sub>1</sub>	134.58±2.92 <sup>b</sup>	233.83±7.06 <sup>a</sup>	394.50±1.70 <sup>b</sup>	505.51±2.62 <sup>b</sup>	589.58±5.43 <sup>b</sup>	626.93±3.16 <sup>a</sup>	2484.81±8.52 <sup>c</sup>
T <sub>2</sub>	130.75±3.38 <sup>ab</sup>	223.88±3.88 <sup>a</sup>	391.79±1.98 <sup>a</sup>	483.67±1.48 <sup>ab</sup>	538.33±9.28 <sup>a</sup>	581.23±3.82 <sup>a</sup>	2349.66±6.24 <sup>ab</sup>
T <sub>3</sub>	127.25±2.38 <sup>ab</sup>	228.46±3.44 <sup>a</sup>	373.52±7.56 <sup>a</sup>	501.52±2.51 <sup>b</sup>	580.59±2.75 <sup>ab</sup>	586.42±1.69 <sup>a</sup>	2397.71±7.36 <sup>b</sup>

Means with different superscripts differ significantly (P≤0.05)

**Feed Consumption and Feed Consumption Ratio**

The total feed consumption of broilers was 4219.91 g in group T0, 4427.28g in group T1, 4228.14 g in T2, and 4319.24 g in T3. The maximum feed consumption was observed in group T1 (AV/AGP/10 treated group) followed by T3 (Salinomycin treated group) and T2 (Herbiotic treated group) ( Table 3). However, the FCR was also significantly better in the supplemented groups as compared to the control (Table 4).

**Table 3: Mean weekly and total feed consumption (gm) of experimental broilers**

Tr.	1 <sup>st</sup> week	2 <sup>nd</sup> week	3 <sup>rd</sup> week	4 <sup>th</sup> week	5 <sup>th</sup> week	6 <sup>th</sup> week	Total
T <sub>0</sub>	146.35±1.62 <sup>a</sup>	305.69±5.75 <sup>a</sup>	614.48±4.92 <sup>a</sup>	842.48±4.14 <sup>a</sup>	1044.91±10.50 <sup>a</sup>	1266.02±3.43 <sup>a</sup>	4219.91±10.16 <sup>a</sup>
T <sub>1</sub>	158.75±0.88 <sup>c</sup>	318.04±5.58 <sup>a</sup>	607.47±4.13 <sup>a</sup>	879.60±4.76 <sup>bc</sup>	1096.60±7.51 <sup>d</sup>	1366.69±3.64 <sup>c</sup>	4427.28±19.86 <sup>c</sup>
T <sub>2</sub>	154.25±1.75 <sup>bc</sup>	302.32±4.09 <sup>a</sup>	611.13±0.73 <sup>a</sup>	846.49±4.68 <sup>a</sup>	1016.43±5.64 <sup>b</sup>	1297.53±4.69 <sup>b</sup>	4228.14±17.15 <sup>a</sup>
T <sub>3</sub>	150.08±1.78 <sup>ab</sup>	303.82±3.35 <sup>a</sup>	590.08±1.69 <sup>a</sup>	887.79±3.97 <sup>c</sup>	1091.39±6.16 <sup>d</sup>	1295.95±3.45 <sup>b</sup>	4319.24±28.90 <sup>b</sup>

Means with different superscripts differ significantly (P≤0.05)

**Table 4: Mean weekly and total feed conversion ratio of experimental broilers**

Tr.	1 <sup>st</sup> week	2 <sup>nd</sup> week	3 <sup>rd</sup> week	4 <sup>th</sup> week	5 <sup>th</sup> week	6 <sup>th</sup> week	Total
T <sub>0</sub>	1.19±0.02 <sup>a</sup>	1.36±0.03 <sup>a</sup>	1.59±0.03 <sup>a</sup>	1.80±0.02 <sup>a</sup>	1.95±0.02 <sup>a</sup>	2.26±0.02 <sup>a</sup>	1.84±0.00 <sup>a</sup>
T <sub>1</sub>	1.18±0.03 <sup>a</sup>	1.36±0.02 <sup>a</sup>	1.54±0.05 <sup>a</sup>	1.74±0.02 <sup>a</sup>	1.86±0.03 <sup>a</sup>	2.18±0.02 <sup>a</sup>	1.78±0.01 <sup>c</sup>
T <sub>2</sub>	1.18±0.04 <sup>a</sup>	1.35±0.04 <sup>a</sup>	1.57±0.09 <sup>a</sup>	1.76±0.06 <sup>a</sup>	1.89±0.02 <sup>a</sup>	2.25±0.14 <sup>a</sup>	1.80±0.01 <sup>bc</sup>
T <sub>3</sub>	1.18±0.02 <sup>a</sup>	1.33±0.01 <sup>a</sup>	1.58±0.04 <sup>a</sup>	1.77±0.01 <sup>a</sup>	1.88±0.02 <sup>a</sup>	2.21±0.00 <sup>a</sup>	1.80±0.01 <sup>bc</sup>

Means with different superscripts differ significantly (P≤0.05)

**Mean Digestibility Coefficient**

The mean digestibility coefficient of organic nutrients have been listed in Table 5. The mean digestibility coefficient of dry matter and and crude fibre was significantly (P<0.05) higher in group T1 (70.37 and 25.08 respectively) (AV/AGP /10 treated group), mean digestibility coefficient of crude protein was higher in group T3 (70.76) (Salinomycin treated group) , the mean digestibility coefficient of ether extract varied non significantly.

**Table 5: Mean digestibility co-efficient of organic nutrients**

Tr.	Dry matter	Crude protein	Ether extract	Crude fibre
T <sub>0</sub>	65.52±0.37 <sup>a</sup>	67.51±0.28 <sup>a</sup>	81.00±0.51 <sup>a</sup>	24.26±0.26 <sup>a</sup>
T <sub>1</sub>	70.37±0.24 <sup>c</sup>	69.48±0.37 <sup>b</sup>	80.98±0.37 <sup>a</sup>	25.08±0.12 <sup>b</sup>
T <sub>2</sub>	68.77±0.29 <sup>b</sup>	69.59±0.26 <sup>b</sup>	80.94±0.20 <sup>a</sup>	23.89±0.15 <sup>a</sup>
T <sub>3</sub>	69.59±0.35 <sup>bc</sup>	70.76±0.56 <sup>c</sup>	81.20±0.46 <sup>a</sup>	25.83±0.21 <sup>c</sup>

Means with different superscripts differ significantly (P≤0.05)

**Retention % of Nitrogen, Calcium and Phosphorus**

The nitrogen retention % was highest in AV/AGP/10 supplemented group T1 (62.77%) and the retention % of calcium and Phosphorus varied non significantly in all the groups (Table 6).

**Table 6: Percentage of retention of nitrogen, calcium and phosphorus of experimental broilers**

Tr.	Nitrogen	Calcium	Phosphorus
T <sub>0</sub>	59.25±0.76 <sup>a</sup>	51.76±0.50 <sup>a</sup>	50.80±0.30 <sup>a</sup>
T <sub>1</sub>	62.77±0.96 <sup>b</sup>	51.97±0.27 <sup>a</sup>	50.98±0.28 <sup>a</sup>
T <sub>2</sub>	61.16±1.04 <sup>ab</sup>	52.07±0.37 <sup>a</sup>	51.10±0.36 <sup>a</sup>
T <sub>3</sub>	62.34±0.70 <sup>b</sup>	52.23±0.19 <sup>a</sup>	51.02±0.34 <sup>a</sup>

Means with different superscripts differ significantly (P≤0.05)

**Carcass Characteristics**

The live weight carcass weight, dressing % and abdominal fat % were highest in group T1 (AV/ AGP/10 treated group) at 2.55 kg, 1.88 kg, 73.73% and 2.18% respectively. The intestinal pH was highest in group T2 (6.8) (Herbiotic treated group). The colour of meat was light pink in all the groups (Table 7).

**Table 7: Carcass characteristics**

Tr.	Live weight (Kg)	Carcass weight (Kg)	Dressing percentage	Abdominal fat (% of carcass weight)	Intestinal pH	Colour of meat
T <sub>0</sub>	2.350±0.03 <sup>a</sup>	1.680±0.02 <sup>a</sup>	71.49±0.01 <sup>a</sup>	2.11±0.01 <sup>a</sup>	6.3±0.12 <sup>a</sup>	Light pink
T <sub>1</sub>	2.550±0.02 <sup>c</sup>	1.880±0.04 <sup>a</sup>	73.73±1.05 <sup>a</sup>	2.18±0.10 <sup>a</sup>	6.5±0.00 <sup>a</sup>	Light pink
T <sub>2</sub>	2.400±0.02 <sup>ab</sup>	1.720±0.03 <sup>ab</sup>	71.69±0.8 <sup>a</sup>	2.15±0.01 <sup>a</sup>	6.8±0.04 <sup>a</sup>	Light pink
T <sub>3</sub>	2.450±0.03 <sup>b</sup>	1.780±0.05 <sup>bc</sup>	72.65±1.32 <sup>a</sup>	2.11±0.03 <sup>a</sup>	6.6±0.06 <sup>a</sup>	Light pink

Means with different superscripts differ significantly (P<0.05)

**Intestinal Micrometry**

Villus height: crypt depth ratio of duodenum and ileum was higher in herbal supplement groups in comparison to Salinomycin and non supplemented group (Table 8). The GALT (Gut associated lymphoid tissue) was found to vary non significantly between the groups. Mortality rate decreased in the supplemented groups as compared to control group (Table 8).

**Table 8: Intestinal Micrometry**

Parameters	Dietary Treatments			
	T <sub>0</sub>	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>
<b>Duodenum</b>				
a) Villus height (µm)	753.50	790.21	787.21	781.61
b) Villus width (µm)	111.87	152.24	215.76	198.54
c) Crypt depth (µm)	94.60	97.03	96.32	96.66
d) Villus height : Crypt depth	7.97	8.14	8.17	8.09
<b>Ileum</b>				
a) Villus height (µm)	353.50	379.27	387.21	361.78
b) Villus width (µm)	108.89	110.49	116.14	106.60

c) Crypt depth (µm)	76.60	80.53	79.53	79.37
d) Villus height : Crypt depth	4.61	4.71	4.87	4.56
GALT	1-2	1-2	1-2	1-2
Incidence of diseases	Mixed bacterial infection in all the treatment groups during 2 <sup>nd</sup> and 3 <sup>rd</sup> week			
No. of mortalities	4	2	2	1
Mortality rate (percent)	6.67	3.33	3.33	1.67

**Economics of product supplementation**

Results indicate that maximum profit was made from group T1 (Rs.58.18/bird) (AV/AGP/10 treated group) followed by T3 (Rs. 53.15/bird) (Salinomycin treated group), and T2 (Rs. 51.45/bird) (Herbiotic treated group) as compared to control group (Rs 46.62/ bird)(Table 9).

**Table 9: Economics of product supplementation**

Parameters	Dietary Treatments			
	T <sub>0</sub>	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>
Cost per day old chick (Rs.)	24	24	24	24
Av. Total Feed consumed (Kg)	4.220	4.427	4.228	4.319
Av. Cost of per Kg feed (Rs.)	34	34	34	34
Cost of total feed consumed (Rs.)	143.48	150.52	143.75	146.85
Miscellaneous cost per bird (Rs.)	20	20	20	20
Av. total cost per bird (Rs.)	187.48	194.52	187.75	190.85
Av. Live weight per bird (Kg)	2.341	2.527	2.392	2.440
Market price per Kg live weight (Rs.)	100	100	100	100
Av. Total price earning per bird (Rs.)	234.10	252.70	239.20	244.00
<b>Av. Profit per bird (Rs.)</b>	<b>46.62</b>	<b>58.18</b>	<b>51.45</b>	<b>53.15</b>
Av. Profit per Kg live weight (Rs.)	19.91	23.02	21.51	21.78

**Discussion**

The increased weight gain also bears resemblance to the earlier reports of Rahman *et al.* (2012) who recorded an increase in body weight after use of growth promoters. The increase may be attributed to the antimicrobial effect of the herbs viz. *Allium sativum*, *Zingiber officinale* (Karuppiah and Rajaram, 2012) present in AV/AGP/10 which result in inhibition of intestinal bacteria leading to the reduced bacterial competition with the host for available nutrients and diminution in the level of toxic bacterial metabolites as a result of lessened bacterial fermentation resulting in the improvement of protein and energy digestibility; thereby ameliorate the performance of bird weight. The increase in digestibility coefficient may be due to presence of *Allium sativum* in AV/AGP/10. Similar result resembling the present study was also reported where the digestibility of total tract DM, CP and EE digestibility were improved (P < 0.05) by the addition of the garlic (*Allium sativum*) powder compared to that in the control diet. (Issa and Omar, 2011). The short chain fatty acids which are by products of bacterial fermentation stimulate the proliferation of epithelial cells of the bowel (Ichikawa *et al.*, 1999). The increase in the villous height: crypt depth ratio may be due attributed to bacterial fermentation which is enhanced by the herbal growth promoters.

**Conclusion**

The results indicate that the growth promoters improve overall performance in the broilers significantly aid in enhancing the productivity of broilers. Growth promoters improve the FCR, help to improve carcass characteristics, improve gut function and reduce mortality.

**References**

1. Peric L , Zikic D, Lukic M: Application Of Alternative Growth Promoters In Broiler Production Biotechnology in Animal Husbandry. (2009);25:387-397.
2. Newman KE, Devegowda G: Merging modern agriculture with the herbal revolution: possibilities for livestock production, what we do and do not know. pp. 301-306. Proc. of Alltech's 15th Annu. Symp., Biotechn. in the Feed Industry. Alltech Technical Publications. (1999) Nottingham University Press. Nicholasville, KY.
3. Herttrampjt JW: Alternative antibacterial performance promoters. Poultry Int.(2001); 40: 50-52.

4. Humphrey BD, Huang N, Klasing KC: Rice expressing lactoferrin and lysozyme has antibiotic-like properties when fed to chicks. J. Nutr. (2002); 132:1214-1218.
5. Martins ER, Castro DM, Castellani DC, Dias JE. Plantas medicinais. Viçosa, MG: UFV; 2000.
6. Dorman HJD, Deans SG: Antimicrobial agents from plants: antibacterial activity of plant volatile oils. J. Appl. Microb. (2000); 88: 308-316.
7. Tucker, LA: Evaluation of the effect of the botanical feed ingredient Apex on growing broiler performance. Poultry. Sci. Assoc. 91st Annu. Meeting, 11-14 Aug., 2002, Newark, Delaware. Poultry. Sci. 80 (Suppl. 1), 77 (Abstr.)
8. Guo F, Kwakkel RP, Verstegen MWA: The use of chinese herbs as alternative for growth promoters in broiler diets. Proc. of XII World's Poultry Cong., 20-24 Aug., 2000, Montreal, Canada.
9. Panda A, Rao SR, Raju M: Phytobiotics, a natural growth promoter for poultry. Poultry. Int., July 2009.
10. Rahman MA, Parvin MS , Sarker RR , Islam MT. Effects of growth promoter and multivitamin-mineral premix supplementation on body weight gain in broiler chickens J. Bangladesh Agril. Univ. (2012); 10(2):245-248
11. Bunyan JL, Jefeeries J, Sayers R, Gulliver AL, Coleman K: Antimicrobial substances and chick growth promotion: The growth promoting activities of antimicrobial substances included fifty two used either in therapy or as dietary additives. Br. Poultry. Sci. (1997) ; 18:283-294.
12. Ngantu HN, Keambou CT, Manfo TFP and Ndamukong KJ N: Growth Promoter Effects of Allium Sativum and Zingiber Officinale on Performances Haematological Parameters and Gut Microbiology of the Cameroon Kabir Chicken J Anim Sci Adv. (1997); 6(9): 1766-1778
13. Karuppiah P, Rajaram S: Antibacterial effect of Allium sativum cloves and Zingiber officinale rhizomes against multiple-drug resistant clinical pathogens Asian Pac J Trop Biomed. (2012); 2(8): 597-601.
14. Ichikawa H, Kuroiwa T, Inagaki A, Shineha R, Nishihira T, Satomi S, *et al.* Probiotic bacteria stimulate gut epithelial cell proliferation in rat. Digestive Diseases and Sciences, 1999;44:2119-2123.
15. Issa KJ, Omar JM: Effect of garlic powder on performance and lipid profile of broilers. Open Journal of Animal Sciences. (2012); 2:62-68.
16. Snedecor, GW and Cochran, WG: Statistical Methods. Seventh Edition. Ames Iowa: The Iowa State University Press. (1980).