



SCREENING OF *NYMPHAEA TETRAGONA* GEORGI, A TRADITIONAL TAXON FOR BIOACTIVE METABOLITES

K. Siva Prasad	Department of Botany, S. V. University, Thirupati-517502, A. P. India.
K. Sujatha*	Department of Botany, S. P. W. Degree & P. G. College Tirupati. *Corresponding Author
D. Suhurulatha	Department of Botany, NBKR Science & Arts degree college, Vidhyanagar, Nellore.
N. Savithramma	Department of Botany, S. V. University, Thirupati-517502, A. P. India.

ABSTRACT Bioactive metabolites or phytochemical constituents of medicinal plants performing a crucial role in drug industry as well as in identification of elite germplasm Globally distributed *Nymphaea tetragona* Georgi, an aquatic and Ayurvedic herb with perennial rhizomes and white flowers. Studies on phytochemical evaluation for validation in aquatic plants particularly in *Nymphaea* sps. is scanty. Hence the present work is aimed to screen for the analysis of qualitative and quantitative bioactive metabolites present in various parts of the *N. tetragona* for potential applications. Polar and non-polar solvents were used to extract maximum no of compounds from plant parts. The results revealed that the leaf and flower are the rich source of different phytochemicals like anthocyanins, anthraquinones, emodins, fatty acids, flavonoids, leucoanthocyanins, glycosides, phenols, coumarins, tannins and triterpenoids when compare with stem and root. The findings of the study will be helpful to the pharmacologists and phytochemists for identification of an effective and novel phytochemicals.

KEYWORDS : aquatic plant, bioactive metabolites, *Nymphaea tetragona*, aqueous extract, methanol extract, ethanol extract and chloroform extract.

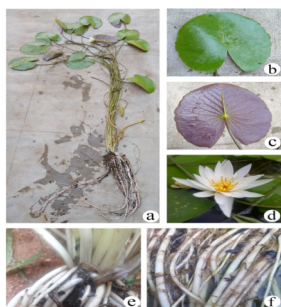
INTRODUCTION

In many spiritual traditions, lotus flowers are sacred as an offering to the divine. Although the lotus is commonly seen in Buddhist art and literature, it has been a religious symbol in Hindu culture much before the onset of Buddhism. Lotus flowers are divided in to two genera like *Nelumbo* and *Nymphaea*. *Nymphaea* have many wild species (*N. alba*, *N. candida*, *N. nouchali*, *N. pubescens*, *N. rubra* and *N. tetragona*) and cultivated (*N. caerulea*, *N. marliacea*, *N. micrantha* and *N. alba*). The flowers are used in Ayurvedic medicine in wine, dyspepsia, enteritis, diarrhoea, urinary problems, fevers and heart palpitations. Phytochemicals which are naturally occurring in the medicinal plants for defense mechanism and protect from various diseases. Phytochemicals mainly secondary constituents like terpenoids, alkaloids and phenolic compounds were observed in the flowers of *N. caerulea*². Through the plant is used conventionally to cure several diseases, reports on evaluation of bioactive metabolites in *N. tetragona* is not carried out so far. Therefore the present study is undertaken to screen the plant for bioactive metabolites present in the root, rhizome, leaf and flower of *N. tetragona* as secondary metabolites present in plants are responsible for curing number of diseases and the plant had been using to treat several affections by our ancient people.

MATERIAL AND METHODS

flowers, leaves, rhizomes and Roots, (Fig.1 d b, c, e and f,) of *Nymphaea tetragona* (Fig. 1. a) were collected from stacked water near Bokkasenugutta, Sibyalala, Rayachoty Mandal, Kadapa District of Andhra Pradesh and washed the parts with tap water and allowed to dry under shade. The dried roots, rhizomes, leaves and flowers were made in to fine powders and stored separately. These powders were primarily extracted separately with water, ethanol, methanol and chloroform; then the preliminary tests were carried out for the detection of primary and secondary metabolites present in these plant parts.

Fig.1. a) Plant b) Leaf abaxial c) Leaf adaxial d) Flower e) Rhizome and f) Roots,



1. Water extraction

10 g of shade dried powders of plants were added to 100 ml of distilled water and boiled for 2 h. The supernatant were collected and same procedures were repeated twice. The collected supernatants at an interval of every 2 h were pooled together and concentrated to make the final volume into one-fourth of the original volume. It was then autoclaved at 121°C and 15 lbs pressure and stored in separate bottles at 4°C.³

2. Chloroform, Ethanol and Methanol extraction

Air dried powders of roots, rhizome, leaves and flowers were extracted with chloroform, ethanol and methanol separately. The crude extracts were prepared by maceration of plant materials (100 g each) with (300 ml) of each solvent⁴. The extracts were filtered and stored in separate bottles.

Screening of Bioactive compounds

The condensed extracts were used for preliminary screening of primary and secondary metabolites such as proteins⁵, starch⁶, sugar⁷, flavonoids⁸, phenols, alkaloids, lignin and steroids⁹, saponins and Terpenoids¹⁰, glycosides¹¹, quinones¹², coumarins¹³, tannins, leucoanthocyanins and emodins¹⁴.

RESULTS AND DISCUSSION

Qualitative and quantitative analysis of the *N. tetragona* revealed that the presence of bioactive metabolites like alkaloids, anthocyanins, anthraquinones, coumarins, emodins, flavonoids, glycosides, leucoanthocyanins, phenols, starch, steroids, sugars, saponins, tannins and terpenoids in flower, leaves, roots and rhizomes as shown in Table 1 and 2. Similar result was observed in the flower, leaves, roots and rhizomes of *N. caerulea*¹⁵.

The primary metabolites like starch, sugars and proteins are also identified from four parts of the plant. The primary metabolites are carbon compounds which are essential to life. All organisms will use primary metabolites as building blocks of the cells and exploit their rich supply of potential energy to maintain life. The rhizome of *N. tetragona* is rich in protein (2.3 gm/dwt) when compare to other plants (Table-2). The presence of higher protein level in plant parts indicates towards possible increase in food value and protein based bioactive compounds could also be isolated in future. Proteins are vital for growth and repair of cells, this may be the reason most of the tribal people store the rhizome, they eat cooked rhizomes. Starch is a nutrient polysaccharides found in rhizome at higher level when compared to other organs of *N. tetragona* (Table-2). These are omnipresence contain storage organs like tubers, seeds and stems are particularly abundant in them¹⁶. Leaves and flowers of *N. tetragona* also source of starch. But there is no indication in chloroform and methanol extracts.

Sugars found in all extracts of leaves whereas aqueous and ethanolic extracts the flowers contain the sugars and absent in aqueous and chloroform extract (Table-1). The four extracts of roots and rhizomes indicate the absence of sugars. The sugar are help in translocation of food in phloem and protects the plants from desiccation¹⁷. Sugars are found at higher levels in flowers followed by leaves, rhizomes and roots. The reason could be that the flowers may contain sugars in the form of honey. The phenols are appeared in all plants of aqueous extract and leaves of chloroform, ethanol and methanolic extracts of flowers. The leaves showed maximum levels of phenols when compare with flowers, rhizome and root. The high amounts of phenols are important in regulation of growth, development and disease resistance and plant phenols have anti oxidant and cardio protective activities¹⁸. They are also used as fungicides, antiseptic disinfectant and in manufacturing of resins.

The aquatic extract contains maximum levels of alkaloids. These are the most diverse group of secondary metabolites present in living

organisms with all arrays of structural types in biosynthetic pathways and various pharmacological activities. The anthocyanins found which are in the leaves, flowers and roots of *N. tetragona* and absent in rhizomes. Anthocyanins possess viridic effect. Not only decreasing influenza viral infection but also prevent the entry of virus to human cell¹⁹. The leaf and flower extracts of aqueous ethanol, methanol and chloroform showed the coumarin, which are potential anti oxidants by scavenging free radicals and chelating metal ions²⁰. Emodins found in the flowers of all extracts. Emodins are one of the natural anti cancer drugs Chinese used as anti cancer drugs in the traditional medicine²¹. Phytochemical screening helps to reveal the chemical nature of the constituents of the plant extract which may also be used to search for bioactive agents that could be used in the synthesis of very vital drugs²². Flavonoids are present in flower, leaf, root and rhizome extracts of aqueous and absent in chloroform extract, root and rhizome extracts of ethanol and methanol. As flavonoids are water soluble and potent as free radical scavengers, super antioxidants, prevent oxidative cell damage and have strong anti-cancer activity²³.

Table 1. Qualitative analysis of bioactive metabolites of various parts of *N. tetragona*

S. No	Phytochemical constituents	Aqueous				Chloroform				Ethanol				Methanol				
		F	L	R	Rh	F	L	R	Rh	F	L	R	Rh	F	L	R	Rh	
1.	Starch	+	+	-	-	-	+	-	-	+	+	-	-	-	+	-	-	+
2.	Sugars	+	+	-	-	-	+	-	-	+	+	-	-	-	+	-	-	+
3.	Proteins	+	+	-	+	-	+	-	-	+	+	-	-	+	+	+	+	+
4.	Alkaloids	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
5.	Anthocyanins	+	+	+	-	-	-	-	-	+	+	-	-	+	+	-	-	-
6.	Anthraquinones	+	+	+	-	-	-	-	-	+	+	+	-	+	+	-	-	-
7.	Coumarins	+	+	-	-	+	+	-	-	+	+	-	-	+	+	-	-	-
8.	Emodins	+	+	-	-	-	-	-	-	-	+	-	-	+	-	-	-	-
9.	Flavonoids	+	+	-	+	-	+	-	-	+	+	+	-	+	+	-	-	-
10.	Glycosides	-	-	-	-	-	+	-	-	-	-	-	-	+	+	-	-	-
11.	Leucoanthocyanins	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
12.	Steroids	+	+	-	+	+	-	-	-	+	+	-	+	+	+	-	-	+
13.	Phenols	+	+	+	+	-	+	-	-	+	+	-	-	+	+	-	-	-
14.	Saponins	+	+	+	+	+	-	-	-	+	-	-	-	+	+	+	-	-
15.	Tannins	+	+	-	+	-	+	-	-	+	+	-	+	+	+	-	-	-
16.	Triterpenoids	+	+	-	-	-	-	-	-	-	+	-	-	+	-	-	-	-

Note: '+' indicates presence, '-' indicates absence (F) Flower, (L) Leaf, (R) Root and (Rh) Rhizome)

Table 2. Quantitative analysis of bioactive metabolites of various parts of *N. tetragona*

S. No	<i>N. tetragona</i>	Secondary bioactive metabolites			Primary bioactive metabolites		
		Flavonoids mg/g dwt	Phenols mg/g dwt	Tannins mg/g dwt	Starch mg/g dwt	Sugar mg/g dwt	Proteins mg/g dwt
1	Flower	0.026 ± 0.002	280 ± 2.943	0.13 ± 0.008	3.7 ± 0.061	6.9 ± 1.632	1.8 ± 0.163
2	Leaf	0.0127 ± 0.001	285 ± 4.198	0.16 ± 0.012	3.7 ± 0.526	4.0 ± 0.816	1.8 ± 0.081
3	Rhizome	-	210 ± 3.681	-	4.1 ± 0.552	3.4 ± 1.632	2.3 ± 0.205
4	Root	-	180 ± 2.943	-	33.7 ± 0.526	2.3 ± 1.632	1.2 ± 0.124

Note: '-' indicates absence and ± indicates stranded error

Glycosides are present in flower and leaf extracts of aqueous, chloroform, ethanol and methanol and root extract of ethanol, rhizome extracts of aqueous and ethanol. They are absent in root extracts of aqueous, chloroform and methanol, rhizome extracts of chloroform methanol. Glycosides compounds are containing a carbohydrate and non-carbohydrate residues (moiety) in the same molecule. The carbohydrate moiety is attached to an acetyl linkage carbon-I to the non-carbohydrate residue (aglycone). They all contain steroid as aglycone component in combination with sugar molecules. They are important in medicine because of their action on heart and are used in cardiac insufficiency²⁴.

Leucoanthocyanins are present in flower and leaf extracts of aqueous only and absent in chloroform, ethanol and methanol of flower, leaf, root and rhizome extracts. Leucoanthocyanins are occupying an important position among the water soluble organic compounds. They have been implicated as being responsible for the astringent taste of unripe fruit they are also responsible for the chill haze that develops in beer and for the browning of white wine. They influence the storage stability of wine and juice²⁵. Steroids are present in flower and leaf extracts of aqueous, ethanol and methanol; flower extract of chloroform, stem extract of aqueous, chloroform, ethanol and methanol and absent in leaf extract of chloroform, root extract of aqueous, chloroform, ethanol and methanol. Steroids are important in pharmacy as they related to sex hormones.²⁶

Saponins are found in flowers, leaves, roots and rhizome extracts of aqueous; flowers, leaves and rhizome extracts of chloroform; flower

and rhizome extracts of ethanol, leaves and roots extracts of methanol and absent in root extract of chloroform and ethanol; leaf extracts of ethanol, flower and rhizome extracts of methanol. The saponins are used as detergents, pesticides and molluscides along with their industrial applications in foaming and surface active agents. They are involved in controlling of onset of labour pains along with oxytocin²⁷.

Tannins are present in flowers, leaves, roots and rhizome extracts of aqueous; leaf extract of chloroform; flower and leaf extracts of ethanol and methanol and absents in flower, root and rhizome extracts of chloroform, root and rhizome extract of ethanol and methanol. Tannins contribution in wound healing and reduced the inflammation of mucus membranes. They possess astringent, anti diarrhoeal and wide pharmaceutical properties²⁸.

Triterpenoids are present in flower and leaf extracts of aqueous, chloroform and methanol. Triterpenoids are absent in flower and leaf extract of ethanol, root and rhizome extracts of aqueous, chloroform, ethanol and methanol. Triterpenoids are attributed for analgesic and anti-inflammatory activities. The results revealed that the saponins and glycosides are found in all extracts used in present work. Aqueous extract is the best among the solvents used for extraction of secondary metabolites. Flowers and leaves are rich source of phytochemical constituents in order.

Based on the screening of bioactive metabolites among the flavonoids, phenols, tannins, starch, sugar and proteins highest content of flavonoids were found in flowers (0.026± 0.002 mg/g dwt) followed by

leaves (0.0127 ± 0.001 mg/g dwt). Flavonoids are reported to possess many useful properties, including anti-inflammatory, antimicrobial, enzyme inhibition, oestrogenic, antiallergic, antioxidant and antitumour activity^{29,30}.

The present study is evident that the *N. tetragona* possess number of medicinal properties, that might be the reason the taxon has been using in traditional events since ancient times. *N. tetragona* is proved that the plant parts are rich in bioactive metabolites like other *Nymphaea* species like *N. caerulea*³¹, *N. pubescens*³², *N. rubra*³³ and can be utilised in potential pharmacological applications.

CONCLUSION.

N. tetragona plant appears to be rich in primary and secondary metabolites particularly rich source of flavonoids, saponins, glycosides and steroids. Widely had used in traditional medicine to combat and cure various ailments. The anti-inflammatory, anti-spasmodic, anti-cancer, anti-microbial, anti-analgesic, anti-oxidants and anti-diuretic activities can be attributed to their high amount of steroids, tannins, terpenoids, flavonoids, phenols, emodins and saponins. Exploitation of these pharmacological properties involves further investigation of these active ingredients by implementation techniques of extraction, purification, separation, crystallization and identification.

REFERENCE

- Mitra RL (1990). Nymphaeaceae. In: Nayar MP, Thothathri K, Sanjappa M (eds.) Fascicles of flora of India, Fascicles 20. Botanical Survey of India, Kolkata. 11–25.
- Sivaprasad K., Savithramma N., (2016). Screening of Phytochemical Constituents of *Nymphaea Caerulea* savigny. An Aquatic Plant Resource for Drug Development American Journal of Advanced Drug Delivery. ISSN 2321-547X
- Parekh Jigna, sumitra CV (2007). Invitro antimicrobial activity and phytochemical analysis of some Indian medicinal plants. Turk J Biol. 31: 53-58.
- Alanis AD, Calzada F and Caervantes JA (2005). Antimicrobial properties of some plants used in Mexican Traditional Medicine for the treatment of gastrointestinal disorders. Ethnopharmacology, 100: 153-157.
- Lowry OH, Rosebrough NJ, Farr AL, Randall RJ. Protein measurement with the folin phenol reagent. J Biol Chem, 1951; 193:265-
- McCready RM, Guggoiz J, Silveira V, Owens HS. Determination of starch and amylase in vegetables. Anal Chem, 1950; 22(9):1156-1158
- DuBoise M, Gilles K, Hamilton J, Rebers PA, Smith F. Colorimetric method for determination of sugars and related substances. Anal Chem, 1956; 28(3):350-356.
- Peach K and Tracey MV (1956). Modern methods of plant analysis. Springer Verlag, Berlin. 3.
- Gibbs RD. Chemotaxonomy of Flowering Plants, Mc Gill Queen's University Press, Montreal and London, 1, 1974.
- Ayoola GA, Coker HAB, Adesegun SA, Adepoju-Bello AA, Obaweya K, Ezennia EC and Atangbayila TO (2008). Phytochemical plants used for malaria therapy in South Western Nigeria. Trop J Pharm Res. 7: 1019-1024.
- Kokate CK, Purohit AP and Gokhale SB (1999). Pharmacognosy, Nirali publishers, Pune, 1-2.
- ASEAN countries (1993). Standard of ASEAN herbal medicine, Jakatra Buena Printing. 1: 116-28.
- Rizk AM, Constituents of plant growing in Qatar. I. A chemical survey of sixty plants. Fitoterapia, (1982); 52-42.
- Treare GE and Evans WC (1985). Pharmacognosy, Bahive Tinal, London. 17: 149.
- Sivaprasad K and Savithramma N, (2016). Screening of Phytochemical Constituents of *Nymphaea Caerulea* savigny. An Aquatic Plant Resource for Drug Development. ISSN 2321-547X
- Sabnis SD and Daniel M (1990). A phytochemical approach to economic Botany, Kalyani Publishers, New Delhi. 15: 65.
- Fogliano V, Verde V, Randazzo G and Ritenti A (1999). Method of measuring antioxidant activity and its application to monitoring the antioxidant capacity of wines. J Agric Food Chem. 47:1035-1040
- Liu F, Zhou N, Cao H, Fang X, Deng L, Chen W, Nin X, Liu L, Zhao H. Chinese college freshmen's mental health problems and their subsequent help-seeking behaviours: A cohort design (2005-2011). Plos One. 12:e0185531. PMID 29040266 DOI: 10.1371
- Tseng A (1991). Chemoprevention of tumors in MTV-H ras transgenic mice with coumarins. Proc. Am. Assoc. Cancer Res., 32:2257.
- Khemkaran A, Jain SK (2011). Aloeemodin novel anticancer herbal drug. In J Phytomed. 3:27-31.
- Sibanda, T., Okoh, A. I. (2008). In vitro evaluation of the interactions between acetone extracts of *Garcinia kola* seeds and some antibiotics. Afr. J. Biotech. 7: 1672-1678.
- Bolwell E G, Rice E and Evans C (1995). Polyphenolic flavonoids as scavenger of aqueous phase radicals and chain breaking antioxidants. Arch Biochem. 2:239-346.
- Balch JF, Balch PA (2000). Prescription for Nutritional Healing. New York. A very.penguin Putnam Inc. 267-270.
- Josly MA, Goldstein L (1953). Astringency of fruit and fruit products in relation to leucoanthocyanin content. Progress report of Agricultural research, The University of California Division of Agricultural Science.
- Santhi, R., Lakshmi G., Priyadarshini A.M., Anandaraj L., 2011. Phytochemical screening of *Nerium oleander* leaves and *Momordica charantia* leaves. Int. Res. J. Pharm., 2: 131-135.
- Okwu DE (2001). Improving the nutrition value of Cassava tapioca meal with local species. Nutraceutical, Functional and medicinal food. 3: 43-51.
- Killedar SG, More HN (2010). Estimation of tannins in different parts of *Memecylonum bellatum*. Burm J Phar Res. 3(3): 554-556.
- Havesteen B (1990). Flavonoids a class of natural products for antimicrobial agents. Eur J Clin Microbial Infect Dis, 9: 455-61.
- Harborne JB, Williams CA (2000). Advance in flavonoid research since 1992. Phytochemistry. 55: 481-504.
- Sivaprasad K, Savithramma N, (2015). Biosynthesis and Validation of SNPs from *Nymphaea caerulea* Savigny. AJADD149-159 ISSN 2321-547X
- Sivaprasad K, Savithramma N, (2016). *Nymphaea rubra* ROXB.- An aquatic source against bacterial proliferation. World Journal of Pharmaceutical Research, Volume 5, Issue 10, 1201-1210. Research Article ISSN 2277-7105

- Sivaprasad K, Savithramma N, (2016). Green Synthesis of Silver Bionanoparticles from Aquatic Resources to Control Bacterial Cell Proliferation. J. Pharm. Sci. & Res. Vol. 8(8), 2016, 721-724