



SPECTRAL ANALYSIS OF ETHYL 6-(CHLOROMETHYL)-4-(3-CHLOROPHENYL)-2-OXO-1,2,3,4-TETRAHYDROPYRIMIDINE-5-CARBOXYLATE– USING DFT COMPUTATIONS

G. Shakila*

Department of Physics, Bharathidasan Government College for Women, Puducherry-605003, India. *Corresponding author

H. Saleem

Department of Physics, Annamalai University, Annamalainagar-608002, Tamilnadu, India.

ABSTRACT In this work, the FT-IR, FT-Raman spectra, the NMR and UV-Vis spectra of ethyl 6-(chloromethyl)-4-(3-chlorophenyl)-2-oxo-1,2,3,4-tetrahydropyrimidine-5-carboxylate (ECM3CTC) have been recorded and analyzed. All the theoretical parameters in this work have been calculated using DFT at B3LYP/6-311++G(d,p) level. The molecular structures, fundamental vibrational frequencies and intensity of the vibrational bands were investigated theoretically and interpreted comparing with experimental values. The optimized bond parameters of ECM3CTC were compared with X-ray diffraction data of the molecule. The vibrational wavenumber were assigned on the basis of total energy distribution (TED) determined using Veda4 program. Stability of the molecule arising from hyper conjugative interactions, UV transition and charge delocalization was analyzed using natural bond orbital (NBO) analysis. In addition, the ¹H and ¹³C NMR chemical shifts values of ECM3CTC in the ground state were also calculated using Gauge independent atomic orbital (GIAO) method. The docking of the Molecule ECM3CTC was performed by using Glide module implemented in Maestro version 9.3.5 of Schrödinger software suite, 2011.

KEYWORDS : FT-IR, FT-Raman, NMR, UV analysis, NBO and Molecular docking studies.

1. INTRODUCTION

Pyrimidine nucleus contains two nitrogen atoms at positions 1 and 3 of the six-membered ring. Pyrimidine and its derivatives is the most important heterocyclic aromatic organic compound. It is exhibiting remarkable pharmacological and biological activities, because it is an essential constituent of all cells and thus all living matter [Sasada et al., 2009]. Pyrimidine ring is found in cytosine, uracil and thymine which are the essential building blocks of nucleic acids namely DNA and RNA [Amir et al., 2007]. It is also found in vitamin B2, riboflavin and folic acid [Stanisaw, 2009]. They are also found to exhibit remarkable pharmacological activities such as anti-cancer, anti-tumor, anti-microbial, anti-viral, analgesic, anti-HIV, anti-inflammatory and antifungal [Desai et al., 2006, Amr et al., 2007, Fujiwara et al., 2008 and Wagner et al., 2008] etc. They are also widely used as agrochemicals, pharmaceuticals, dyes, organic additives in electroplating of steel and in the polymerization process [Sharma et al., 2014 and Vaisalini et al., 2012]. Many pyrimidine derivatives have been developed as chemotherapeutic agents. Dihydro pyrimidinones, the product of the Biginelli reaction, are also used in the pharmaceutical industry [Beena et al., 2012]. Phenyl and its derivatives is biologically and industrially useful compound. They are used in the manufacturing of dyes, drugs, plastics, explosives, pesticides, etc. The inclusion of chlorine with phenyl ring leads to considerable variation of charge distribution in benzene ring and consequently greatly affects the structural, electronic and vibration properties. Generally chlorine or any halogen is referred to as electron withdrawing substituent in the aromatic systems [Bist et al., 1967 and Evans, 1960].

Chlorophenyl has got a variety of uses ranging from preparation of preservatives to insecticides. Thus, Owing to the wide applications and interesting properties of various constituent of the molecule ECM3CTC, a systematic spectral analysis supported by quantum computational methods based on DFT theories has been carried out in the present study. A detailed structural, vibrational, UV-Visible electronic transitional and NMR chemical shift studies have been reported. The non-covalent bonding and anti-bonding interaction can be quantitatively described in terms of the NBO analysis, which is expressed by means of the second-order perturbation interaction energy ($E^{(2)}$). This energy represents the estimation of the off-diagonal NBO Fock matrix elements. It can be deduced from the second-order perturbation approach.

2. COMPUTATIONAL DETAILS

The density functional theory (DFT) has been proven to be extremely effective in determining the various physical and chemical properties of the molecules. All the quantum chemical computations in the present work are performed using the Gaussian 09 software programs

[Frisch et al., 2004 and Schlegel, 1982] on a Pentium IV/3.02GHz personal computer. The Total energy distribution (TED) values are computed for every vibrational modes using VEDA4 program [Jamroz, 2004]. The Raman activities were transformed into Raman intensities using *Raint* program [Michalska, 2003 and Michalska et al., 2005]. The wave numbers and geometrical parameters were computed using B3LYP methods with 6-311++G (d,p) basis set. The geometry corresponding to the real minimum on the potential energy surface (PES) has been obtained and the UV-Visible spectra electronic transition such as HOMO-LUMO excitation energies and oscillator strength were calculated using time-dependent TD-SCF-B3LYP method and also using experimental method. The NMR chemical shift was carried out by GIAO method in B3LYP method with 6-311++G(d,p) basis set. The natural bonding orbital (NBO) computations were done using B3LYP function with 6-311++G(d,p) basis set based on a second-order perturbation theory.

3. EXPERIMENTAL DETAILS

3.1 Synthesis Procedure

The compound under study was prepared by heating a mixture of ethyl-4-chloroacetoacetate (4.1 ml, 0.025 mol), 3-chlorobenzaldehyde (3.6 g, 0.025 mol), and urea (4.5 g, 0.075 mol) in ethanol (5 ml) under reflux in the presence of concentrated HCl (1 mL) for 8 hrs (monitored by TLC). The reaction mixture, after being cooled to room temperature, was poured onto crushed ice and stirred for 5-10 min. The precipitate was washed with sodium bicarbonate solution, filtered, dried and again washed with petroleum ether (40-60%) and dried over in a vacuum. The compound was recrystallized from absolute ethanol with melting point 151-153° C and yield 72%. The schematic diagram of the title molecule is shown in Fig. 1a.

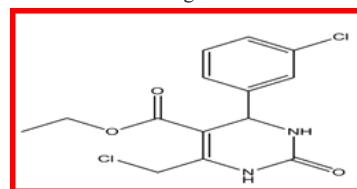


Fig.1a. Scheme of ECM3CTC

3.2. Spectral measurements

3.2.1. FT-IR, FT-Raman, UV-Vis and NMR Spectra Details

The FTIR spectrum was recorded in the spectral range 4000-400 cm^{-1} using KBr pellet technique with a FTIR-Shimadzu spectrometer. The spectrum was recorded at room temperature with a scanning speed of 10 cm^{-1} per minute with the spectral resolution of 2.0 cm^{-1} at Instrumentation laboratory, Jamal Mohamed College, Tiruchirappalli,

Tamilnadu. FT-Raman spectrum was recorded using Nd:YAG 1064 nm laser as excitation source in the region 4000-50 cm^{-1} on Bruker IFS 66v spectrophotometer equipped with a FRA 106 FT-Raman module accessory with spectral resolution of 4 cm^{-1} at SAIF Laboratory, IIT Madras. The UV-Vis absorption spectrum of ECM3CTC was recorded in the range of 200-500 nm using a Shimadzu-2600 spectrometer in the department of Chemistry, Jamal Mohamed College, Tiruchirappalli, Tamilnadu. ^1H NMR spectra were recorded on a Bruker 400 MHz spectrometer and ^{13}C NMR spectra were recorded on a Bruker 100 MHz spectrometer in the department of chemistry, Annamalai University, Annamalainagar. Chemical shift values are reported in parts per million (ppm) and tetramethylsilane (TMS) as internal standard.

4. RESULTS AND DISCUSSION

4.1 Geometrical Analysis

The structure of the ECM3CTC molecule was optimized at B3LYP/6-311++G(d,p) level. Fig. 1b shows the optimized structure of ECM3CTC molecule showing the atom symbols and numbering scheme, respectively. The single crystal X-ray structure for the novel ethyl 6-(chloromethyl)-4-(3-chlorophenyl)-2-oxo-1,2,3,4-tetrahydro pyrimidine-5-carboxylate molecule reported at the earlier work [Bharanidharan et al., 2014] was used for comparison and analysis. The selected bond parameters such as bond lengths, bond angles and dihedral angles values are presented in Table.1 against the reported XRD data. In this study, the tetrahydropyrimidine ring is found to be almost planar; their bond lengths and bond angles nearly coincide with literature value [Bharanidharan et al., 2014]. From the theoretical values, it is found that most of the optimized bond lengths are slightly larger than the experimental values. The CC bond length values inside the pyrimidine ring lies in the range 1.403 Å to 1.374 Å, the corresponding XRD values are 1.357 Å and 1.394 Å, respectively. But the phenyl ring CC values lies in the range 1.38 Å to 1.39 Å. This difference in values indicate the conjugational difference between the pyrimidine and phenyl ring, which shows the electronic distribution inside these two rings are slightly different and there is neither single bonded nor double bonded CC inside these rings. The single bonded C-C has the bond length values lies in the range 1.45 to 1.50 Å, while the double bonded CC values in the range 1.33 to 1.34 Å. The table shows C10-C11 bond length is 1.529 Å and that of C11-C24 is 1.487 Å, which indicates they are purely single bonded but with a different electronic density or distribution. According to the bond length values computed for this molecule, there is no CC bond which is purely double bonded but some of the bonds in the phenyl and pyrimidine rings are closer to double bonded CC, which can be identified through vibrational analysis. The average bond distances of C12-N18 and C14-N18 in the pyrimidine ring calculated by DFT method are 1.403 Å and 1.374 Å, respectively. The breakdown of hexagonal symmetry of the pyrimidine ring is obvious from the shortening of C12-N18 (~0.046 Å) and elongation of C14-N18 (~0.019 Å) due to the substitution of oxo group to the ring. There is a shortening of C20-CL23 bond length (~0.056 Å) and C24-O26 bond length (~0.029 Å) due to the substitution of chloro methyl group and carboxylate group to the pyrimidine ring.

The asymmetry of the pyrimidine ring is also evident from the negative deviation of C10-C11-C14 (118°) and positive deviation of N16-C12-O17 (125°) from the normal value of 120°. The carbon atoms usually take two types of hybridization SP^2 and SP^3 when they are in rings, with bond angle 120° and 109° respectively. In this molecule, for many of the bond angles around carbon show the values in the range 118° to 122° both in phenyl and pyrimidine rings which shows the deviation from the SP^2 hybridization at these atoms. At H21-C20-H22, H21-C20-CL23 and H22-C20-CL23 the bond angles are 108.4°, 107.4°, 106.6° respectively, this shows the deviation from the SP^3 hybridization. It also indicates that the SP^3 takes place when there is an H atom on one side of the carbon atom. All these deviation from the ideal angles 120° and 109° indicate that the regular hexagonal structure is distorted due to the substitutional groups and presence of N atoms in the pyrimidine group.

Also, the N18-C12-N16-C10, N16-C12-N18-C14 and O17-C12-N18-C14 dihedral angle is positively (~2.2°), (~3.4°) and (~4.4°) deviated from the corresponding XRD values. The O17-C12-N16-C10 dihedral angle is negatively (~4.1°) deviated from the corresponding XRD value. This deviation is mainly due to the substitution of oxo group and chloro methyl group to the ring. The C10-C1-C2-C3 dihedral angle is positively (~1.1°) deviates from the corresponding XRD value is due to the substitution of chloro phenyl group to the ring. The carboxylate

group C11-C24-O26-C27 dihedral angle parameter is positively (~2.5°) deviated with reported XRD value, due to the reason for the crystal disorder over two positions in ethyl group.

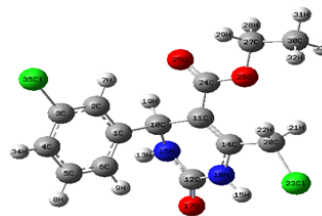


Fig. 1b. Optimized structure of ECM3CTC

Table 1. The selected bond parameters of ECM3CTC

Bond Parameters	B3LYP/6-311++G(d,p)	XRD ^a
Bond Lengths (Å)		
C1-C2	1.4026	1.351
C1-C6	1.3997	1.381
C1-C10	1.5387	1.526
C2-C3	1.3917	1.354
C3-C4	1.3943	1.381
C3-CL35	1.7617	1.723
C4-C5	1.3933	1.364
C5-C6	1.396	1.358
C10-C11	1.529	1.503
C10-N16	1.4622	1.482
C11-C24	1.4666	1.487
C12-N16	1.3663	1.333
C12-O17	1.2203	1.228
C12-N18	1.4036	1.357
C14-N18	1.3744	1.394
C20-CL23	1.8167	1.76
C24-O25	1.2221	1.205
O26-C27	1.4496	1.46
Bond Angles (°)		
C2-C1-C6	119.1487	122.3
C1-C2-C3	119.5599	117.8
C2-C3-C4	121.7374	121.6
C3-C4-C5	118.3725	119.6
C4-C5-C6	120.8042	119.9
C1-C6-C5	120.3738	118.8
C1-C10-N16	113.2246	110.7
C11-C10-N16	108.5973	108.6
N16-C12-O17	125.3629	123
O17-C12-N18	121.0855	121.3
C10-N16-C12	123.148	123.8
O25-C24-O26	121.9093	122.4
Dihedral Angles (°)		
C6-C1-C2-C3	0.6511	1.0
C10-C1-C2-C3	-179.0294	-177.9
C10-C1-C6-C5	179.0242	179.0
O17-C12-N16-C10	160.2809	164.3
N18-C12-N16-C10	-21.6273	-19.4
N16-C12-N18-C14	-9.4486	-6.0
O17-C12-N18-C14	168.7343	164.3
C11-C24-O26-C27	179.8018	177.3

^aBharanidharan et al., J.New developments in chem., 1:2 (2016) 1-25.

4.2 Vibrational Investigations

The title molecule under investigation has 35 number of atoms and 99 normal modes of fundamental vibrations, which can be distributed using the irreducible representations as $\Gamma_{\text{vib}} = 67 A' + 32 A''$. All the vibrations are found active collectively in both IR and Raman spectra. The assignments of all the fundamentals have been made on the basis of TED values along with the comparison of available literature on the structurally similar molecules. The calculated wave numbers are found slightly higher than the observed values for the majority of the normal

modes. Two factors may be responsible for the discrepancies between the experimental and computed wave numbers; the first is caused by the unpredictable electronic distribution among the different bonds in the molecule and the second reason is the anharmonic nature of the vibrations [Scott et al., 1996] which cannot be accounted completely by theory. Scaling strategies were used to bring computed wave numbers coincide with observed values. In this, study, the scaling factor used is 0.9608 as advised by the earlier work [Radom et al., 1970]. The intensities, reduced mass and Force constants along with the wave numbers and assignment are presented in Table 2. The theoretical and experimental IR and Raman spectra are also constructed and shown in Figs. 2 & 3.

4.2.1 CH vibrations

In the aromatic compounds, the C-H stretching normally occurs in the region of 3100–3000 cm^{-1} [Michalska, 2003]. For the title molecule, the C-H stretching vibrations of chloro phenyl group are observed at 3228, 3119 cm^{-1} in IR region and 3075 cm^{-1} in Raman (mode: 3, 4, 7). The corresponding calculated values occur at 3228, 3223 and 3193 cm^{-1} . This observation indicates that all the stretching vibrations are in the higher range and this is due to the considerable impact of substitution on the rings. Especially the bands are predominated by the Cl atom in the phenyl ring. The scaled harmonic frequencies in the range 3139-3056 cm^{-1} (mode nos: 8,9,11, 13, 14) are attributed to C-H stretching vibrations of carboxylate group attached to the pyrimidine ring in ECM3CTC. All these mixing modes are observed in the FT-Raman spectrum and all these vibrations are in the lower range due to the impact of O atom in the carboxylate group. The pure mode of wave number 2972 cm^{-1} in the FT-Raman spectrum (mode 12) with TED 99% contribution is due to the C-H stretching vibration of the pyrimidine ring. This mode is shifted to the lower wave number region due to the substitution of chloro phenyl group in the adjacent position. The corresponding computed unscaled value is 3097 cm^{-1} and this coincides with the assignment made by Alam et al. [Alam et al., 2016]. The theoretically calculated values of C-H in-plane bending vibrations of phenyl ring fall in the region 1519-1198 cm^{-1} (mode nos: 21, 32, 39, 41). Similarly the harmonic bands due to C-H out-of-plane bending vibration fall in the region 985-790 cm^{-1} (mode nos: 51, 54, 59, 61). In

this study, some of the experimental bands are slightly higher for β_{CH} mode, which may be due to the steric factor of bulky group (pyrimidine ring). In general, the β_{CH} and τ_{CH} modes are within the expected region with few exceptions and also having considerable TED values: >20 and mode no: 54 is a pure out-of-plane bending mode with TED 79%, respectively.

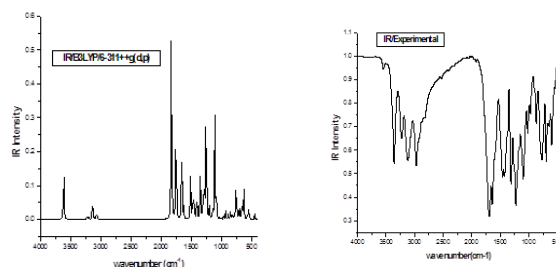


Fig. 2. The combined theoretical and experimental FT-IR Spectra of ECM3CTC

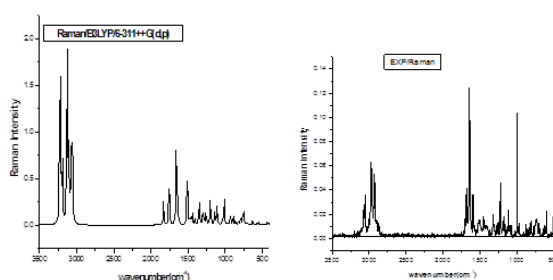


Fig. 3. The combined theoretical and experimental FT-Raman Spectra of ECM3CTC

Table 2. The experimental and calculated frequencies of ECM3CTC using B3LYP/6-311++G(d,p) level of basis set [harmonic frequencies (cm^{-1}), IR, Raman intensities (Km/mol), reduced masses (amu) and force constants (mdynA^{-1})]

Mode No.	Calculated Frequencies (cm^{-1})		Observed Frequencies (cm^{-1})		IR Intensity		Raman Intensity		Reduced Masses	Force Constants	Vibrations Assignments $\geq 10\%$ (TED) ^d
	Un Scaled	Scaled ^a	IR	Raman	Rel. ^b	Abs.	Rel. ^c	Abs.			
1	3631	3511	3544		38.75	6.93	31.00	2.24	1.08	8.37	ν_{N16H13} (100)
2	3613	3493	3360		153.45	27.46	27.56	1.99	1.08	8.29	ν_{N18H15} (100)
3	3228	3121	3228		5.83	1.04	21.90	1.58	1.09	6.70	ν_{C2H7} (99)
4	3223	3116	3119		4.76	0.85	72.55	5.23	1.10	6.71	ν_{C4H34} (79)+ ν_{C5H8} (11)
5	3216	3109			3.30	0.59	15.63	1.13	1.09	6.65	ν_{C4H34} (14)+ ν_{C6H9} (81)
6	3212	3106			0.14	0.03	13.85	1.00	1.10	6.70	ν_{C20H21} (15)+ ν_{C20H22} (84)
7	3193	3087		3075	6.89	1.23	36.11	2.60	1.09	6.53	ν_{C5H8} (84)+ ν_{C6H9} (10)
8	3139	3034		3056	35.63	6.38	5.53	0.40	1.11	6.42	ν_{C27H28} (13)+ ν_{C27H29} (13)+ ν_{C30H32} (36)+ ν_{C30H33} (37)
9	3132	3028			21.81	3.90	50.82	3.67	1.10	6.37	ν_{C30H31} (67)+ ν_{C30H32} (16)+ ν_{C30H33} (15)
10	3128	3024			4.28	0.77	48.86	3.52	1.07	6.14	ν_{C20H21} (84)+ ν_{C20H22} (15)
11	3112	3009			1.97	0.35	40.87	2.95	1.11	6.32	ν_{C27H28} (37)+ ν_{C27H29} (36)+ ν_{C30H32} (14)+ ν_{C30H33} (13)
12	3097	2994		2972	3.81	0.68	28.85	2.08	1.09	6.14	ν_{C10H19} (99)
13	3073	2971	2973	2928	15.79	2.83	41.94	3.02	1.06	5.89	ν_{C27H28} (48)+ ν_{C27H29} (50)
14	3056	2954		2872	13.86	2.48	68.83	4.96	1.04	5.70	ν_{C30H31} (32)+ ν_{C30H32} (34)+ ν_{C30H33} (34)
15	1831	1770	1924		558.88	100.00	46.47	3.35	7.96	15.73	ν_{O17C12} (72)+ $\beta_{\text{N16C12N18}}$ (11)
16	1756	1698	1692	1683	347.18	62.12	125.07	9.02	12.17	22.11	ν_{O25C24} (86)

17	1660	1605	1640	1645	163.77	29.30	158.91	11.46	8.54	13.87	VC11C14(66)
18	1648	1593		1597	25.73	4.60	86.57	6.24	5.93	9.49	VC4C3(17)+VC6C5(28)+βC5C4C3(12)
19	1628	1574			39.21	7.02	21.22	1.53	5.70	8.90	VC5C4(52)+VC1C6(54)+βC4C3C2(35)
20	1532	1481			2.86	0.51	2.77	0.20	1.09	1.51	βH28C27H29(54)+βH32C30H33(16)+ΓC27H28C30H29(17)
21	1519	1469			40.56	7.26	3.81	0.27	2.24	3.05	βH7C2C3(20)+βH8C5C4(31)+βH9C6C5(10)
22	1511	1461		1508	1.77	0.32	48.34	3.49	1.05	1.41	βH28C27H29(32)+βH32C30H33(42)
23	1509	1459	1458	1456	113.23	20.26	58.01	4.18	2.39	3.21	vN18C14(26)+βH15N18C14(44)
24	1502	1452			6.79	1.22	55.57	4.01	1.04	1.38	βH31C30H32(65)+τH32C30C27O26(14)+τH33C30C27O26(10)
25	1474	1425	1424		66.09	11.83	20.91	1.51	2.19	2.81	βH13N16C10(10)+βH9C6C5(13)+ΓC10C11H19(21)
26	1464	1415			32.30	5.78	11.68	0.84	1.15	1.45	βH21C20H22(82)
27	1453	1405			44.74	8.00	10.15	0.73	1.91	2.37	βH13N16C10(35)
28	1439	1391			11.75	2.10	32.37	2.33	1.41	1.72	βH31C30C27(23)+βH32C30H33(13)+ΓC27H28C30H29(21)+TH33C30C27O26(10)
29	1408	1361			6.28	1.12	5.20	0.38	1.55	1.81	βH31C30C27(17)+βH32C30H33(15)+ΓC27H28C30H29(29)
30	1406	1360	1374		62.53	11.19	16.71	1.21	1.97	2.29	vN16C12(12)+vN18C12(11)+βH13N16C10(23)+ΓC10C11H19(14)
31	1358	1313	1310	1323	48.67	8.71	16.90	1.22	2.36	2.56	VC4C3(21)+VC1C6(13)+βH19C10C1(39)
32	1351	1306			16.45	2.94	9.63	0.69	1.50	1.61	βH7C2C3(26)+βH34C4C5(18)+βH9C6C5(10)+βH19C10C1(20)
33	1347	1302			126.39	22.61	76.47	5.52	3.56	3.80	VC24C11(19)+ΓC27H28C30H29(15)
34	1308	1265		1275	52.32	9.36	17.41	1.26	1.35	1.36	ΓC20H22C14H21(53)
35	1301	1258			26.84	4.80	4.97	0.36	1.77	1.77	VC4C3(13)+VC1C6(19)
36	1297	1254		1256	1.77	0.32	42.74	3.08	1.10	1.09	βH28C27C30(77)
37	1285	1243			130.45	23.34	32.79	2.37	2.28	2.22	vN18C14(20)+βH15N18C14(31)
38	1257	1215	1226	1225	392.99	70.32	62.50	4.51	2.23	2.08	ΓC10C11H19(25)
39	1206	1166		1193	15.28	2.73	15.28	1.10	1.34	1.15	βH8C5C4(21)+βH9C6C5(26)
40	1202	1162		1172	6.34	1.13	39.90	2.88	1.09	0.93	βH22C20C14(70)+ΓC20H22C14H21(11)
41	1198	1158			27.67	4.95	62.16	4.48	2.10	1.77	VC6C5(15)+βC2C1C6(13)+VC1C10(20)+βH34C4C5(11)+βH8C5C4(13)
42	1183	1144			3.52	0.63	7.84	0.57	1.53	1.26	βH31C30H32(14)+ΓC27C30O26H28(18)+τH28C27C30H31(36)+TH32C30C27O26(22)
43	1149	1111		1119	23.48	4.20	15.13	1.09	2.40	1.86	vO ₂₆ C ₂₄ (10)+vN ₁₆ C ₁₀ (18)+vC ₁₀ C ₁₁ (16)
44	1140	1102	1099		4.43	0.79	48.04	3.46	2.22	1.70	vC ₂₇ C ₃₀ (18)+βH ₃₁ C ₃₀ H ₂₇ (16)+βC ₃₀ C ₂₇ O ₂₆ (15)+τH ₃₂ C ₃₀ C ₂₇ O ₂₆ (14)+τH ₃₃ C ₃₀ C ₂₇ O ₂₆ (14)
45	1124	1087			9.08	1.62	5.80	0.42	1.68	1.25	βH34C4C5(27)+βC4C3C2(10)
46	1113	1076		1080	59.51	10.65	36.28	2.62	2.24	1.64	VC5C4(19)+VC6C5(10)+βC4C3C2(15)
47	1112	1075			280.51	50.19	56.83	4.10	5.71	4.16	VO26C24(20)+VC20C14(12)+VO26C27(12)
48	1087	1051			65.49	11.72	12.92	0.93	3.82	2.66	vN16C10(13)+VC11C10(15)
49	1037	1002	1018	999	6.69	1.20	16.09	1.16	2.93	1.85	VC30C27(45)+VO26C27(18)
50	1012	979	972	973	2.28	0.41	164.77	11.88	6.15	3.71	βC1C2C3(19)+VC6C5(10)+βC6C5C4(29)

51	985	952			0.01	0.00	2.06	0.15	1.30	0.74	τ H34C4C5C6(13)+ Γ C5C4C6H8(62)+ τ H9C6C5C4(20)
52	965	933			18.24	3.26	13.93	1.00	3.71	2.04	ν N16C12(16)+ ν 18C12(29)+ ν C20C14(14)
53	933	902			6.59	1.18	33.13	2.39	1.77	0.91	Γ C20C14C123H 22(74)+ Γ C20C11N18C14(10)
54	928	897			21.14	3.78	28.42	2.05	1.45	0.74	Γ C2C1C3H7(79)
55	919	888		877	1.68	0.30	4.25	0.31	1.36	0.68	τ H34C4C5C6(43)+ τ H9C6C5C4(44)
56	893	863	867		20.34	3.64	61.63	4.44	2.82	1.33	ν C30C27(10)+ ν O26C27(27)+ β H31C30C27(14)
57	869	840		841	3.93	0.70	22.78	1.64	4.36	1.94	ν O26C24(10)+ ν C1C10(12)+ β C5C4C3(12)
58	853	824		810	24.88	4.45	21.41	1.54	6.51	2.79	β O25C24O26(12)+ β N16C12N18(10)
59	823	796			18.15	3.25	22.20	1.60	2.80	1.12	τ H34C4C5C6(11)+ Γ C1C11N16C10(18)
60	813	786			0.52	0.09	3.47	0.25	1.12	0.44	β H28C27C30(12)+ Γ C27C30O26H28(66)
61	790	763	769	770	13.50	2.42	42.27	3.05	1.94	0.71	τ H34C4C5C6(15)+ Γ C5C4C6H8(19)+ τ H9C6C5C4(14)
62	766	740			17.90	3.20	32.86	2.37	7.26	2.51	Γ O25C11O26C24(50)+ Γ O17N16N18C12(10)
63	761	735			72.56	12.98	44.07	3.18	7.82	2.67	ν C123C20(21)+ Γ O25C11O26C24(10)+ Γ O17N16N18C12(30)
64	747	722		739	37.70	6.75	119.99	8.65	7.48	2.46	ν C123C20(16)+ Γ O17N16N18C12(48)
65	715	691	696		16.44	2.94	19.02	1.37	3.20	0.96	τ C1C6C4C5(15)+ Γ O25C11O26C24(11)
66	703	679		703	35.07	6.27	18.38	1.33	4.41	1.28	β C6C5C4(12)+ τ C3C2C4C5(10)+ τ C1C6C4C5(14)
67	675	653	663		8.75	1.57	12.81	0.92	3.79	1.02	β N16C12N18(15)+ τ H15N18C14C11(13)+ τ C3C2C4C5(10)
68	656	634	642	633	58.35	10.44	10.55	0.76	2.68	0.68	β O25C24O26(10)+ β N16C12N18(10)+ τ H13N16C12N18(10)+ τ H15N18C14C11(22)
69	630	610			83.91	15.01	28.71	2.07	2.00	0.47	τ H13N16C12N18(12)+ τ H15N18C14C11(32)
70	624	604	600	602	10.05	1.80	38.09	2.75	5.31	1.22	ν C123C20(13)+ β C1C10N16(11)+ τ C3C2C4C5(12)
71	565	546	542		18.90	3.38	27.12	1.96	1.51	0.28	τ H13N16C12N18(47)+ τ H15N18C14C11(19)
72	549	531			15.57	2.79	16.89	1.22	3.96	0.70	β C11C14N18(14)+ Γ C20C11N18C14(13)
73	548	530			10.87	1.95	21.35	1.54	3.84	0.68	β C11C14N18(14)+ β C11C10N16(11)+ Γ C20C11N18C14(12)
74	525	508		512	6.92	1.24	18.33	1.32	4.06	0.66	β N18C12O17(13)+ β N16C12N18(14)
75	453	438	456		10.21	1.83	1.08	0.08	3.51	0.42	τ C4C3C5C6(36)+ τ C1C6C4C5(17)
76	447	432		424	9.11	1.63	6.72	0.48	6.02	0.71	β N18C12O17(27)+ β O26C24C11(18)+ β C30C27O26(12)
77	427	413			2.52	0.45	50.83	3.67	7.84	0.84	ν C135C3(41)+ β C5C4C3(10)
78	392	379			7.12	1.27	43.07	3.11	4.62	0.42	β O25C24O26(22)+ β C30C27O26(31)
79	378	365		364	0.32	0.06	111.86	8.07	5.47	0.46	β C4C3C135(14)
80	356	345			2.24	0.40	36.24	2.61	6.48	0.49	β C2C1C6(10)+ β C11C10N16(10)+ β C4C3C135(22)
81	321	310			17.97	3.22	49.18	3.55	7.72	0.47	β O26C24C11(10)+ β C27O26C24(18)+ β C123C20C14(12)
82	302	292			0.63	0.11	55.07	3.97	5.67	0.31	ν C1C10(11)+ β C6C1C10(11)+ β C1C10N16(11)+ β C4C3C135(16)
83	288	278			1.38	0.25	58.94	4.25	6.57	0.32	ν C24C11(12)+ β C1C10N16(14)+ β C27O26C24(11)+ β C123C20C14(10)
84	270	261			1.05	0.19	9.90	0.71	1.28	0.06	τ H28C27C30H31(40)+ τ H33C30C27O26(33)
85	227	220			3.67	0.66	55.93	4.03	6.06	0.18	β C10C11C24(29)+ β C30C27O26(11)
86	216	209		201	1.69	0.30	113.83	8.21	4.59	0.13	τ C3C2C4C5(14)+ τ C4C3C5C6(12)+ τ C1C6C4C5(13)+ Γ C24C10C14C11(11)
87	193	187			2.82	0.51	25.94	1.87	5.47	0.12	τ C1C2C3C135(60)
88	172	167		166	1.71	0.31	44.89	3.24	9.68	0.17	β C11C14C20(40)+ β C123C20C14(26)
89	160	155			2.51	0.45	151.81	10.95	5.77	0.09	β C6C1C10(16)+ β C4C3C135(10)+ τ C12N18C11C14(18)
90	154	149			1.22	0.22	93.90	6.77	4.72	0.07	β C6C1C10(17)+ β C4C3C35(11)+ τ C12N18C11C14(39)+ Γ C20C11N18C14(10)
91	128	123			1.73	0.31	3.49	0.25	3.16	0.03	τ C27O26C24C11(57)
92	105	101			0.69	0.12	18.93	1.37	6.13	0.04	τ C14C11C24O26(44)+ τ C30C27O26C24(12)
93	94	91		90	0.18	0.03	25.15	1.81	5.11	0.03	β O26C24C11(23)+ β C10C11C24(18)+ β C27O26C24(19)
94	75	72			0.56	0.10	134.42	9.69	5.78	0.02	τ C10N16N18C12(57)+ Γ C24C10C14C11(10)
95	65	63			3.02	0.54	394.63	28.46	3.94	0.01	τ C10N16C11C14(30)+ τ N18C14C20C123(13)+ Γ C1C11N16C10(11)
96	57	55			0.29	0.05	250.21	18.04	3.17	0.01	τ C30C27O26C24(47)
97	32	31			0.03	0.01	776.69	56.01	4.49	0.00	Γ C14C11C24O26(10)+ τ C30C27O26C24(24)+ τ N18C14C20C123(16)
98	19	19			0.53	0.09	1095.91	79.03	6.20	0.00	τ C6C1C10C11(56)+ τ C14C11C24O26(11)+ τ N18C14C20C123(10)
99	9	9			0.96	0.17	1386.68	100.00	10.30	0.00	τ C6C1C10C11(17)+ τ C10N16C11C14(20)+ τ N18C14C20C123(38)

ν: Stretching, β: in-plane-bending, Γ: out-of-plane bending, τ- Torsion, vw: very weak, w: week, m: medium, s: strong, vs: very strong.
 *Scaling factor: 0.9608 (Random et al., 1970 and Pople et al., 1993).

^bRelative IR absorption intensities normalized with highest peak absorption equal to 100.

^c Relative Raman intensities calculated by Equation (1) and normalized to 100.

^d Total energy distribution calculated at B3LYP/6-311++G(d,p) level.

4.2.2. C-Cl Vibrations

Most aromatic chlorine compounds absorb strongly in the region 600-800 cm^{-1} due to C-Cl stretching vibrations, when there is more than one chlorine atoms on the same ring. The vibrations due to a chlorine atom is worth to discuss since mixing of the vibrations are possible due to lowering of the molecular symmetry and the presence of heavy atom [Mooney, 1964, Mooney, 1963]. [Sundaraganesan et al., 2008] reported C-Cl stretching at 704 (FT-IR), 705 (Raman), and 715 cm^{-1} (DFT) and the deformation bands at 250 and 160 cm^{-1} . In present case, the band observed at 739 cm^{-1} in FT-Raman spectrum is assigned as C-Cl stretching of chloro methyl group and the corresponding theoretical wavenumber at 747 cm^{-1} . Sundaraganesan et al. reported the C-Cl in-plane and out-of-plane bending deformation bands at 250 and 160 cm^{-1} , respectively [Sundaraganesan et al., 2008]. In this study, the C-Cl in-plane bending vibrations of chloro phenyl ring were calculated at 378, 302 cm^{-1} and that of chloro methyl group at 288, 172 cm^{-1} . Similarly, the C-Cl out-of-plane bending vibrations of chloro phenyl ring and that of chloro methyl group were calculated at 193 & 65 and 32 & 19 cm^{-1} respectively.

4.2.3. C-N Vibrations

Generally the C=N/C-N stretching vibrations were observed in the regions of 1600-1670 cm^{-1} /1266-1382 cm^{-1} [Socrates, 2001, Silverstein et al., 1998] respectively. Khalaji et al. reported that the C=N (aromatic) stretching mode appeared in the region 1490-1570 cm^{-1} [Khalaji et al., 2009]. For the title molecules, the four C-N stretching vibrations are identified with the help of TED. The CN stretching vibrations are assigned to harmonic wavenumbers 1509, 1406, 1285, 1149 cm^{-1} (mode nos: 23, 30, 37, 43). The observed bands occurred in FTIR spectra at 1458 (1456: FT-Raman), 1374, 1119 cm^{-1} respectively. These assignments are supported by their TED values. The assignment of CN stretching modes in this molecule indicate that the CN is single bonded rather than double bonded, as the wave number lies in the lower range. The in-plane bending vibrations: $\beta_{C_1C_{10}N_{16}}$, $\beta_{C_{11}C_{14}N_{18}}$; $\beta_{N_{16}C_{12}N_{18}}$ are attributed to mode nos: 70, 72, 73, 74, 67 and 68 respectively. The out-of-plane bending mode of Γ_{C-N} contributes as mixed vibrations of $\Gamma_{O_{17}N_{16}N_{18}C_{12}}$ and $\Gamma_{C_{20}C_{11}N_{18}C_{14}}$ (mode nos: 63, 90).

4.2.4. C-O Vibrations

The carboxylate C=O stretching mode of ester group [Roeges, 1994 and Colthup et al., 1975] is expected to occur in the region 1750-1600 cm^{-1} and for ECM3CTC, this mode is observed with very strong band at 1692 cm^{-1} in FT-IR spectrum and 1683 cm^{-1} in FT-Raman. The theoretically calculated wavenumber at 1756 cm^{-1} (mode no. 16) corresponds to C=O stretching vibration mode which is pure mode with TED contribution of 86%. In C-O group, the absorption is sensitive for both the carbon and oxygen atoms. Normally the C-O stretching vibration occurs in the region 1000-1260 cm^{-1} . [Meganathan et al. 2010] have assigned at 1251 cm^{-1} (FTIR) and 1248 cm^{-1} (FT-Raman) to above mentioned band. The intensity of the carboxylate group increases, due to the conjugation (or) formation of hydrogen bonds. The increase in conjugation, which increase the intensity of Raman lines as well as the IR band intensities [Varghese et al., 2009]. According to the above facts, the harmonic frequencies 1149, 1112, 1037 (mode nos: 43, 47, 49) are designated as $\nu_{C_{24}O_{26}}$, $\nu_{C_{27}O_{26}}$ modes, respectively with small TED values (10, 20, 18%) as well as relative IR intensities (23.48, 280.51, 6.69). Further the mode no: 43 appeared as FT-Raman band: 1119 cm^{-1} with medium intensity and mode no: 49 appeared in both FT-IR and FT-Raman region. The in-plane bending vibrations of β_{C-O} are observed as mixed vibration of $\beta_{C_{27}C_{30}O_{26}}$ at 424 cm^{-1} in FT-Raman spectrum, whereas the calculated frequencies are: 447, 392 (mode nos: 76, 78). The C-O out-of-plane bending modes are also contribute as mixed vibration of $\Gamma_{C_{14}C_{11}C_{20}O_{26}}$ and $\tau_{C_{14}C_{11}C_{20}O_{26}}$, but shifted to low wave number value due to the substitution of chloro phenyl group in the adjacent position.

4.2.5. CC Vibrations

The ring stretching vibrations are very much important and are characteristic of the phenyl and pyrimidine rings. The C=C stretching vibrations were reported in the regions of 1590-1625, and C-C in the region 1280-1380 cm^{-1} with variable intensity by [Varsanyi, 1973]. In this study, the bands observed at 1597, 1323, 1080 cm^{-1} in FT-Raman

and at 1310 cm^{-1} in FT-IR spectra are attributed to C=C/C-C stretching mode of phenyl ring in ECM3CTC and their corresponding harmonic frequencies are: 1648, 1628, 1358, 1301, 1198, 1113 cm^{-1} (mode nos: 18, 19, 31, 35, 41, 46). As it is evident from Table the TED value corresponding to all ν_{CC} modes lie in between 12 and 54% and also mixed with β_{CH} mode. The wavenumbers for in-plane bending modes of phenyl group are determined experimentally at 1080, 973, 841, 703 cm^{-1} in FT-Raman spectrum and 972 cm^{-1} in FT-IR spectrum. The theoretically computed wave numbers by DFT method are at 1113, 1012, 869, 703 cm^{-1} [Sarojini et al., 2013]. The modes: 65, 67, 70, 75 at 696, 663, 600, 456 cm^{-1} (FTIR) respectively were recognized as torsional deformation τ_{CC} modes.

[Alam et al., 2016] assigned the C-C stretching vibrations of pyrimidine derivatives in the wavenumber range 1180-1771 cm^{-1} (FTIR). The theoretically calculated ν_{CC} modes: 1660-1087 cm^{-1} (mode nos: 17, 33, 41, 43, 47, 48, 52) of the pyrimidine ring have been obtained to be consistent with the recorded spectral values: 1640, 1428, 1365, 1212 cm^{-1} (FTIR bands) and 1645, 1119 cm^{-1} (FT-Raman). The harmonic wave numbers of β_{CCC} and Γ_{CCC} modes of pyrimidine ring were computed at 302, 227, 172 and 216, 75 cm^{-1} respectively. Compared to the above literature values [Alam et al., 2016], these harmonic wave numbers are shifted to the lower range due to the presence of chloro phenyl, carboxylate group and chloro methyl group to the tetra hydro perimidine ring. According to TED results the β_{CCC} and Γ_{CCC} vibrations are mixed with β_{CH} , β_{CCN} and τ_{CH} , τ_{CCN} modes of the neighbouring group.

4.2.6. N-H Vibrations

The N-H stretching vibrations usually appear in the region of 3500-3300 cm^{-1} for heterocyclic molecules [Socrates, 2001]. In the present molecule the observed vibrational wave numbers at 3544, 3360 cm^{-1} in FTIR spectra and its corresponding calculated values are at 3631, 3613 cm^{-1} (mode nos: 1 & 2) are assigned to ν_{NH} mode. These modes are in expected region and also find support from TED value (100%). The observed wave numbers are negatively (87 and 253 cm^{-1}) deviated from harmonic frequencies, which is due to the presence of moderate anharmonicity in this vibration. It may also be due to hydrogen-bonding interactions occur within the molecule [Alam et al., 2016]. The N-H in-plane bending β_{NH} vibration is an important mode, which shows their presence as medium to strong bands in the region 1520-1549 cm^{-1} [Socrates, 2001]. In the present molecule, when nitrogen and hydrogen move in opposite with respect to carbon atom the peak observed is at 1458, 1424, 1374 cm^{-1} in FT-IR and 1456 cm^{-1} in FT-Raman band, while the corresponding harmonic wave number at 1509, 1474 and 1406 cm^{-1} (mode nos: 23, 25, 30) are assigned to β_{NH} vibration. Similarly, when the N and H atoms move in same direction it is observed near 1250 cm^{-1} [Dhandapani et al., 2014]. In ECM3CTC, the N-H in-plane bending mode $\beta_{H_{15}N_{18}C_{14}}$ is calculated at 1285 cm^{-1} and the corresponding experimental band is absent. The N-H torsional mode $\tau_{H_{15}N_{18}C_{14}C_{11}}$ is assigned at 663, 642 cm^{-1} in FT-IR spectrum, 634 cm^{-1} in FT-Raman spectrum and for the same mode the corresponding harmonic values are 675, 656 cm^{-1} (mode nos: 67, 68).

4.2.7. Methyl group Vibrations

The compound ECM3CTC contains one CH_2Cl group attached to pyrimidine ring. In CH_2Cl , the C-H stretching occurs at lesser frequencies than those of aromatic C-H stretching (3100-3000 cm^{-1}). The region around 2980 cm^{-1} is assigned to asymmetric C-H stretching mode of CH_2Cl group [Coluthup et al., 1990, Kleiman, 1977, Smith, 1999 and Socrates 1990] and the region around 2870 cm^{-1} is allotted to symmetric C-H stretching mode. In asymmetric stretching of CH_3 mode, the two C-H bonds of the methyl group are expanding while the third one is contracting. In symmetric stretching of CH_3 mode, all the three C-H bonds expand and contract in-phase. The asymmetric stretching for the CH_3 has higher magnitude than the symmetric stretching [Sudha et al., 2011, Varsanyi, 1969 and Socrates, 1980]. In ECM3CTC, the $C_{20}-H_{21}$ and $C_{20}-H_{22}$ of symmetric and asymmetric stretching vibrations of methyl group did not occur in both FT-IR and FT-Raman region. The expected theoretically values obtained by B3LYP/6-311++G(d,p) method at 3212, 3128 cm^{-1} (mode numbers 6, 10). The substitution of chlorine atom to methyl group has affected the C-H vibration to higher value.

4.3. NBO Analysis

NBO analysis is used for studying intra molecular bonding interactions, and also a means to investigating charge transfer or conjugative interaction in molecular systems. Of the different

molecular orbitals, some are electron donors and some are acceptors, the energy difference between such bonding and anti-bonding orbitals makes the molecule susceptible for interactions. The bonding, non-bonding and anti-bonding interactions can be quantitatively described by means of the second-order perturbation interaction energy $E^{(2)}$. This energy represents the estimation of the off-diagonal Fock matrix elements and can be deduced from the second-order perturbation as [James et al., 2006, Na et al., 2005, Liu et al., 2005]

$$E^{(2)} = \Delta E_{ij} = q_i \frac{F(i, j)^2}{\epsilon_j - \epsilon_i} \quad (5)$$

where q_i is the donor orbital occupancy, ϵ_i and ϵ_j are diagonal elements (orbital energies) and $F(i, j)$ is the off diagonal NBO Fock matrix elements. The larger energy difference $E^{(2)}$ value, the more intensive is the interaction i.e. more is donating tendency of electron from one orbital and more is the accepting tendency of other orbital, which makes the interaction between them stronger.

Table 3. The second order perturbation theory analysis of Fock Matrix in NBO basis for ECM3CTC

Type	Donor NBO (i)	ED/e	Acceptor NBO (j)	ED/e	^a E ⁽²⁾ KJ/mol	^b E(j)-E(i) a.u.	^c F(i,j) a.u.
$\pi - \pi^*$	BD (2) C1 - C6	1.659 63	BD*(2) C2 - C3	0.3737	86.86	0.27	0.067
			BD*(2) C4 - C5	0.32409	88.24	0.28	0.069
$\pi - \pi^*$	BD (2) C2 - C3	1.681 65	BD*(2) C1 - C6	0.34885	79.24	0.3	0.068
			BD*(2) C4 - C5	0.32409	80.25	0.3	0.067
$\pi - \pi^*$	BD (2) C4 - C5	1.668 47	BD*(2) C1 - C6	0.34885	80.33	0.29	0.066
			BD*(2) C2 - C3	0.3737	87.07	0.27	0.068
$\pi - \sigma^*$	BD (2) C11 - C14	1.845 52	BD*(1) C1 - C10	0.04684	14.27	0.67	0.044
			BD*(2) C11 - C14	0.24204	14.77	0.31	0.03
			BD*(1) C20 - H21	0.01685	7.57	0.72	0.034
			BD*(2) C24 - O25	0.30813	98.87	0.29	0.077
$\pi - \pi^*$	BD (2) C12 - O17	1.987 16	BD*(2) C12 - O17	0.33555	8.45	0.46	0.03
$\pi - \pi^*$	BD (2) C24 - O25	1.979 55	BD*(2) C11 - C14	0.24204	20.71	0.4	0.042
$n - \sigma^*$	LP (1) N16	1.743 31	BD*(1) C1 - C10	0.04684	34.39	0.66	0.069
			BD*(1) C12 - O17	0.0377	11.51	0.83	0.045
			BD*(2) C12 - O17	0.33555	190.25	0.32	0.11
$n - \sigma^*$	LP (1) O17	1.975 16	BD*(1) C12 - N16	0.07435	11.55	1.14	0.051
			BD*(1) C12 - N18	0.0902	8.79	1.08	0.043
$n - \sigma^*$	LP (2) O17	1.843 77	BD*(1) C12 - N16	0.07435	103.47	0.7	0.12
			BD*(1) C12 - N18	0.0902	118.24	0.64	0.123

$n - \pi^*$	LP (1) N18	1.662 2	BD*(2) C11 - C14	0.24204	180.25	0.3	0.104
			BD*(2) C12 - O17	0.33555	182.13	0.32	0.106
$n - \sigma^*$	LP (2) C123	1.983 62	BD*(1) C20 - H21	0.01685	11.67	0.77	0.041
			BD*(1) C20 - H22	0.0158	8.08	0.78	0.035
$n - \sigma^*$	LP (3) C123	1.960 56	BD*(1) C14 - C20	0.03655	18.12	0.69	0.049
			BD*(1) H15 - N18	0.02652	16.19	0.78	0.049
$n - \sigma^*$	LP (1) O25	1.974 41	BD*(1) C11 - C24	0.05791	12.55	1.14	0.053
$n - \sigma^*$	LP (2) O25	1.848 79	BD*(1) C11 - C24	0.05791	70.96	0.71	0.101
			BD*(1) C24 - O26	0.10017	138.62	0.62	0.13
$n - \sigma^*$	LP (1) O26	1.957 69	BD*(1) C20 - C123	0.01951	7.03	0.66	0.03
			BD*(1) C24 - O25	0.01801	30.54	1.16	0.083
$n - \pi^*$	LP (2) O26	1.807 13	BD*(2) C24 - O25	0.30813	190.08	0.33	0.113
			BD*(1) C27 - H28	0.01952	17.41	0.78	0.053
			BD*(1) C27 - H29	0.01943	17.15	0.78	0.053
$n - \sigma^*$	LP (2) C135	1.973 7	BD*(1) C2 - C3	0.02575	15.77	0.88	0.052
			BD*(1) C3 - C4	0.02665	15.61	0.87	0.051

^a $E^{(2)}$ means energy of hyper conjugative interaction (stabilization energy).

^b Energy difference between donor (i) and acceptor(j) nbo orbitals.

^c $F(i, j)$ is the Fock matrix element between i and j nbo orbitals.

The $E^{(2)}$ energy values and types of the transition are listed in Table 3. The strong intra-molecular hyperconjugative interaction of the σ and π electrons of C-C, C=C, C=N to the anti C-C, C=C, C=N bonds of the ring as well as C=O group leads to stabilization of some part of the ring system in ECM3CTC. The hyperconjugative interactions in this molecular are $\pi(C_4-C_5) \rightarrow \pi^*(C_1-C_6)$, $\pi(C_1-C_6) \rightarrow \pi^*(C_2-C_3)$, $\pi(C_4-C_5) \rightarrow \pi^*(C_2-C_3)$, $\pi(C_1-C_6) \rightarrow \pi^*(C_4-C_5)$, $\pi(C_{11}-C_{14}) \rightarrow \sigma^*(C_{24}-O_{25})$ with corresponding stabilization energy: 80.33, 86.86, 87.07, 88.24 & 98.87 KJ/Mol respectively. In ECM3CTC, the lone pair of oxygen, and nitrogen atoms play great role, i.e. the $O_{17}, O_{25}, N_{18}, O_{26}$ & N_{16} atoms transfer maximum energy 118.24, 138.62, 180.25, 182.13, 190.08 & 190.25 KJ/Mol to $(C_{12}-N_{18}), (C_{24}-O_{26}), (C_{11}-C_{14}), (C_{12}-O_{17}), (C_{24}-O_{25})$ & $(C_{12}-O_{17})$ bonds, respectively.

In hetero cyclic atoms, the maximum hyperconjugative $E^{(2)}$ energy exhibited during the inter-molecular interaction, which leads the molecule towards medicinal and biological application. According to the stabilization energy values, the first five most probable electronic transitions in this molecule in the descending order are: 1. C2 - C3 to C4 - C5 ($\pi^* - \pi^*$ and $E^2 = 249.13$), 2. C2 - C3 to C4 - C5 ($\pi^* - \pi^*$ and $E^2 = 238.8$), 3. LP(1)N16 to C12 - O17 ($n - \pi^*$, $E^2 = 45.47$), 4. LP(1)N18 to C12 - O17 ($n - \pi^*$, $E^2 = 43.53$), and 5. LP(1)N18 to C12 - O17 ($n - \pi^*$, $E^2 = 43.08$). But among the five most probable transitions, only one or two will take place with high intensity which can be observed in the experimental UV-Visible spectrum also, whose values are determined by the high oscillator strength value.

4.4. Molecular docking

Molecular docking studies were carried out to study the precise binding site of ligand on protein. A molecular docking study is a key tool in structural molecular biology and computer-assisted drug design. Molecular docking trials for compound ECM3CTC was performed by using Glide module implemented in Maestro version 9.3.5 of Schrödinger software suite, 2011. The 3D co-ordinates of crystallographic structure of the Dengue virus NS2B/NS3 protease was retrieved from the Protein Data Bank (PDB) using PDB ID: 3LD6. The Ketoconazole protein (PDB ID: 3LD6) was downloaded from protein Data Bank (<http://www.rcsb.org/pdb>). The molecular docking studies of the ligands and the protein were performed by GLIDE. Glide provides three different levels of docking precision, high throughput virtual screening (HTVS), standard precision (SP), extra precision (XP). Calculations were carried out in XP mode.

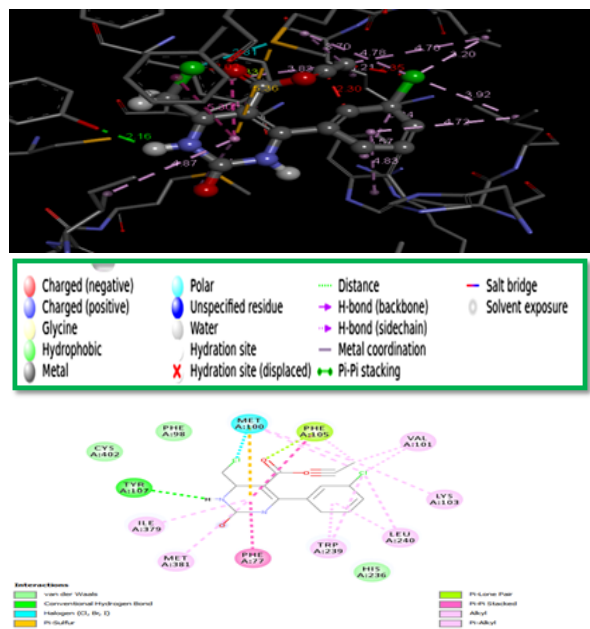


Fig. 4. 2D and 3D interactions of ECM3CTC

For the given structures of a ligand and a protein, docking problem is to predict the structures of the resulting complex. The geometry of the complex reflects the global minimum energy [Chamberlain, 1974]. The synthesized analogue docked with crystal structure of human lanosterol 14- α dimethylase in complex with ketoconazole (3LD6). Analogue showed best ligand pose energy 13.5 kcal/mol in 3LD6 protein. The possible number of protein ligand interactions were predicted and shown in Fig.4. The ligand is surrounded by hydrophilic interactions with the amino acids like PHE 104, TYR107, ILE379, MET100, LEU240, MET381, PHE77, VAL101, PHE105 and TRP 239. The TYR107 amino acid interacts with pyrimidine ring and PHE 105 interacts with carboxylate group via hydrogen bonding interaction. The Vander walls interaction within PHEA:98, CYSA:402 and HISA:236 are surrounding the interaction. The protein-ligand interaction value indicates that the synthesized compound ECM3CTC exhibits better anti-inflammatory property. From this interaction it can be predicted as the activity may be due to inhibition of human lanosterol 14- α dimethylase in complex with ketoconazole.

4.5. UV-Analysis

The UV-Vis absorption spectrum of ECM3CTC is recorded in the range 200-800 nm. The UV-Vis spectral analyses of the molecule have been investigated in Gas phase, Chloroform and DMSO by TD-SCF/B3LYP/6-311++G(d,p) calculations. The electronic transitions, the corresponding excitation energies and the oscillator strengths for the three phases are presented Table 4. From the table, it is found that the electronic transition of ECM3CTC with high oscillator strengths and low absorption wavelength for all the three phases, thereby giving rise to significant electronic properties. The observed and simulated UV-Vis spectra are shown in Fig. 5. Typically, according to the Frank-Condon principle, the maximum absorption peak (λ_{max}) in a UV-Vis spectrum corresponds to vertical excitation.

The theoretical calculation predicts one intense electronic transition at 303 nm with an oscillator strength $f = 0.1233$ is in agreement with the experimental data $\lambda_{exp} = 300$ nm. Since the absorption wavelength lies in the $n-\pi^*$ transition, according to NBO analysis, the following transitions causes the UV-Visible transition. I) LP(1)N 16 to C 12 - O 17 ($n-\pi^*$, $E^2 = 45.47$), II) LP(1)N 18 to C 12 - O 17 ($n-\pi^*$, $E^2 = 43.53$), and III) .LP(1)N 18 to C 12 - O 17 ($n-\pi^*$, $E^2 = 43.08$).

According to the oscillator strength value, only the second transition in the above list appear in the theoretical spectrum. However, the experimental spectrum shows that there is overlapping of all these most probable transitions as their oscillator strengths are close to each other.

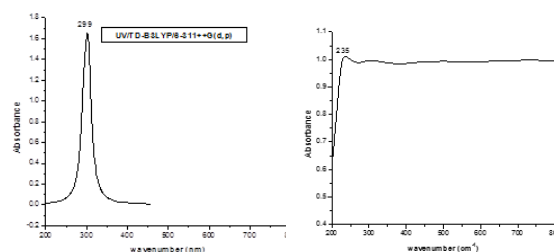


Fig. 5. The combined calculated and experimental UV Vis spectra of ECM3CTC

Table 4. The electronic transition of ECM3CTC using B3LYP/6-311++G(d,p) basis set

Calculated at B3LYP/6-311++G(d,p)	Oscillator strength	Calculated Band gap (eV/nm)	Experimental Band gap (eV/nm)	Type
Excited State-1	Singlet-A ($f=0.0122$)	4.0069eV/309.43nm		$n-\pi^*$
83 -> 86	0.20686			
84 -> 86	0.59833			
85 -> 86	-0.27667			
Excited State-2	Singlet-A ($f=0.1233$)	4.0862eV/303.42nm		$n-\pi^*$
82 -> 86	0.10804			
83 -> 86	0.28378			
84 -> 86	0.12259			
85 -> 86	0.55827			
Excited State-3	Singlet-A ($f=0.0674$)	4.2228 eV/293.61nm	235	$\pi-\pi^*$
80 -> 86	0.17095			
81 -> 86	0.12911			
83 -> 86	0.5481			
84 -> 86	-0.2977			
85 -> 86	-0.17863			

4.6. NMR Spectra Analysis

NMR chemical shift analysis is used to elucidate the types of functional groups in the molecules or samples. It is also used to predict details about the composition and chemical environment of atoms in the molecule. The chemical shifts were computed for H and C atoms in the molecule using B3LYP functional and 6-311++G(d,p) basis set along with GIAO functional, which is meant for this purpose. The same was also experimentally measured using ^1H and ^{13}C NMR spectrometer and the results were presented in Table 5. The ^1H and ^{13}C experimental shifts in ppm for the molecule ECM3CTC are also given in Figs. 6a & 6b. The chemical shift for carbon atoms in benzene ring is expected in the range 100-120 ppm, this is found to be the case in this molecule also except for C1 whose value is found to be 130 ppm. This observation is in accordance with the charge prediction by MEP where all carbon atoms within benzene are equally negative except C1. Because of the positive charge on C1 the deshielding is more on this atom compared to other atoms which lead to higher chemical shift. The same trend is also observed in Pyrimidine ring, where all positively charged C atoms C12 & C14 are found to have higher chemical shift

due to increased deshielding. The other two carbon atoms C10 & C11 due to the enhanced negative charges show much shielding effect, hence their chemical shift is proportionately less 47.31 & 89.93 ppm respectively. Hence, the chemical shift values clearly distinguishes benzene ring and pyrimidine ring and demonstrates that these two rings have different types of conjugations. The carbon atoms in the carboxylate groups have chemical shift values clearly less than 50 ppm which indicate they are aliphatic. The C atom which is attached with the chlorine atom shows the least value around 5 ppm, this indicates the complete change in the environment of this atom.

The chemical shift for all ¹H atoms of ECM3CTC fall in the range of 1.25 – 8.08 ppm. The ¹H chemical shift values of aromatic ¹H atoms have the range 7– 8 ppm. The H atoms in the pyrimidine ring show the values in the range 4 to 6 ppm while in the aliphatic chain around 1 ppm. Thus there is clear distinction in the values of H atoms chemical shifts showing the different types of electronic distribution, even though this is not explicitly visible in charge analysis or in vibrational analysis. The observed ¹H and ¹³C NMR spectra of ECM3CTC is plotted in Figs. 6a & 6b.

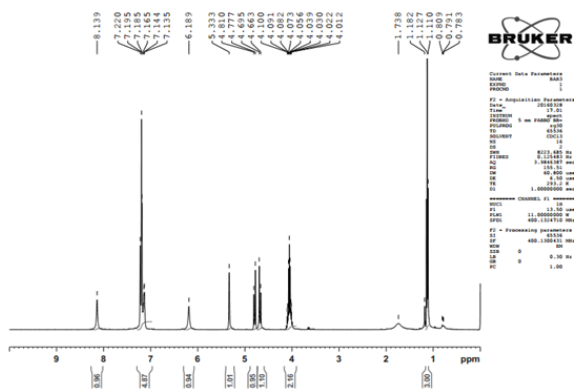


Fig. 6a. The ¹H-NMR spectrum of ECM3CTC

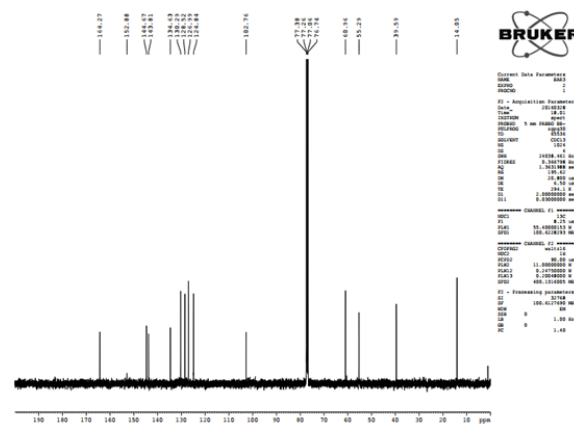


Fig. 6b. The ¹³C-NMR spectrum of ECM3CTC

Table 5. Theoretical and experimental ¹H and ¹³C chemical shift (ppm) of ECM3CTC

Atoms	Isotropic chemical shielding tensor (σ) (ppm)	Shift (ppm)	Experimental values Ppm
C1	50.96	131.5	
C2	68.86	113.59	
C3	52.33	130.13	
C4	69.13	113.33	
C5	67.87	114.58	
C6	74.53	107.90	102.76
C10	135.15	47.31	
C11	92.33	89.93	
C12	48.64	133.81	134.63
C14	52.68	129.78	130.29
C20	141.13	41.33	
C24	32.66	149.93	152.88
C27	129.93	52.55	55.29

C30	177.22	5.771	
7H	23.79	8.08	8.139
8H	24.43	7.44	
9H	24.55	7.32	7.220
13H	27.82	4.06	
15H	25.16	6.71	6.189
19H	26.28	5.57	5.333
21H	27.31	4.56	
22H	26.98	4.90	4.810
28H	27.52	4.35	
29H	27.55	4.32	
31H	30.62	1.25	1.182
32H	30.44	1.43	
33H	30.42	1.45	
34H	24.61	7.27	

5. CONCLUSION

The title molecule was synthesized and characterized by spectral analysis such as FTIR, FT-Raman, UV-Vis and NMR spectral studies. A complete vibrational analysis was carried out for the first time for the molecule ECM3CTC. The bond parameters were calculated and also compared with the reported XRD data. NBO analysis reflects the charge transfer occur within the molecule. The Lewis and non-Lewis structure of ECM3CTC provides ED contribution of individual atoms, which leads the molecular vibration with respect to hybridization of chemical bonds. The conjugation of pyrimidine ring π -orbital with carboxylate group σ^* -orbital was found to be strong when both orbitals lie in the same plane. Molecular docking studies revealed that the target molecule ECM3CTC exhibited appreciable interaction with 3LD6 protein and the protein-ligand interaction value indicates that the title compound exhibits better anti-inflammatory property. The observed UV-Vis absorption bands are assigned to $n-\pi^*$ & $\pi-\pi^*$ transition. The chemical shift for all ¹H atoms of ECM3CTC fall in the range of 1.25 – 8.08 ppm. The carbon atoms in the carboxylate groups have chemical shift values clearly less than 50 ppm which indicate they are aliphatic. The C atom which is attached with the chlorine atom shows the least value around 5ppm, this indicates the complete change in the environment of this atom. There is clear distinction in the values of H atoms chemical shifts showing the different types of electronic distribution, and therefore confirm the formation of target compound.

REFERENCES

1. Alam, M., and Lee, D. U. (2016), 64, 185-201.
2. Amir M, Javed SA and Kumar H; Indian J. Pharm. Sciences, 2007; 69(3): 337-343.
3. Amr EA, Nermien MS, Abdulla MM. Monatsh Chem, 2007; 138: 699- 707.
4. Bharanidharan, S., Saleem, H., Gunasekaran, B., Syed Ali Padusha, M., Suresh, M. Crystal structure of ethyl 6-(chloromethyl)-4-(4-chlorophenyl)-2-oxo-1,2,3,4-tetrahydropyrimidine-5-carboxylate, Acta Cryst. 2014, E70, 1185–1186.
5. Beena, K.P., Akelesh, T. Int. Res. J. Pharm. 2012, 3, 303–304.
6. Bist, H.D., Brand, J.C.D., Williams, D.R. J. Mol. Spectrosc. 1967, 24, 402–412.
7. Chamberlain, N. F. The Practice of NMR Spectroscopy with Spectra-Structure Correlations for Hydrogen-1, Plenum Press (1974).
8. Colthup, N.B., Paly, L.H., Wiberley, S.E. Introduction to Infrared and Raman Spectroscopy, third ed., Academic press, Boston, MA, 1990.
9. Dhandapani, A., Manivarma, S., Subashchandrabose, S., Chem. Phys. Lett. (2016), <http://dx.doi.org/10.1016/j.cpl.2016.04.009>.
11. Desai K, Patel R, Chikhalia K. J Ind Chem, 2006; 45: 773-778.
12. Evans, J.C. Spectrochim. Acta. 1960, 16, 1382–1392.
13. Frisch, M.J., Pople, J.A. Gaussian 03, Revision C.02, Gaussian Inc., Wallingford, CT, 2004.
14. Fujiwara N, Nakajima T, Ueda Y, Fujita HK, Awakami H. Bioorg Med Chem, 2008; 16: 9804-9816
15. Geskin, V.M., Lambert, C.J., Bredas, L. Origin of high second and third-order nonlinear optical response in ammonio/borato diphenylpolyene zwitterions: the remarkable role of polarized aromatic groups, J. Am. Chem. Soc. 125 (2003) 15651–15658.
16. James, C., Amalraj, A., Reghunathan, R., Hubert Joe, I., JayaKumar, V.S., J. Raman Spectrosc. 37 (2006) 1381–1392.
17. Jamroz, M.H. Vibrational Energy Distribution Analysis: VEDA4 program, Warsaw, Poland, 2004.
18. Khalaji, A. D., Chermahini, A. N., Fejfarova, K. & Dusek, M. Journal of the Structural Chemistry, (2009) 21, 153-157.
19. Kleiman, D.A., phys. Rev. 126 (1962) 1977.
20. Kleinman, D.A., (1962). Nonlinear Dielectric Polarization in Optical Media, Physics Reviews, 126, 1977-1979.
21. Liu, J., Chen, Z., and Yuan, S. Journal of Zhejiang University Science, 6B(6)(2005), 584-589.
22. Meganathan, C., Sebastian, S., Kurt, M., Lee, K. W., & Sundaraganesan, N. Journal of Raman Spectroscopy, 41(10), (2010), 1369-1378.
23. Michalska, D. Raint Program, Wroclaw University of Technology, 2003.
24. Michalska, D. and Wysokinski, R. Chem. Phys. Lett., 403 (2005) 211-217.
25. Mooney, M.F., Spectrochim. Acta 20 (1964) 1021.
26. Mooney, E.F., Spectrochim. Acta 19 (1963) 877.
27. Na, L.J., Rang, C.Z., Fang, Y.S., Zhejiang J., Univ. Sci. 6B (2005) 584–589.
28. Stanisaw, Jord. J. Chem. (2009), 4: 1-15.
29. Radom, L., John A. Pople, J. American. Chem. Soc. 92 (1970) 4786-4795
30. Roeges, N.P.G. A Guide to the Complete Interpretation of Infrared Spectra of Organic Compounds, Wiley, New York, 1994.

32. Sarojini, K., Krishnan, H., Charles Kanakam, C., Muthu, S., Spectrochim. Acta A108 (2013) 159-170
33. Schlegel, H.B. J. Comput. Chem. 1982, 3, 214–218.
34. Sharma, A.; Khare, R.; Kumar, V.; Gupta, G. K.; Beniwal, V. Int. J. Pharm. Pharm. Sci. 2014, 6, 171–175.
35. Sasada, T., Kobayashi, F., Sakai, N., Konakahara, T., Organic Letters, 2009; 11: 2161-2164.
36. Scott, A.P., Random, L., J. Phys. Chem. 100 (1996) 16502-16513.
37. Silverstein, R.M., and Webster, F.X. Spectrometric identification of organic compounds sixth edn. Wiley, New York (1998).
38. Smith, B., Infrared Spectral Interpretation. A Systematic Approach, CRC Press, Washington, D C, 1999.
39. Socrates, G. Infrared Characteristics Frequencies, Wiley-interscience Publication, New York, 1990.
40. Socrates, G. Infrared and Raman Characteristic group Frequencies, Tables and Charts, 3rd Ed. Wiley, Chichester (2001).
41. Sudha, S., Karabacak, M., Kurt, M., Cinar, M., Sundaraganesan, N. Spectrochim. Acta A 84 (2011) 184-195.
42. Sundaraganesan, N., Meganathan, C., Joshua, B.D., Mani, P., Jayaprakash, A., Spectrochim. Acta A 71 (2008) 1134-1139.
43. Vaisalini, N.B., Rao, N.V., Harika, V.B.M.L., Desu, P.K., Nama, S., Int. J. Pharm. Chem. Res. 2012,
44. Varghese, H. T., Panicker, C. Y., Madhavan, V. S., Mathew, S., Vinsova, J., & Van. Alsenoy, C. Journal of Raman Spectroscopy, 40(9), (2009), 1211-1223
45. Varsanyi, G., Assignments for Vibrational Spectra of Seven Hundre Benzene Derivatives, vol. 1/2, Academic Kiado, Budagset, (1973), 2278–8700.
46. Varsanyi, G. Vibrational Spectra of Benzene Derivatives, Academic Press, New York, 1969.
47. Wagner E, Al-Kadasi K, Zimecki M, Sawka-Dobrowolska W. Eur J Med Chem, 2008; 43: 2498-2504.