



RECENT ADVANCES IN CRANIOSYNOSTOSIS AND FUTURE PROSPECTIVE

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ABSTRACT **Objective:** Craniosynostosis (CS) is the premature ossification and genetically heterogeneous in conditions. We aim to focus on recent advances with future prospective on Craniosynostosis.

Material and Methods: We observed (since 1990 to till 2017) through different databases of world literature and add our experience with recent advances and future prospective in Craniosynostosis in both the group (syndromic and non syndromic) and additional anomalies.

Results: The pathologies result from complete loss of gene function, biochemical mechanisms involving haploinsufficiency, dominant gain-of-function and recessive hypomorphic mutation (RHM). X-linked cellular interference process been implicated few of the genes involved could predicted based on their expression patterns. Genes play much wider roles in embryonic development or cellular homeostasis (CH). Here we argue that they fit into a limited number of functional modules active at different stages of cranial suture development (CSD).

Conclusion: Recent advances in neurosurgery, neuromonitoring and neurointensive care (NIC) have dramatically improved the outcome in patients affected by surgical lesions of central nervous system (CNS). Most of these techniques were applied first in the adult population; paediatric patients present a set of inherent challenges. Recently their developing and maturing neurological and physiological status, apart from the CS disease process and phenotypic pattern.

KEYWORDS : craniofacial; craniosynostosis; gene mutations; malformation; skull sutures; recent advances; prospective

INTRODUCTION

Craniosynostosis (CS) is the premature ossification of one or more skull sutures [1]. It is a clinically and genetically a group of heterogeneous congenital anomaly (HCA), affecting approximately one in 2,500 live births globally. In Indian scenario 1:1000 [2-3]. CS occurs as an isolated congenital anomaly, that is, nonsyndromic craniosynostosis (NCS) [4]. The major causes of the disease are genetic, and environmental. Other causes remain largely unknown [5]. Researcher believe that some of the midline NCS cases may be explained by two loci inheritance, approximately in 25-30% of the patients [6]. In craniofacial disorders, upper airway obstruction (UAO) is one of the primary causes for morbidity and mortality, in the neonatal period, including Pierre Robin sequence, which is high risk for obstructive sleep apnea syndrome (OSAS) [7-8-9]. Nonsyndromic craniosynostosis (NSC) is basically associated with significant learning disability, later in life [10]. This small cohort supported by the primary goal of surgery in allowing for more normalized brain growth [11]. Large sample, and correlating degree of normalization with cognitive performance in NSC, is warranted [12]. In this article, we discuss the recent advances in our understanding of the embryology of craniofacial conditions, and we focus on the use of animal models to guide rational therapies [13]. Further Genetics and biochemistry may lead for further future prospective of Craniosynostosis (CS) and other additional anomalies [14-15].

MATERIAL AND METHODS

We observed (since 1990 to till 2017) through different databases of world literature and add our experience with recent advances and future prospective in CS in both the group (syndromic and non syndromic). We provides current approaches in craniofacial surgery for treating states of bone excess and deficit, recent advances in our understanding of the molecular and cellular processes underlying craniosynostosis, a pathological state of bone excess (PSOBE) and current research efforts in cellular-based therapies (CBT) for bone regeneration and its recent advances and future prospective for craniosynostosis (CS) amongst both the group.

RESULTS

Recently, new approach to dissect the underlying causes from investigation of clinical samples, and recent advances in high-throughput DNA sequencing have dramatically enhanced the human subject as the preferred as model [16-17-18]. Most CS, investigation of mechanisms requires more conventional model organisms (CMO) [19]. In mouse, similarities in cranial suture development (CSD)

present a framework for classifying genetic causes of craniosynostosis (CS) [20]. Current understanding of cranial suture biology with molecular and developmental pathogenesis and pathologies result from complete loss of gene function (CLOGF) [21]. Biochemical mechanisms involving haploinsufficiency, dominant gain-of-function and recessive hypomorphic mutations (RHM) and X-linked cellular interference process are important [22]. Expression patterns of the genes play much wider roles in embryonic development or cellular homeostasis at different stages of cranial suture development (CSD) [23-24]. CS defining as the potential avenues for devising pharmacological approaches for new molecular targeted therapy (MTT) [25].

Development of the craniofacial region is a remarkably complex and tightly orchestrated process with genetic and environmental insults frequently results in craniofacial anomalies [26]. In our knowledge of their cell fate in etiology and pathogenesis is still scarce, limiting our efforts for preventing diseases [27]. New standardized protocols have been developed to guide clinical and surgical interventions on the most recent research advances on craniofacial conditions [28], from genomics and epigenetics to ontology and medical care are discussed with emphasis on the most common anomalies (CA) of the craniofacial region like (facial clefts, craniosynostosis, craniofacial microsomia, facial dysostosis, Robin sequence, jaw and dentition anomalies, and anterior neural tube defects) [29-30]. As a phenotypic variability and the importance of standardized terminology to better distinguish between phenotypes, new technologies for genetic diagnosis, and the use of mouse models to study these conditions, complex phenotypic and genetic aspects are highlighted [31-32]. Recent advances in neurosurgery, neuromonitoring and neurointensive care have dramatically improved the outcome in patients affected by surgical lesions of central nervous system (CNS) [33]. Although most of these techniques were applied first in the adult population, paediatric patients present a set of inherent challenges because of their developing and maturing neurological and physiological status, apart from the CNS disease process [34-35-36]. Syndromic craniosynostosis (SCS) and (CBT) exhibits variable clinical and genetic heterogeneity condition [37]. The fibroblast growth factor receptor genes (FGFR1, FGFR2, FGFR3 (encoding fibroblast growth factor receptors), TWIST1 (functions as an upstream regulator of FGFRs) and EFNB1 (gene encoding fibrillin1) [38-39-40]. Recently, advances in molecular genetics have led to a discover of other genes implicated in different craniosynostosis syndromes as a priority [41-42]. The transcription factor Twist (TWIST) plays vital roles during embryonic development

through regulating/controlling cell migration [43]. The transcriptional activities of Twist support cancer cells to disseminate from primary tumours and subsequently establish a secondary tumour growth (STG) in distant organs [44-45]. Recent advances in Twist regulation and activity, with a focus on phosphorylation-dependent Twist activity, potential upstream kinases contribution of these factors in transducing biological signals from upstream signalling complexes (USC) [46-47-48]. Recent advances in particular areas have new light on the phosphorylation-dependent regulation (PDR) of the Twist proteins promotes or suppresses among the scene [49]. The Twist activity (TA) now leading to differential regulation of Twist transcriptional targets and thereby influencing the cell fate in easiest way in the cell fate (CF) [50-51].

DISCUSSION

Recent studies have demonstrated the impressive improvements in treatment outcome with the use of tyrosine kinase inhibitors (TKIs) [52]. High-resolution genomic profiling (HRGP), of genetic alterations and gene expression has revolutionized our understanding of the genetic basis of CS [53-54] activating mutations of Janus kinases, and rearrangement of the lymphoid cytokine receptor gene CRLF2 [55]. Recent progress in elucidating the molecular and cellular mechanisms governing bone formation will have significant role in developing advanced therapies for the treatment of pathological states of bone excess and deficit (BEAD) and reengineering the bone access [56-57-58]. The health care system (HCS) are built on the evolution of technology fetal medicine technology (FMT), prenatal imaging (PI) / allow us to see and diagnose abnormalities [59-60]. The oral and maxillofacial surgeon on the fetal diagnosis and treatment team required for correct deformity [61].

Recent advances in molecular genetics have led to a better understanding of the role of specific genes (fibroblast growth factor receptor (FGFR) and Twist as the root cause of cranial bone malformations (CBMF) and osteoblast abnormalities (OA) [62-63-64]. The list of genes that are involved in CS includes those coding FGFR and a ligand of ephrin receptors (ER) [65-66]. Genes encoding transcription factors, FGFR, MSX2 and TWIST genes are equally involved in skull formation, odontogenesis, providing an explanation for associations of CS and tooth malformations [67-68]. Bone ridging seen on the ectocranial and endocranial surfaces of fused sagittal suture (FSS) pattern are not observed stenosis of coronal and lambdoid sutures [69-70]. This making it specific to sagittal suture only [69]. Thus there is a complex arrangement of the structure of the human cranium and the process of craniosynostosis, with some differences in final structure depending on the affected suture [70-71]. Fibroblast growth factor receptor (FGF) signalling pathway is involved in evolution and plays crucial roles in development of CS [72]. FGF signal pathway plays important role in suture and synchondrosis regulation (SR) [73]. FGF receptors relate to syndromic and non-syndromic CS basically located (Fgf10/Fgf2b signal loop) is critical for palatogenesis and submandibular gland formation (SMGF) through mutation [74-75].

The principle of abnormal skull growth due to restriction of skull growth at the fused sutures [76], and the realization by Moss that the sutures at the skull base are equally affected, have been the main intellectual driving forces behind the majority of cranial expansion procedures in children with craniosynostosis (CS) diseases [77-78]. The main subtypes of craniosynostosis and craniofacial dysostosis presented, including specific clinical features presently available surgical options [79].

Development and new application of various vehicles and tissue engineered constructs to deliver different cytokines, gene products and short segment DNA, to treat CS [80]. Such therapy based on gene product may be used as adjuncts to surgery and manage postoperative resynostosis [81]. Study of TWIST, FGFR-1, FGFR-2 and FGFR-3 genes in a cohort of patients with CS led to the diagnosis of Saethre-Chotzen syndrome (SCS) [82]. It is a newly diagnosed a new micro deletion disorder (MD) and reports the first example of a gene-environment interaction (GEI) [83].

Basic to Clinical approach through animal model and organism:

Many laboratories are investigating murine cranial suture biology as a model for human cranial suture development and fusion [84]. Normal murine cranial suture biology (MCSB) is very complex [85]. Evidence suggests that the dura mater provides the bimolecular

blueprints, which guide the fate of the pluripotent osteogenic fronts (POF) [86]. We have very little understanding the fundamental mechanisms of cranial suture fusion (CSF) [87]. Interestingly, recent advances in premature human and programmed murine suture fusion (PMSF) have revealed unexpected results [88]. We present recent advances in the understanding of mechanisms of CS, with particular emphasis on the biology of programmed cranial suture fusion (PCSF) in rodents [89-90]. Several surgical techniques have been described for correction of scaphocephaly [91].

Advances in the field of craniofacial surgery have allowed more extensive operative procedures [92]. For the treatment of cranial vault anomalies (CVA) aim of surgical treatment is to prevent early refusion of the parietal bones [93]. After the sagittal craniotomy associated with the widening of the biparietal diameter [94]. In children with older than 6 months, these procedures result are unsatisfactory, and provides only a partial correction of the malformation [95]. Therefore operative techniques of total cranial vault (TCV) reshaping are reported in the literature [96]. Additionally occipital remodeling is useful in young infants with a marked skull deformity. This technique will provide good results [97-98]. Recently, human genome initiative has accelerated positional cloning efforts toward identification of a number of genes responsible for human developmental anomalies (HAD) [99-100-101].

CURRENT STATE OF KNOWLEDGE AND FUTURE PROSPECTIVE

Premature fusion of the metopic suture is an uncommon form of craniosynostosis (CS) [102]. The reported with an incidence of less than 10% among the various forms of craniosynostoses [103]. The most obvious deformity associated with premature fusion of a single suture with its prominent frontal keel, narrow forehead, and close-set eyes [104]. We discuss the timing and long-term results, of surgery [105]. Frontal bone advancement (FBA) and compensatory craniofacial growth (CCG) changes in rabbits with experimental coronal suture immobilization (CSI) very nicely [106]. Clinical advances in the surgical correction of coronal suture synostosis (CSS) involve the overcorrection of a frontal bone segment (FBS) to allow for unrestricted expansion of skull [107]. Animals those underwent frontal bone advancement (FBA) exhibited normal overall craniofacial growth by 18 weeks of age, compared with control animals [108]. However, surface dysmorphology (SD) of the head in Apert syndrome known for a century [109]. A computer-assisted medical imaging technology (CAMIT) allows in vivo nondestructive "dissection." [110]. Presently, the surgical correction of brachycephalic airway syndrome (BAS) in dogs has been reported good long-term outcomes [111-112].

CONCLUSION

Recent advances have been achieved in craniofacial surgery (CS), improved the strategies for addressing states of bone excess (BE) and bone deficit (BD) in the craniofacial region (CR) are needed. The biomolecular events involved in craniosynostosis (CS and NSCS) and cellular-based bone tissue engineering (CBBTE) soon be added to the armamentarium of surgeons treating craniofacial dysmorphologies as an emerging cellular base therapy (CBT) for resolve these types of diseases as a priority in future management and early planning for patient benefit and research.

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Conflict of Interest: Nil

REFERENCES

- Kalantar Hormozi A, Mahdavi N, Foroozanfar MM, Razavi SS, Mohajerani R, Eghbali A, Mafi AA, Hashemzadeh H, Mahdavi A. Effect of Perioperative Management on Outcome of Patients after Craniosynostosis Surgery. *World J Plast Surg.* 2017; 6:48-53.
- Barik M, Bajpai M, Panda SS, Malhotra A, Samantaray JC, Dwivedi SN. Strengthening molecular genetics and training in craniosynostosis: The need of the hour. *J Neurosci Rural Pract* 2014;5:428-32
- Stricker PA, Goobie SM, Cladis FP, Haberkern CM, Meier PM, Reddy SK, Nguyen TT, Cai L, Polansky M, Szmuk P, Fiadjo J, Soneri C, Falcon R, Petersen T, Kowalczyk-Derderian C, Dalesio N, Budac S, Groenewald N, Rubens D, Thompson D, Watts R, Gentry K, Ivanova I, Hetmaniuk M, Hsieh V, Collins M, Wong K, Binstock W, Reid R, Poteet-Schwartz K, Gries H, Hall R, Koh J, Bannister C, Sung W, Jain R, Fernandez A, Tuite GF, Ruas E, Drozhinin O, Tetreault L, Muldowney B, Ricketts K, Fernandez P, Sohn L, Hajduk J, Taicher B, Burkhardt J, Wright A, Kugler J, Barajas-DeLoa L, Gangadharan M, Busso V, Stallworth K, Staudt S, Labovsky KL, Glover CD, Huang H, Karlberg-Hippard H, Capehart S, Streckfus C, Nguyen KT, Manyang P, Martinez JL, Hansen JK, Levy HM, Brzenski A, Chiao F, Ingelmo P, Mujallid R, Olutoye OA, Syed T, Benzon H, Bosenberg A; Pediatric Craniofacial Collaborative Group. Perioperative Outcomes and Management in Pediatric Complex Cranial Vault Reconstruction: A Multicenter Study from the Pediatric Craniofacial Collaborative Group. *Anesthesiology.* 2017; 126:276-287.

4. Pawar N, Padmavathy S, Maheshwari D, Ravindran M, Ramakrishnan R. OCT-documented optic atrophy in nonsyndromic craniosynostosis and lacunar skull. *J AAPOS*. 2017;21:78-81.
5. Magge SN, Snyder K, Sajja A, DeFreitas TA, Hofferr SE, Broth RE, Keating RF, Rogers GF. Identical Twins Discordant for Metopic Craniosynostosis: Evidence of Epigenetic Influences. *J Craniofac Surg*. 2017;28:14-16.
6. Cox TC, Luquetti DV, Cunningham ML. Perspectives and challenges in advancing research into craniofacial anomalies. *Am J Med Genet C Semin Med Genet*. 2013;163C:213-7.
7. Cielo CM, Montalva FM, Taylor JA. Craniofacial disorders associated with airway obstruction in the neonate. *Semin Fetal Neonatal Med*. 2016;21:254-62.
8. Carvalho FR, Lentini-Oliveira DA, Prado LB, Prado GF, Carvalho LB. Oral appliances and functional orthopaedic appliances for obstructive sleep apnoea in children. *Cochrane Database Syst Rev*. 2016 5;10:CD005520.
9. Joosten KF, Larramona H, Miano S, Van Waardenburg D, Kaditis AG, Vandenbussche N, Ersu R. How do we recognize the child with OSAS? *Pediatr Pulmonol*. 2017 ;52(2):260-271.
10. Faria AC, Rabbini-Bortolini E, Reboucas MR, de S Thiago Pereira AL, Frasson MG, Atique R, Lourenço NC, Rosenberg C, Kobayashi GS, Passos-Bueno MR, Errera FI. Craniosynostosis in 10q26 deletion patients: A consequence of brain underdevelopment or altered suture biology? *Am J Med Genet A*. 2016;170A:403-9.
11. Thwin M, Schultz TJ, Anderson PJ. Morphological, functional and neurological outcomes of craniectomy versus cranial vault remodeling for isolated nonsyndromic synostosis of the sagittal suture: a systematic review. *JBI Database System Rev Implement Rep*. 2015;13:309-68.
12. Brooks ED, Yang J, Beckett JS, Lacadie C, Scheinost D, Persing S, Zellner EG, Oosting D, Keifer C, Friedman HE, Wyk BV, Jui R, Sun H, Gary C, Duncan CC, Constable RT, Pelphey KA, Persing JA. Normalization of brain morphology after surgery in sagittal craniosynostosis. *J Neurosurg Pediatr*. 2016;17:460-8.
13. Romaneli Tavares VL, Zechi-Ceide RM, Bertola DR, Gordon CT, Ferreira SG, Hsia GS, Yamamoto GL, Ezuquina SA, Kokitsu-Nakata NM, Vendramini-Pitolli S, Freitas RS, Souza J, Raposo-Amaral CA, Zatz M, Amiel J, Guion-Almeida ML, Passos-Bueno MR. Targeted molecular investigation in patients within the clinical spectrum of Auriculocondylar syndrome. *Am J Med Genet A*. 2017;173:938-945.
14. Goodwin AF, Kim R, Bush JO, Klein OD. From Bench to Bedside and Back: Improving Diagnosis and Treatment of Craniofacial Malformations Utilizing Animal Models. *Curr Top Dev Biol*. 2015;115:459-92.
15. Sanchez-Lara PA. Clinical and Genomic Approaches for the Diagnosis of Craniofacial Disorders. *Curr Top Dev Biol*. 2015;115:543-59.
16. Ishii M, Sun J, Ting MC, Maxson RE. The Development of the Calvarial Bones and Sutures and the Pathophysiology of Craniosynostosis. *Curr Top Dev Biol*. 2015;115:131-56.
17. Kim S, Twigg SR, Scanlon VA, Chandra A, Hansen TJ, Alsabait A, Fenwick AL, McGowan SJ, Lord H, Lester T, Sweeney E, Weber A, Cox H, Wilkie AO, Golden A, Corsi AK. Localized TWIST1 and TWIST2 basic domain substitutions cause four distinct human diseases that can be modeled in *C. elegans*. *Hum Mol Genet*. 2017 Mar 27; doi: 10.1093/hmg/ddx107.
18. Whyte MP, McAlister WH, Fallon MD, Pierpont ME, Bijanki VN, Duan S, Otaifiy GA, Sly WS, Mumm S. Raine Syndrome (OMIM #259775), Caused By FAM20C Mutation, Is Congenital Sclerosing Osteomalacia With Cerebral Calcification (OMIM 259660). *J Bone Miner Res*. 2017;32:757-769.
19. De Coster PJ, Mortier G, Marks LA, Martens LC. Cranial suture biology and dental development: genetic and clinical perspectives. *J Oral Pathol Med*. 2007;36:447-55.
20. Rice DP, Rice R, Thesleff I. Molecular mechanisms in calvarial bone and suture development, and their relation to craniosynostosis. *Eur J Orthod*. 2003;25:139-48.
21. Elliott AM, Wilcox WR, Spear GS, Field FM, Steffens TS, Friedman BD, Raimo DL, Lachman RS. Osteocraniosynostosis-hypomineralized skull with gracile long bones and splenic hypoplasia. Four new cases with distinctive chondro-osseous morphology. *Am J Med Genet A*. 2006 15;140:1553-63.
22. Rice DP, Rice R, Thesleff I. Molecular mechanisms in calvarial bone and suture development, and their relation to craniosynostosis. *Eur J Orthod*. 2003;25:139-48.
23. Bae SC, Lee YH. Phosphorylation, acetylation and ubiquitination: the molecular basis of RUNX regulation. *Gene*. 2006 17;336:58-66.
24. Merrill AE, Bochkova EG, Bruggner SM, Ishii M, Pilz DT, Wall SA, Lyons KM, Wilkie AO, Maxson RE Jr. Cell mixing at a neural crest-mesoderm boundary and deficient ephrin-Eph signaling in the pathogenesis of craniosynostosis. *Hum Mol Genet*. 2006 ;15:1319-28.
25. Twigg SR, Wilkie AO. A Genetic-Pathophysiological Framework for Craniosynostosis. *Am J Hum Genet*. 2015 ;97:359-77.
26. Schock EN, Struve JN, Chang CF, Williams TJ, Snedeker J, Attia AC, Stottmann RW, Bruggner SA. A tissue-specific role for intracellular transport genes during craniofacial development. *PLoS One*. 2017 27;12:e0174206.
27. Garcia GA, Tian JJ, Apinyawasuk S, Kim S, Akil H, Sadun AA. Clues from Crouzon: Insights into the potential role of growth factors in the pathogenesis of myelinated retinal nerve fibers. *J Curr Ophthalmol*. 2016;28:232-236.
28. Sperber GH. Pathogenesis and morphogenesis of craniofacial developmental anomalies. *Ann Acad Med Singapore*. 1999;28:708-13.
29. Cohen MM Jr. Malformations of the craniofacial region: evolutionary, embryonic, genetic, and clinical perspectives. *Am J Med Genet*. 2002;115:245-68.
30. Sperber GH. The aetiopathogenesis of craniofacial anomalies. *Ann Acad Med Singapore*. 1992;21:708-14.
31. Feng W, Choi I, Clouthier DE, Niswander L, Williams T. The Ptc1(DL) mouse: a new model to study lamdboid craniosynostosis and basal cell nevus syndrome-associated skeletal defects. *Genesis*. 2013;51:677-89.
32. Czerwinski M, Kolar JC, Fearon JA. Complex craniosynostosis. *Plast Reconstr Surg*. 2011;128:955-61.
33. Rath GP, Dash HH. Anaesthesia for neurosurgical procedures in paediatric patients. *Indian J Anaesth*. 2012;56:502-10.
34. Aldridge K, Marsh JL, Govier D, Richtsmeier JT. Central nervous system phenotypes in craniosynostosis. *J Anat*. 2002;201:31-9.
35. Breik O, Mahinda A, Moore MH, Molloy CJ, Santoreneos S, David DJ. Central nervous system and cervical spine abnormalities in Apert syndrome. *Childs Nerv Syst*. 2016 ;32:833-8.
36. Chan CT, Thorogood P. Pleiotropic features of syndromic craniosynostoses correlate with differential expression of fibroblast growth factor receptors 1 and 2 during human craniofacial development. *Pediatr Res*. 1999;45:46-53.
37. Jezela-Stanek A, Krajewska-Walasek M. Genetic causes of syndromic craniosynostoses. *Eur J Paediatr Neurol*. 2013;17:221-4.
38. Marie PJ, Kaabeche K, Guenou H. Roles of FGFR2 and twist in human craniosynostosis: insights from genetic mutations in cranial osteoblasts. *Front Oral Biol*. 2008;12:144-59.
39. Anderson PJ, Cox TC, Roscioli T, Elakis G, Smithers L, David DJ, Powell B. Somatic FGFR and TWIST mutations are not a common cause of isolated nonsyndromic single suture craniosynostosis. *J Craniofac Surg*. 2007;18:312-4.
40. Taylor GM, Cooper GM, Losee JE, Mooney MP, Gilbert J. Molecular Analysis of Ephrin A4 and Ephrin B1 in a Rabbit Model of Craniosynostosis: Likely Exclusion as the Loci of Origin. *Cleft Palate Craniofac J*. 2017 Jan 30. doi: 10.1597/16-135.
41. Melville H, Wang Y, Taub PJ, Jabs EW. Genetic basis of potential therapeutic strategies for craniosynostosis. *Am J Med Genet A*. 2010;152A:3007-15.
42. Makarov R, Steiner B, Gucev Z, Tasic V, Wieacker P, Wieland I. The impact of CFNS-causing EFNB1 mutations on ephrin-B1 function. *BMC Med Genet*. 2010;11:98.
43. Passos-Bueno MR, Armelin LM, Alonso LG, Neusteine I, Sertie AL, Abe K, Pavanello Rde C, Elkis LC, Koiffmann CP. Craniosynostosis associated with ocular and distal limb defects is very likely caused by mutations in a gene different from FGFR, TWIST, and MSX2. *Am J Med Genet*. 2002;113:200-6.
44. Jadico SK, Huebner A, McDonald-McGinn DM, Zackai EH, Young TL. Ocular phenotype correlations in patients with TWIST versus FGFR3 genetic mutations. *J AAPOS*. 2006;10:435-44.
45. Passos-Bueno MR, Armelin LM, Alonso LG, Neusteine I, Sertie AL, Abe K, Pavanello Rde C, Elkis LC, Koiffmann CP. Craniosynostosis associated with ocular and distal limb defects is very likely caused by mutations in a gene different from FGFR, TWIST, and MSX2. *Am J Med Genet*. 2002;113:200-6.
46. Anderson PJ, Cox TC, Roscioli T, Elakis G, Smithers L, David DJ, Powell B. Somatic FGFR and TWIST mutations are not a common cause of isolated nonsyndromic single suture craniosynostosis. *J Craniofac Surg*. 2007;18:312-4.
47. El Ghouzzi V, Lajeunie E, Le Merrer M, Cormier-Daire V, Renier D, Munnich A, Bonaventure J. Mutations within or upstream of the basic helix-loop-helix domain of the TWIST gene are specific to Saethre-Chotzen syndrome. *Eur J Hum Genet*. 1999 ;7:27-33.
48. Johnson D, Iseki S, Wilkie AO, Morriss-Kay GM. Expression patterns of Twist and Fgf1, -2 and -3 in the developing mouse coronal suture suggest a key role for twist in suture initiation and biogenesis. *Mech Dev*. 2000 ;91:341-5.
49. El Ghouzzi V, Lajeunie E, Le Merrer M, Cormier-Daire V, Renier D, Munnich A, Bonaventure J. Mutations within or upstream of the basic helix-loop-helix domain of the TWIST gene are specific to Saethre-Chotzen syndrome. *Eur J Hum Genet*. 1999;7:27-33.
50. Xue G, Hemmings BA. Phosphorylation of basic helix-loop-helix transcription factor Twist in development and disease. *Biochem Soc Trans*. 2012;40:90-3.
51. Huang Y, Meng T, Wang S, Zhang H, Mues G, Qin C, Feng JQ, D'Souza RN, Lu Y. Twist1- and Twist2-haploinsufficiency results in reduced bone formation. *PLoS One*. 2014;9:e99331.
52. Singh HN, Rajeswari MR. Role of long purine stretches in controlling the expression of genes associated with neurological disorders. *Gene*. 2015;572:175-83.
53. Bessenyei B, Nagy A, Szakson K, Mokánszki A, Balogh E, Ujfalusi A, Tihanyi M, Novák L, Bognár L, Oláh É. Clinical and genetic characteristics of craniosynostosis in Hungary. *Am J Med Genet A*. 2015 ;167A:2985-91.
54. Flöttmann R, Knaus A, Zemojtel T, Robinson PN, Mundlos S, Horn D, Spielmann M. FGFR2 mutation in a patient without typical features of Pfeiffer syndrome--The emerging role of combined NGS and phenotype based strategies. *Eur J Med Genet*. 2015 ;58:376-80.
55. Hunger SP, Raetz EA, Loh ML, Mullighan CG. Improving outcomes for high-risk ALL: translating new discoveries into clinical care. *Pediatr Blood Cancer*. 2011;56:984-93.
56. Wan DC, Kwan MD, Kumar A, Bradley JP, Longaker MT. Craniofacial surgery, from past pioneers to future promise. *J Maxillofac Oral Surg*. 2009;8:348-56.
57. Moazen M, Peskett E, Babbs C, Pauws E, Fagan MJ. Mechanical properties of calvarial bones in a mouse model for craniosynostosis. *PLoS One*. 2015;10:e0125757.
58. Coll G, Arnaud E, Collet C, Brunelle F, Sainte-Rose C, Di Rocco F. Skull base morphology in fibroblast growth factor receptor type-2-related faciocraniosynostosis: a descriptive analysis. *Neurosurgery*. 2015;76:571-83.
59. Helfer TM, Peixoto AB, Tonni G, Araujo Junior E. Craniosynostosis: prenatal diagnosis by 2D/3D ultrasound, magnetic resonance imaging and computed tomography. *Med Ultrason*. 2016;18:378-85.
60. Guibaud L, Collardeau-Frachon S, Lacalm A, Massoud M, Rossi M, Cordier MP, Vianey-Saban C. Antenatal manifestations of inborn errors of metabolism: prenatal imaging findings. *J Inher Metab Dis*. 2017;40:103-112.
61. Costello BJ, Edwards SP, Clemens M. Fetal diagnosis and treatment of cranioaxillofacial anomalies. *J Oral Maxillofac Surg*. 2008;66:1985-95.
62. Marie PJ, Kaabeche K, Guenou H. Roles of FGFR2 and twist in human craniosynostosis: insights from genetic mutations in cranial osteoblasts. *Front Oral Biol*. 2008;12:144-59.
63. Miri S, Mittermiller P, Buchanan EP, Khosla RK. Facial twist (asymmetry) in isolated unilateral coronal synostosis: does premature facial suture fusion play a role? *J Craniofac Surg*. 2015;26:655-7.
64. Kim S, Twigg SR, Scanlon VA, Chandra A, Hansen TJ, Alsabait A, Fenwick AL, McGowan SJ, Lord H, Lester T, Sweeney E, Weber A, Cox H, Wilkie AO, Golden A, Corsi AK. Localized TWIST1 and TWIST2 basic domain substitutions cause four distinct human diseases that can be modeled in *C. elegans*. *Hum Mol Genet*. 2017 Mar 27. doi: 10.1093/hmg/ddx107.
65. Ko JM, Jeong SY, Yang JA, Park DH, Yoon SH. Molecular genetic analysis of TWIST1 and FGFR3 genes in Korean patients with coronal synostosis: identification of three novel TWIST1 mutations. *Plast Reconstr Surg*. 2012 ;129:814e-21e.
66. De Coster PJ, Mortier G, Marks LA, Martens LC. Cranial suture biology and dental development: genetic and clinical perspectives. *J Oral Pathol Med*. 2007;36:447-55.
67. Qin Q, Xu Y, He T, Qin C, Xu J. Normal and disease-related biological functions of Twist1 and underlying molecular mechanisms. *Cell Res*. 2012;22:90-106.
68. Sauerhammer TM, Patel K, Oh AK, Proctor MR, Mulliken JB, Rogers GF. Combined metopic and unilateral coronal synostoses: a phenotypic conundrum. *J Craniofac Surg*. 2014;25:437-40.
69. Nah H. Suture biology: Lessons from molecular genetics of craniosynostosis syndromes. *Clin Orthod Res*. 2000;3:37-45.
70. Anderson PJ, Netherway DJ, David DJ, Self P. Scanning electron microscope and micro-CT evaluation of cranial sutures in health and disease. *J Craniofac Surg*. 2006 ;17:909-19.
71. Lajeunie E, Catala M, Renier D. Craniosynostosis: from a clinical description to an understanding of bone formation of the skull. *Childs Nerv Syst*. 1999;15:676-80.
72. Johnson D, Wall SA, Mann S, Wilkie AO. A novel mutation, Ala315Ser, in FGFR2: a gene-environment interaction leading to craniosynostosis? *Eur J Hum Genet*. 2000 ;8:571-7.
73. Lu C, Huguley S, Cui C, Cabaniss LB, Waite PD, Sarver DM, Mamaeva OA, MacDougall M. Effects of FGFR Signaling on Cell Proliferation and Differentiation of Apert Dental Cells. *Cells Tissues Organs*. 2016;201:26-37.
74. Nie X, Luukko K, Kettunen P. FGF signalling in craniofacial development and developmental disorders. *Oral Dis*. 2006;12:102-11.
75. Lu C, Huguley S, Cui C, Cabaniss LB, Waite PD, Sarver DM, Mamaeva OA, MacDougall M. Effects of FGFR Signaling on Cell Proliferation and Differentiation of Apert Dental Cells. *Cells Tissues Organs*. 2016;201:26-37.
76. Rice DP, Aberg T, Chan Y, Tang Z, Kettunen PJ, Pakarinen L, Maxson RE, Thesleff I. Integration of FGF and TWIST in calvarial bone and suture development. *Development*.

- 2000;127:1845-55.
77. Sgouros S. Skull vault growth in craniosynostosis. *Childs Nerv Syst.* 2005 ;21:861-70.
 78. Teven CM, Farina EM, Rivas J, Reid RR. Fibroblast growth factor (FGF) signaling in development and skeletal diseases. *Genes Dis.* 2014;1:199-213.
 79. Guimarães-Ferreira J, Miguéns J, Lauritzen C. Advances in craniosynostosis research and management. *Adv Tech Stand Neurosurg.* 2004;29:23-83.
 80. Mooney MP, Moursi AM, Opperman LA, Siegel MI. Cytokine therapy for craniosynostosis. *Expert Opin Biol Ther.* 2004;4:279-99.
 81. Wan DC, Kwan MD, Lorenz HP, Longaker MT. Current treatment of craniosynostosis and future therapeutic directions. *Front Oral Biol.* 2008;12:209-30.
 82. Johnson D. A comprehensive screen of genes implicated in craniosynostosis. *Ann R Coll Surg Engl.* 2003;85:371-7.
 83. Mooney MP, Moursi AM, Opperman LA, Siegel MI. Cytokine therapy for craniosynostosis. *Expert Opin Biol Ther.* 2004;4:279-99.
 84. Arnaud E, Molina F, Mendoza M, Fuente del Campo A, Ortiz-Monasterio F. Bone substitute with growth factor. Preliminary clinical cases for cranio- and maxillo-facial indications. *Ann Chir Plast Esthet.* 1998 ;43:40-50.
 85. Grova M, Lo DD, Montoro D, Hyun JS, Chung MT, Wan DC, Longaker MT. Models of cranial suture biology. *J Craniofac Surg.* 2012;23:1954-8.
 86. Atsawasuwan P, Lu X, Ito Y, Zhang Y, Evans CA, Luan X. Ameloblastin inhibits cranial suture closure by modulating MSX2 expression and proliferation. *PLoS One.* 2013 ;8:e52800.
 87. Slater BJ, Lenton KA, James A, Longaker MT. Ex vivo model of cranial suture morphogenesis and fate. *Cells Tissues Organs.* 2009;190:336-46.
 88. Warren SM, Longaker MT. The pathogenesis of craniosynostosis in the fetus. *Yonsei Med J.* 2001 ;42:646-59.
 89. Mehra BJ, Longaker MT. New developments in craniofacial surgery research. *Cleft Palate Craniofac J.* 1999 ;36:377-87.
 90. Kwan MD, Wan DC, Wang Z, Gupta DM, Slater BJ, Longaker MT. Microarray analysis of the role of regional dura mater in cranial suture fate. *Plast Reconstr Surg.* 2008 ;122:389-99.
 91. Stadler JA 3rd, Cortes W, Zhang LL, Hanger CC, Gosain AK. A reinvestigation of murine cranial suture biology: microcomputed tomography versus histologic technique. *Plast Reconstr Surg.* 2006;118:626-34.
 92. Byron CD, Maness H, Yu JC, Hamrick MW. Enlargement of the temporalis muscle and alterations in the lateral cranial vault. *Integr Comp Biol.* 2008 ;48:338-44.
 93. Chummun S, McLean NR, Flapper WJ, David DJ. The Management of Nonsyndromic, Isolated Sagittal Synostosis. *J Craniofac Surg.* 2016 ;27:299-304.
 94. Kung TA, Vercler CJ, Muraszko KM, Buchman SR. Endoscopic Strip Craniectomy for Craniosynostosis: Do We Really Understand the Indications, Outcomes, and Risks? *J Craniofac Surg.* 2016 ;27:293-8.
 95. Spruijt B, Rijken BF, den Ottelander BK, Joosten KF, Lequin MH, Loudon SE, van Veelen ML, Mathijssen IM. First Vault Expansion in Apert and Crouzon-Pfeiffer Syndromes: Front or Back? *Plast Reconstr Surg.* 2016;137:112e-121e.
 96. Maggi G, Aliberti F, Pittore L. Occipital remodeling for correction of scaphocephaly in the young infant. Technical note. *J Neurosurg Sci.* 1998 ;42:119-22.
 97. Scheuerle AE. Recent advances in craniofacial genetics. *J Craniofac Surg.* 1995;6:440-2.
 98. Muenke M. Finding genes involved in human developmental disorders. *Curr Opin Genet Dev.* 1995;5:354-61.
 99. Winter RM. Recent molecular advances in dysmorphology. *Hum Mol Genet.* 1995;4 :1699-704.
 100. Lewanda AF, Jabs EW. Genetics of craniofacial disorders. *Curr Opin Pediatr.* 1994 ;6:690-7.
 101. Eppley BL, Sadove AM. Surgical correction of metopic suture synostosis. *Clin Plast Surg.* 1994;21:555-62.
 102. Losken HW, Mooney MP, Hurwitz DJ, Siegel MI, Losken A, Zhang LP, Swan J. Frontal bone advancement and compensatory craniofacial growth changes in rabbits with experimental coronal suture immobilization. *J Craniofac Surg.* 1991;2:86-94.
 103. Mommaerts MY, De Vos W. Suture Autotransplantation and Dural Stripping for Craniosynostosis: A Long-Term Growth Study in Humans. *J Craniofac Surg.* 2015 ;26:2014-7.
 104. Tahiri Y, Swanson JW, Taylor JA. Distraction Osteogenesis Versus Conventional Fronto-Orbital Advancement for the Treatment of Unilateral Coronal Synostosis: A Comparison of Perioperative Morbidity and Short-Term Outcomes. *J Craniofac Surg.* 2015 ;26:1904-8.
 105. Pickrell BB, Lam SK, Monson LA. Isolated Unilateral Frontosphenoidal Craniosynostosis: A Rare Cause of Anterior Plagiocephaly. *J Craniofac Surg.* 2015 ;26:1944-6.
 106. Marsh JL, Galic M, Vannier MW. The craniofacial anatomy of Apert syndrome. *Clin Plast Surg.* 1991 ;18:237-49.
 107. Shen W, Cui J, Chen J, Weiping S. Molding of top skull in the treatment of Apert syndrome. *J Craniofac Surg.* 2015 ;26:516-7.
 108. LoPresti M, Daniels B, Buchanan EP, Monson L, Lam S. Virtual surgical planning and 3D printing in repeat calvarial vault reconstruction for craniosynostosis: technical note. *J Neurosurg Pediatr.* 2017 ;19:490-494.
 109. Ree JJ, Milovancev M, MacIntyre LA, Townsend KL. Factors associated with major complications in the short-term postoperative period in dogs undergoing surgery for brachycephalic airway syndrome. *Can Vet J.* 2016;57:976-80.
 110. Rinkinen J, Zhang P, Wang L, Enchakalody B, Terjimanian M, Holcomb S, Wang SC, Buchman SR, Levi B. Novel temporalis muscle and fat pad morphomic analyses aids preoperative risk evaluation and outcome assessment in nonsyndromic craniosynostosis. *J Craniofac Surg.* 2013;24:250-5.