Pathology



PATHOLOGY AND RENAL OUTCOME OF THREE UNCOMMON FACES OF CRESCENTRIC GLOMERULONEPHRITIS

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ABSTRACT AIMS: Rapidly progressive glomerulonephritis (RPGN) presents with rapidly deteriorating renal function (> 50% loss of glomerular filtration rate /GFR within 3 months) associated with nephritic urinary sediments and crescents in biopsy. Crescentric IgA Nephropathy, Anti-GBM (Glomerular basement membrane) disease and combined IgA Nephropathy with Anti-GBM disease are three uncommon reasons of RPGN. We have compared clinicopathological. Immunofluorescence (DIF) and renal outcome of three groups. Setting and designs: Prospective, cross-sectional, single centre study.

METHODS AND MATERIALS: Ultrasonography guided core biopsies obtained, one stained with hematoxylin-eosin, periodic acid-Schiff, Masson's trichrome, and silver methenamine stain another one with immunofluorescence conjugated IgG, IgM, IgA, C3, C1q, kappa and lambda stain. Demographic, clinicopathological and therapeutic parameters with survival data were collected.

STATISTICALANALYSIS USED: Done using software (GraphPad PRISM 6).

RESULTS: We have included 9 cases of crescentric IgA Nephropathies, 6 cases of AntiGBM diseases and 2 cases of combined IgA Nephropathy and AntiGBM diseases. Significant difference seen in the incidences of hypertension, hemoptysis, serum creatinine, anti GBM antibody, total number of crescents and mesangial hypercellularity, fragmentation of GBM etc. IgG, IgA and kappa positivity in DIF show significant difference. Survival analysis and mortality versus dialysis dependence and complete and partial remission versus no remission showed no difference between these three groups.

CONCLUSIONS: Proper and early clinicopathological diagnosis is important since all are of poor renal outcome. Further renal outcome of the combined disease is same as that of individual ones.

KEYWORDS : Rapidly progressive glomerulonephritis; Anti-GBM disease; IgA Nephropathy; crescentric glomerulonephritis; combined IgA Nephropathy with Anti-GBM disease

INTRODUCTION:

Rapidly progressive glomerulonephritis is very rare worldwide. The incidence in the United States of America is around 7 cases per 1 million person-years, while it is 2 cases per 1 million person-years reported in the United Kingdom. There are other reported clusters all over the world, suggesting a possible environmental influence on the pathogenesis¹. It is more common among the White population, and some reported incidences in the Asian population as well. It is relatively uncommon in African Americans.

Anti-GBM disease is the cause of 10 to 20% of RPGN. When associated with pulmonary hemorrhage in nearly 50% of patients it is called Goodpasture's syndrome. Genetically susceptible individuals develop anti-GBM IgG (rarely IgA and IgM) antibodies directed against specific epitopes located on the carboxy terminal or non-collagenous domain (NC1) of the alpha-3 chain of type IV collagen following a triggering event². The specific organ injury reflects the predominant tissue distribution of the alpha-3 chain in glomerular and alveolar basement membranes3. The fenestrated endothelium of glomerular capillaries allows auto-antibody binding and development of glomerulonephritis. Therefore, hallmark of this disease is continuous linear deposition of immunoglobulin, usually immunoglobulin G (IgG) along GBMs, demonstrated by DIF microscopy ⁷. Renal injury typically manifests as diffuse necrotizing and crescentic glomerulonephritis⁴⁶. Similar linear staining of tubular basement membrane (TBM) correlates with anti-tubular BM antibodies and tubulointerstitial disease⁸. Anti-GBM disease without major renal dysfunction is present in up to 30% of patients with anti-GBM auto-antibodies demonstrable in serum with conventional enzyme-linked immunosorbent assay (ELISA)9. The presence of anti-GBM antibodies is diagnostic, but the sensitivity is low. Thus, a negative auto-antibody test does not exclude anti-GBM antibody disease.

IgA nephropathy is the most common glomerulonephritis in adults¹⁰. The most typical presentation is macroscopic hematuria or rarely nephrotic syndrome shortly after a mucosal infection such as upper respiratory tract infection and bronchitis. Prognosis of the disease is highly variable with some patients showing a rapid progression causing of end stage renal disease (ESRD)¹¹. Factors including male gender, persistent microscopic hematuria, increased serum creatinine, proteinuria more than 1 g/d, and hypertension at presentation are segmental sclerosis, tubular atrophy, interstitial fibrosis, interstitial

cellular infiltrate, and peripheral capillary wall alterations such as deposits or endocapillary proliferation also indicate a poor prognosis¹².

IgA nephropathy can present with crescents in histology but simultaneous presentation of Anti-GBM disease with IgA nephropathy has been rarely described in the literature. In this study we are illustrating two cases of rare occurrence of anti-GBM glomerulonephritis with mesangial IgA deposition, suggesting IgA nephropathy. The association of these two pathogenetically unrelated glomerulonepritis may open up new windows and throw light in the pathogenesis and prognosis of both the diseases.

In this study we have also included the treatment history with a short followup. Hemodialysis and Plasmapheresis was also used when required in the patients of all the three groups. Response of the patient to all these as well as the survival was recorded and the prognosis were assessed in all the three.

AIMSAND OBJECTIVES:

1. To evaluate and compare the clinicopathological, histopathological and DIF microscopic features of cresentric IgA Nephropathy, Anti-GBM disease and combined IgA Nephropathy with Anti-GBM disease.

2. To evaluate and compare the renal outcome of cresentric IgA Nephropathy, Anti-GBM disease and combined IgA Nephropathy with Anti-GBM disease.

3. To compare the findings with the findings of the other researchers.

MATERIALAND METHODS:

Patient and Public Involvement: Patients attending the Nephrology department and undergoing renal biopsy are included in this study after taking proper informed consent. Study was conducted in the nephropathology unit of department of Pathology.

STUDY DESIGN: Prospective, cross-sectional, single centre study in a tertiary care hospital.

Information on patients' demographics (age and sex), course of disease and co morbidities were collected. Ultrasonography guided percutaneous needle biopsies with two core of renal biopsies were obtained from each patient. The mean number of glomeruli in all 17 specimens was 17.94 ± 7.2 , with a minimum no of 7 glomeruli and maximum 30 glomeruli.

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Tissue for light microscopy was fixed in neutral buffered formalin, embedded in paraffin, and sectioned at 2-3 μ m intervals. They were then stained with hematoxylin-eosin (HE), periodic acid-Schiff (PAS), Masson's trichrome, and silver methenamine stain. Frozen sections were examined for DIF after staining with fluorescein-conjugated (FITC) anti sera specific for human IgG, IgM, IgA, C3, C1q, kappa and lambda light chains (Dako, Denmark). They were then washed in phosphate-buffered saline, mounted in buffered glycerol, and examined in a fluorescent microscope. Fluorescence was graded as absent, trace, mild, moderate, bright or blazing intensity (0/0.5/1+/2+/3+), and the staining pattern was described as linear, granular, or lumpish deposition along glomerular capillary wall or in mesangium. The staining intensity was scored by two pathologists, and the average score was used.

Following laboratory tests were undertaken:

1. Urine protein quantification using a 24-hour collection.

2. Serum albumin, triglyceride, creatinine, complement 3 and 4 and effective glomerular filtration rate (eGFR).

3. AntiGBM antibody estimated by conventional ELISA method. Sera with low levels (<40%) of anti-GBM antibodies are further identified by Western-blot analysis using purified human α 3 (IV)NC1 as solid-phase ligands¹³.

4. Routine urine examination done for proteinuria, microscopic hematuria, pus cells and casts etc. The severity of proteinuria was graded 0 to 3+ with 0 representing the mildest and 3+ representing the most severe stage.

Outcome parameters. The following outcome and prognostic parameters were recorded and used for statistical analysis: development of ESRD, all-cause mortality, a permanent 50% reduction in eGFR compared with the baseline value/ no remission (NR), complete remission (CR), and partial remission (PR). CR was defined as urinary protein excretion <0.3 g/d (uPCR <300 mg/g or <30 mg/mmol) accompanied by a normal serum albumin concentration and a normal serum creatinine. PR was defined as urinary protein excretion <3.5 g/d (uPCR <3500mg/g or <350mg/mmol), a \geq 50% reduction from peak values, an improvement or normalization of the serum albumin concentration, and stable serum creatinine^{14,15}. ESRD was defined as a permanent drop in eGFR to 15mL/min/1.73m2 requiring dialysis or kidney transplantation. Poor renal outcome was indicated by NR, ESRD, hemodialysis dependence and death. Good renal outcome was indicated by PR or Cr¹⁶.

Ethical issues: This study is approved by the Institutional Ethics Committee and Research Advisory Committee of the Institute. All the patients gave informed consent before participating in this study.

STATISTICALANALYSIS:

Mean value with two standard divisions were calculated for the serological parameters. We performed the oneway ANOVA(and nonparametric) test for comparisons between three groups, p-Value <0.05 was considered as significant. Survival analysis was done by Kaplan Meyer curve and comparison of survival curves were done by Log-rank (Mantel-Cox) test. p-Value <0.05 was considered as significant. We have used statistical software (GraphPad PRISM 6) for analysis.

RESULTS:

We have included 9 cases of IgA Nephropathy with crescents, 6 cases of AntiGBM disease and 2 cases of combined IgA Nephropathy and AntiGBM disease [Figure 1]. All the cases presented clinically with RPGN and histopathologically with crescentric glomerulopathy. Average age at presentation in three groups are 32.78 ± 12.04 years, 45.00 ± 11.38 years and 38.50 ± 13.44 years respectively. All the three groups showed male prepondarence with male female ratio being 1.25:1, 2:1 and 2:0 resepectively. More than half of the patients of IgA Nephropathy(55.5%) and AntiGBM disease (66.6%) presented with edema whereas it was not seen in the patients of combined disease. Diabetes was rarely seen in three groups. History of infection and hypertension were predominantly seen in patients of IgA Nephropathy and combined disease. Association of hypertension was statistically significant. Most prominant pulmonary symptom was hemoptysis which was predominantly seen among the patients of AntiGBM disease (83.3%) and combined disease (50%) and the association was statistically significant [Table 1].



Figure 1: Distribution of the cases of Rapidly Progreesive Glomerulonephritis

In most patients serum albumin was reduced slightly and triglyceride was normal and none showed nephrotic syndrome. Anti GBM antibody was statistically significantly higher in AntiGBM disease (178.8 \pm 140.6IU/mL) and combined disease (295.0 \pm 190.9IU/mL). Both complement 3 and 4 levels were within normal limits in all the three groups. Among the renal function indicators serum creatinine was statistically significantly high in AntiGBM disease (10.8 \pm 4.5 mg/dL) and combined disease (8.3 \pm 1.7 mg/dL) than in IgA Nephropathy (4.6 \pm 3.9 mg/dL). Quantitative protein estimation show 24 hour proteinuria in the range of 2.3 \pm 0.84 gm, 2.4 \pm 0.57 gm and 3.2 \pm 0.56 gm respectively in three groups. Effective GFR was reduced in all the three groups. All the cases were ANCA (MPO ANCA and PR3 ANCA) negative. All the patients were tested for HIV serology along with HbsAg and HCV antibody which were negative [Table 1].

In routine urine examination red blood cell casts were statistically significantly associated with IgA Nephropathy (77.7%) rather than AntiGBM (0%) and combined (50%) disease. Other features like presence of RBC, pus cell and granular cast are comparable in all three groups. Urine protein excreation was 1+ to 2+ in dipstick examination in all the three groups [Table 1].

	CRESENTRIC	ANTI-GBM	IgA AND	р
	IgANEPHROPA	DISEASE	ANTIGBM	VALUE
	THY (n=9)	(n= 6)	COMBINAT	
			ION (n=2)	
	DEMO	GRAPHICS		
Age (Years)	32.78 ± 12.04	45.00 ± 11.38	$38.50{\pm}13.44$	0.1865
Men, n (%)	5 (55.5%)	4 (66.6%)	2 (100%)	0.5404
	COMORE	BIDITIES, n (%)	
Edema	5 (55.5%)	4 (66.6%)	0 (0%)	0.2939
Hypertention	9 (100%)	2 (33.3%)	2 (100%)	0.0030
Diabetes	1 (11.1%)	1 (16.6%)	0 (0%)	0.8432
Infection	7 (77.7%)	1 (16.6%)	1 (50%)	0.0687
Pulmpnary	0 (0%)	5 (83.3%)	1 (50%)	0.0006
disease				
	SEF	ROLOGY		
Serum	2.6 ± 0.53	2.3±0.57	3.2 ± 0.56	0.2102
Albumin				
(g/dL)				
Serum	158.4 ± 40.9	166.0 ± 54.1	$217.0{\pm}~42.4$	0.2971
Triglyceride				
(mg/dL)				
AntiGBM	2.4 ± 2.5	178.8 ± 140.6	295.0 ± 190.9	0.0018
antibody				
(IU/mL)	111.1.05.5	100.0.10.7	1.50.5.05.0	0.1.111
Complement	111.4 ± 35.5	120.8 ± 13.7	159.5 ± 26.2	0.1411
3 (g/dL)				
Complement	26.67 ± 6.4	25.50 ± 7.0	33.00 ± 5.6	0.3916
4 (g/dL)	DENIAL FUDIO		TODO	
	RENAL FUNC	TION INDIC.	ATORS	
Serum	4.6± 3.9	10.8 ± 4.5	8.3 ± 1.7	< 0.0001
Creatinine				
(mg/dL)				

TABLE 1: Clinicopathological parameters and prognosis of three groups of patients

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24 hour	2.3 ± 0.84	2.4 ± 0.57	3.2 ± 0.56	0.3490
Proteinuria				
(gm)				
eGFR	21.80 ± 14.5	18.63 ± 24.9	38.03 ± 14.4	0.4657
(mL/min/				
1.73 m^2)				
	URINE ROUTI	NE EXAMIN	ATION	
RBC (n/hpf)	16 ± 5.6	23 ± 28.1	7 ± 2.8	0.5126
Pus cell	13.5 ± 7.3	5.8 ± 2.3	10.5 ± 6.4	0.0819
(n/hpf)				
Red cell cast	7 (77.7%)	0 (0%)	1 (50%)	0.0063
n, (%)				
Granular	3 (33.3%)	0 (0%)	0 (0%)	0.2278
cast n, (%(
Urine protein	2.1 ± 0.6	1.6 ± 0.8	1.5 ± 0.7	0.3645
CR n,%	3, 33.3%	1, 16.6%	0	0.5561
PR n, %	4, 44.4%	3, 50%	1, 50%	
NR n, %	2, 22.2%	2, 33.3%	1, 50%	
HD	4, 44.4%	2, 33.3%	1, 50%	0.7842
dependence				
n, %				
Death n, %	2, 22.2%	3, 50%	1, 50%	0.2558

Number of glomeruli detected in the kidney biopsy from all the three groups was comparable with the range of 7 to 30 glomeruli. Global sclerosis detected in all the three groups indicating chronicity. Segmental sclerosis seen in IgA Nephropathy [Figure 2] and combined disease [Figure 3] but not in AntiGBM disease [Figure 4]. Cresents are present in significantly higher number in AntiGBM (16.83±7.6) and combined (16.00 ± 8.5) disease than in IgA Nephropathy (7.55 ± 3.2). Incidence of cellular and fibrocellular cresents are comparable in three groups whereas fibrous cresents are signicantly common in combined disease. Mesangial hypercellularity and matrix expansion were significantly more in IgA Nephropathy and combined disease. Endocapillary hypercellularity was present in IgA Nephropathy only whereas fragmentation of GBM was present in AntiGBM and combined diseases and this difference was statistically significant. Intraglomerular necrosis seen in IgA Nephropathy and AntiGBM disease. In the tubulointerstitial compartment proximal tubular epithelial injury in the form of vacuolar degenation, loss of brush border etc, tubular atrophy, presence of intratubular cast like granular cast, colloid cast and red cell cast, interstitial mononuclear inflammatory cell infiltration focally or diffusely and interstitial fibrosis are comparable in all the three groups. Different type of vascular changes identified. Commonest were hypertrophy of tunica media and arteriosclerosis. Others are mucoid intimal changes and leukocytoclastic vasculitis was comparable in all the three groups [Table 2].

In DIF microscopy IgG and kappa showed statistically significant expression along the GBM in linear fashion in Anti GBM and combined disease. Whereas IgA show significant mesangial positivity in IgA Nephropathy and combined disease. Lambda light chain and C3c showed comparable expression in all the three groups. IgM was segmentally positive in IgA Nephropathy whereas C1q was absolutely negative in all the three groups [Table 2].

TABLE 2: Histopathological and DIF microscopic finding in three groups

	CRESENT RIC IgA	ANTI-GBM DISEASE	IgA AND ANTIGBM	P Value
	NEPHROP	(n=6)	COMBINATI	
HISTO	PATHOLOGI	CAL CHARA	CYERISTICS	
Number of Glomeruli	16.22±7.4	19.50± 7.9	21.00± 4.2	0.5916
Global Sclerosis	3.78 ± 4.4	1.50 ± 2.5	1.00 ± 1.4	0.4180
Segmental sclerosis	0.89±1.2	0	1.00±1.4	0.2091
Total no of Crescents	7.55± 3.2	16.83±7.6	16.00± 8.5	0.0166
Cellular Crescents	5.22± 3.7	15.50± 9.3	11.00±15.5	0.0611
Fibrocellular Crescents	3.33± 4.9	1.33±2.4	3.00± 4.2	0.6573

Fibrous 0.11 ± 0.3 0 2.00 ± 2.8 0.0201 Crescents 0.0063 Mesangial cell 0.78 ± 0.4 0 0.50 ± 0.7 proliferation (present-1, absent-0) Mesangialmatrix 1.00 ± 0.0 0.17 ± 0.4 1.00 ± 0.0 < 0.0001 expansion (present-1 absent-0)) Endocapillary 0.33 ± 0.5 0 0 0.2278 hypercellularity (present-1, absent-0) Fragmentation 0 0.83 ± 0.4 1.00 ± 0.0 < 0.0001 of GBM (present-1, absent-0) Necrosis 0.44 ± 0.5 0.33 ± 0.5 0 0.5404 (present-1, absent-0) Tubular injury 0.50 ± 0.7 0.67 ± 0.5 0.33 ± 0.5 0.4976 (present-1, absent-0) Intratubuler cast 0.78 ± 0.4 0.50 ± 0.5 1.00 ± 0.0 0.3645 (present-1, absent-0) Tubular atrophy 18.89 ± 11.6 15.00 ± 13.4 30.00 ± 14.1 0.3457 (%) Interstitial 0.22 ± 0.4 0.50 ± 0.5 0.50 ± 0.7 0.5404 inflammation (diffuse-1,focal-0) Interstitial 18.89 ± 11.6 15.00 ± 13.4 30.00 ± 14.1 0.3457 fibrosis (%) IFTA (%) 18.89 ± 11.6 15.00 ± 13.4 30.00 ± 14.1 0.3457 Vascularchanges 0.55 ± 0.5 0.17 ± 0.4 1.00 ± 0.0 0.1017 (present-1, absent-0) 0.11 ± 0.3 0 0.50 ± 0.7 0.1871 Arteriosclerosis (present-1, absent-0) DIF MICROSCOPY (INTENSITY SCORE) 0.28 ± 0.26 2.83 ± 0.41 2.50 ± 0.71 < 0.0001 IgG 2.44 ± 0.53 2.00 ± 0.0 < 0.0001 0 IgA IgM 0.22 ± 0.26 0 0 0.1068 1.00 ± 0.43 0.4538 C3c 0.67 ± 0.82 0.50 ± 0.71

All patients were treated and followed up with a range of 8 months to 3 months. CR was seen in 33.3% and 16.6% of IgA Nephropathy and Anti GBM disease respectively but not in combined disease. On the other hand 22.2%, 33.3% and 50% of IgA Nephropathy, Anti GBM disease and combined disease respectively were totally nonresponsive to drugs and treated with hemodialysis among which 44.4% of IgA Nephropathy, 33.3% of Anti GBM disease and 50% of combined disease become dialysis dependent. 22.2% of IgA Nephropathy and 50% each of Anti GBM and combined disease died during follow up [Table 1,3,4 &5]. Kaplan Meyer survival analysis curve for all cause mortality versus dialysis dependence [Figure 2] and CR,PR versus NR [Figure 3] show no difference of survival between three groups as the comparisons are statistically insignificant in Log-rank test.

0

 2.50 ± 0.84

 2.17 ± 0.75

0

 2.50 ± 0.71

 1.50 ± 0.71

< 0.0001

0.0985

Different microscopic features of IgA Nephropathy [Figure 4], Anti GBM disease [Figure 5] and combined disease [Figure6] respectively.

TABLE 3	: Followup	of IgA Ne	phropa	thy case
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0

 0.55 ± 0.46

 1.11 ± 0.93

C1q

Kappa

Lambda

	DURATION	DEATH				
	(Months)	RESPONSE	DEPENDANCE			
1	8	PR	Y	N		
2	6	PR	N	Y		
3	5	CR	N	N		
4	6	NR	Y	N		
5	7	CR	Ν	N		
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Volume - 11 | Issue - 07 | July - 2021 | PRINT ISSN No. 2249 - 555X | DOI : 10.36106/ijar

6	5	PR	Ν	Y
7	5	NR	Y	Ν
8	7	CR	Ν	N
0	6	PB	V	N

TABLE 4: Followup of Anti-GBM disease

ANTI- GBM	DURATIO	TYPE OF	DIALYSIS	DEATH
DISEASE	N (Months)	RESPONSE	DEPENDANCE	
1	5	NR	N	Y
2	7	PR	Y	N
3	3	NR	N	Y
4	6	PR	N	Y
5	7	CR	Ν	N
6	3	PR	Y	N

TABLE 5: Followup of Combined IgA Nephropathy and Anti-GBM disease

IgA	DURATI	TYPE OF	DIALYSIS	DEATH
NEPHROPATHY	ON	RESPONSE	DEPENDA	
AND ANTI- GBM	(Months)		NCE	
DISEASE				
1	5	NR	N	Y
2	7	PR	Y	N

Survival proportions: All cause mortality vs Dialysis dependence



Figure 2: Percent survival of All cause mortality versus Dialysis dependance

Survival proportions: No remission versus partial & complete remission







Figure 4: IgA Nephropathy with Crescent formation **4A:** Marked mesangial hypercellularity in IgA Nephropathy (H&E X 400)

4B: Fibrocellular crescent in IgA Nephropathy (MT X 100) **4C:** Interstitial fibrosis and tubular atrophy in IgA Nephropathy (H&E

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X 40) Inset: Crescent in in IgA Nephropathy (JMS X 100) 4D: Mesangial granular IgA deposit in DIF (X200)



Figure 5: Anti-GBM Disease

5A: Cresents present in most glomeruli in Anti-GBM Disease (H&E X 100)

5B: Cellular crescent in Anti-GBM Disease (H&E X 400)

5C: Cellular crescent in Anti-GBM Disease (JMS X 400)5D: Linear positivity of IgG along glomerular basement membrane (DIF X 200)



Figure 6: Combined IgA Nephropathy and Anti-GBM Disease
6A: Fibrocellular cresents in the glomeruli (H&E X 100)
6B: Fibrocellular cresents in the glomeruli (JMS X 100)
6C: Mesangial granular IgA deposits (DIF X 100)
6D: Linear positivity of IgG along glomerular basement membrane (DIF X 400)

DISCUSSION:

RPGN is suspected when there is a rapid decline in renal function (reduction of more than 50% of GFR within 3 months) along with nephritic urinary sediment and a normal or enlarged kidney size¹⁷. It is usually caused by one of the three following mechanisms: anti-GBM antibody disease with or without pulmonary hemorrhage, pauciimmune glomerulonephritis, and severe immune complex mediated glomerulonephritis¹⁸. Untreated patients with RPGN may rapidly progress to ESRD.

The main pathologic finding is extensive glomerular crescent formation. The percentage of glomeruli that exhibit crescents usually correlates with the severity of disease¹⁸. The ubiquitous pathological feature of crescentic glomerulonephritis is a focal rupture of glomerular capillary walls that can be seen by light microscopy and electron microscopy.

In our study we have compared the clinical and serological parameters, renal function indicators, routine urine examination, renal biopsy findings by light and DIF microscopy in three different groups of patients presenting with RPGN- one group with IgA Nephropathy, one group with Anti-GBM disease and another with combined IgA Nephropathy with Anti-GBM disease. Small number of clinicopathological features show statistically significant difference between the three groups like incidence of hypertension, hemoptysis, serum creatinine, anti GBM antibody, red cell casts in urine, total number of crescents including fibrous crescents and mesangial hypercellularity with matrix expansion, fragmentation of GBM etc. In DIF microscopy IgG, IgA and kappa light chain positivity show statistically significant difference between the three groups. After intense search of literature we have found that small number of previous studies have done similar type of comparison. Cui et al done comparison of clinical and laboratory data between the patients of Anti-GBM disease associated with immune complex deposition (10 cases) and and pure Anti-GBM disease (37 cases). Similar to our finding most of the features were comparable between the two groups without any significant difference¹⁹. Oliguria was less common in concurrent IgA Nephropathy and AntiGBM disease (10%) than Anti GBM disease with deposition of other immune complexes (40%). In our study both the patient presented with oliguria with eGFR in the range of 38.03± 14.4 mL/min/ 1.73 m². Percentage of crescent formation was less in concurrent IgA Nephropathy and AntiGBM disease (59%) than that in antiGBM disese with deposition of other immune complexes (93.8%). In our study incidence of crescents was 72% in concurrent IgA Nephropathy and AntiGBM disease.

All the diagnosed cases were treated and followed up. There was no significant difference between the response rate and poor renal outcome indicated by non response, ESRD, dialysis dependence and incidence of death within the three groups. Other researchers Cui et al found that concurrent IgA Nephropathy and Anti-GBM disease show better prognosis than AntiGBM disease alone¹⁹, whereas Gupta et al found just the opposite²⁰. Anti-GBM disease cases less commonly (10%) achieved dialysis indepence than concurrent IgA Nephropathy and AntiGBM disease (60%). Similarly other researcher showed that that Anti-GBM disease wass the most aggressive form of crescentic glomerulonephritis, with the highest frequency of renal insufficiency followed by ANCA-associated vasculitis and then by immune complex-mediated crescentic glomerulonephritis²¹. Fifty percent of our patients with combined disease were dialysis dependent.

Prognosis depends not only on the number of crescents but also on the tubulointerstitial chronicity indicated by interstitial fibrosis and tubular atrophy (IFTA), number of sclerosed glomeruli and the presence of arteriosclerosis. When chronic changes are predominant in renal biopsy, intense immunosuppression protocol was not provided weighing the potential side effects of intense immunosuppression versus the low possibility of salvaging the renal function.

Anti-GBM disease occurs in all age group with a peak in third decade in male and sixth decade in both male and female²². Similarly our study show mean age group of fifth decade with slight male prepondarence. The diagnosis of Anti-GBM disease is made by demonstration of linear IgG positivity along the GBM in renal biopsy by DIF microscopy and is usually supported by the serological detection of circulating anti-GBM antibodies and presence of crescents in light microscopy of renal biopsy²³. The sensitivity of ELISA in detecting anti-GBM antibodies is 87–90%, when combined with Western blot it increases to 95–99%. Early intervention of anti-GBM disease is crucial to preserve renal function and avoid severe pulmonary sequelae. Careful examination of renal biopsy along with clinical evaluation and serological evaluation of of anti-GBM antibody along with close follow-up are necessary in managing these patients.

Main aim of our study is to describe two unusual occurances of concurrent IgA Nephropathy and Anti-GBM disease. First report of anti-GBM disease and mesangial IgA deposits was described in 1998 in a 12 year postrenal transplant recipient from Canada²⁴. After that several studies show similar type of dignosis with varied clinicopathological setup from different countries²⁵⁻²⁹ [Table 6]. IgA Nephropathy may precede or follow Anti-GBM disease. When Anti-GBM disease superimpose on IgA Nephropathy possible mechanism may be IgA-related immune complex exposing the non-collagenous domain of the alpha-3 subunit, that are normally hidden by intrachain methionine cross-links^{19,30,31}. Experimental models and in vivo studied on IgA nephropathy also prove the release of IL-1, TNF, IL-6, oxygen radicals, and eicosanoids by infiltrating glomerular and interstitial monocytes/macrophages and mesangial cells^{23,32}. Inflammatory mediators might initiate immunologic events causing conformational changes of GBM and exposure of antigens, thus facilitating anti-GBM antibody production. Altered IgA deposition in the mesangium characterised by defect in galactosylation of IgA1 occurs in IgA nephropathy33. Aberrant polysaccharide chain of IgA1 triggers anti-GBM IgG2 antibody production which gets deposited in the GBM³⁴. Alternative hypothesis is given by Wang et al, highlighting the

possibility of increased antigen synthesis or capping and shedding of antigen antibody complexes³⁵.

TABLE 6: Comparison of finding of Combined IgA Nephropath	y
and Anti-GBM disease of our study with other studies	

anu Anu-v	3DIVI UIS	ease of ou	li study v	vitii otiiei	studies	
	Yamagu	Troxell	Ge et	Xu et	Suh K-S	Our study
	chi	et al ¹⁰	al	al ¹²	et al' ((2018-
	et al ⁹	(2016)	(2015)	(2016)	2019)	2019)
	(2013)					
Country	Japan	USA	China	China	Korea	India
Number	1	1	1	1	1	2
Age	46	60	24	50	38	38.50± 13.44
Sex	М	F	М	F	F	М
Symptom	Azotemi	Fever,	Edema,	Gross	Fever	Hematuria
s	a, fever	dysphag	heahach	hematuri		& oliguria
		ia, wt	е,	а		
		loss	nocturia			
Hyperten	134/83	Negativ	Positive	Negative	Negative	Positive in
sion	mm Hg	e	194/120			both
	controlle					(162/100 &
	d					170/116)
Diabetes	Positive	Negative	Negative	Negative	Negative	Negative
Infection	Negative	Negative	Negative	Negative	Negative	Positive in 1
Hemonty	Negative	Negative	Negative	Negative	Negative	Positive in 1
sis	ivegative	regative	inegative	inegative	riegative	
Oliguria		Negative	Negative	Negative	Negative	Positive
Nephrotic	Negative	Negative	Positive	Negative	Negative	Negative
syndrome						5.18
Hematuria	Positive	Negative	Negative	Positive	Positive	Positive
24 hour	2.9	0.4	7.04	0.41	14	32+0.56
urine	2.9	0.4	7.04	0.41	1.7	5.2± 0.50
nrotein						
(gm) or in						
spot urine						
Creatinine	5.2	0.73	37	26	5.4	8 3+ 1 7
(mg/dI)	5.2	0.75	5.7	2.0	5.4	0.5± 1.7
(IIIg/uL)	214	120	Dositiva	258	187.2	205.0+
GRMAL	EU/mI	150%	not	EU/mI	II/mI	100.0
ODM AU	LU/IIIL	15070	described	LU/IIIL	U/IIIL	190.9 III/mI
Clamaamali	16	12	21	10	16	21.00 ± 4.2
giomerun	10	12	51	18	10	21.00 ± 4.2
,11 Global	1	2	2	0	5	1.00 ± 1.4
Global	1	5	2	0	5	1.00 ± 1.4
Creasents	15	1 (90/)	10	16	11 (620/)	22 (760/)
crescents	(0.49/)	1 (8%)	10	(800/)	11 (05%)	52 (70%)
Tuno	Collular	Callular	Collular	Collular	Callular	Callular
Type	(100%)	(100%)	(100%)	(750/)	(45, 50/)	(68, 70/)
	(10070)	(10070)	(10070)	(7570),	(43.370), fibrocell	fibrocellular
				ular	ular	(18,7%)
				(25%)	(545%)	fibrous(12)
				(2370)	(34.370)	5%)
Tuaatuaan	IV.	IV.	IV	IV	IV.	IV Matul
Treatmen	IV Matal	IV Matal	IV Matal	IV Matal	IV Matal	DD Oral
ι	DD+Oro	DD+	DD+	DD+Oral	DD+	PD+Orai-
		 Oral	r D+ Oral	PD+OIal		гD
	1-1 D		PD+IV	conheno	+IV	
		Methotr	cyclophs	late	cyclophs	
		exate	nhamide	mofetil	nhamide	
Plasmanh	Dona	Not	Not	Not	Not	Done in 1
riasinapii	failed	norform	norform	norform	nerforme	Done III I
010515	due to	ad	ad	ad	d	
	allerow	eu	eu	eu	u	
Remissio	NP	CP	NP	CP	CP	PR in 1
n type	INIK	UK	INK	CK	CK	IR III I,
Homodi-1	Dialari-	Dialarai-	Dialari-	Dialari-	Dialari-	Dialvaia
riemodial	depende	indeper	depende	indepar	independ	dependent
y 515	nt	dent	nt	dent	ent	in 1
Death	No	No	No	No	No	Death in 1
Deatin	110		110	110		Death III I

When the AntiGBM disease preceds IgA Nephropathy, possible mechanism may be alteration of GBM permeability followed by the circulating immune complex deposition in the mesangium¹⁹. Mesangial IgA deposits are known to occur in association with bacterial and viral infection like HIV^{36,37}. One of our patient also gave

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Volume - 11 | Issue - 07 | July - 2021 | PRINT ISSN No. 2249 - 555X | DOI : 10.36106/ijar

history of upper respiratory tract infection.

CONCLUSION:

In our study we have analysed three relatively uncommon type of RPGN and found that crescentic IgA nephropathy, anti GBM disease and the combined one though have relative difference in presentation have similar therapeutic response and renal outcome. Proper and early clinicopathological diagnosis of these uncommon types of crescentric glomerulonephiritis is important since all are of poor renal outcome

LIMITATIONS:

More cases with an in-depth examination of underlying pathogenic relationships between anti-GBM disease and IgA nephropathy are needed to be analyzed.

Funding: Funding is provided by our Institution.

Ethics: This study is approved by the Institutional ethical committee. All the participants gave informed consent in writting at the begining of the study.

Provenance and peer review Not commissioned; externally peer reviewed

Data availability statement All data relevant to the study are included in the Article.

Data sharing statement: No additional data available.

Conflicts of interest: we don't have any conflicting interest.

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REFERENCES:

- Berti A, Cornec-Le Gall E, Cornec D, Casal Moura M, Matteson EL, Crowson CS, 1. Ravindran A, Sethi S, Fervenza FC, Specks U. Incidence, prevalence, mortality and chronic renal damage of anti-neutrophil cytoplasmic antibody-associated glomerulonephritis in a 20-year population-based cohort. Nephrol. Dial. Transplant. 2019 Sep 01;34(9):1508-1517.
- 2 Austin HA 3d, Muenz LR, Joyce KM, Antonovych TT, Balow JE. Diffuse proliferative lupus nephritis: identification of specific pathologic features affecting renal outcome. Kidney Int 1984;25(4):689-95. Sasatomi Y, Kiyoshi Y, Takabeyashi S. A clinical and pathological study on the
- 3. characteristics and factors influencing the prognosis of crescentic glomerulonephritis using a cluster analysis. Pathol Int 1999; 49(9):781-5.
- using a cluster analysis. Pathol Int 1999; 49(9):781-5. Jennette JC, Nickeleit V. Anti-glomerular basement membrane glomerulonephritis and Goodpasture syndrome. In:Jennette JC, Silva FG, Olson JL et al. (eds). Heptinstall's Pathology of the Kidney. 7th edn. Philadelphia, PA: Wolters Kluwer, 2015, pp. 657–684 Cui Z, Zhao M-H. Advances in human antiglomerular basement membrane disese. Nat Rev Nephrol 2011; 7: 697–705 Fischer EG, Lager DJ. Anti-glomerular basement membrane glomerulonephritis: a morphologic study of 80 cases. Am J Clin Pathol 2006; 125: 445–450 4. 5.
- 6.
- 7.
- Lahmer T, Heemann U. Anti-glomerular basement membrane antibody disease: a rare autoimmune disorder affecting the kidney and the lung. Autoimmun Rev 2012; 12: 169-173
- 8. Andres G, Brentjens J, Kohli R, et al. Histology of human tubulo-interstitial nephritis associated with antibodies to renal basement membranes. Kidney Int 1978;13(6):480-
- Kalluri R, Sun MJ, Hudson BG, Neilson EG. The Goodpasture autoantigen. Structural delineation of two immunologically privileged epitopes on alpha3 (IV) chain of type IV collagen. J Biol Chem 1996; 271(15):9062-8. 9.
- 10 Li LS, Liu ZH. Epidemiologic data of renal diseases from a single unit in China: Analysis based on 13,519 renal biopsies. Kidney Int. 2004;66:920–923. Berthoux FC, Mohey H, Afiani A. Natural history of primary IgA nephropathy. Semin 11.
- Nephrol. 2008;28:4-9.
- Fogo AB, Kashgarian M. Diagnostic atlas of renal pathology. 1st ed. Spain: Elsevier-12. Saunders; 2005. pp. 107–120. Kamimura H, Honda K, Nitta K, et al. Glomerular expression of alpha2(IV) and 13.
- alpha5(IV) chains of type IV collagen in patients with IgA nephropathy. Nephron 2002; 91:43-50
- 14 KDIGO clinical practice guideline for glomerulonephritis. Abstract. Kidney Int Suppl (2011). 2, 142 (2012). Qin, H. Z. et al. Combined Assessment of Phospholipase A2 Receptor Autoantibodies 15
- and Glomerular Deposits in Membranous Nephropathy. *Journal of the American Society* of Nephrology: JASN. 27, 3195–3203 (2016).
- Chen X, Chen Y, Shi K, Lv Y, Tong H, Zhao G, Chen C, Chen B, Li D, Lu Z. Comparison of prognostic, clinical, and renal histopathological characteristics of overlapping 16 idiopathic membranous nephropathy and IgA nephropathy versus idiopathic membranous nephropathy. Sci Rep. 2017 Sep 13;7(1):11468, doi:10.1038/s41598-017-11838-1. PMID: 28904360; PMCID: PMC5597578. Anders HJ. Diagnosis and Management of Crescentic Glomerulonephritis: State of the
- 17. Art. Saudi J Kidney Dis Transpl 2000;11:353-61 Couser WG. Rapidly progressive glomerulonephritis: classification, pathogenetic
- 18 mechanisms, and therapy. Am J Kidney Dis 1988;11(6):449-64. Cui Z, Zhao MH, Wang SX, Liu G, Zou WZ, Wang HY. Concurrent antiglomerular
- 19 basement membrane disease and immune complex glomerulonephritis. Ren Fail 2006; 28:7-14.
- Gupta Y, Swain M, Gowrishankar S.Antiglomerular basement membrane disease 20.
- 21
- Hudson BG, Tryggvason K, Sundaramoorthy M, Neilson EG. Mechanisms of disease Alport's syndrome, Goodpasture's syndrome, and type IV collagen. N Engl J Med 2003;348:2543-56
- Ikezumi Y, Hurst LA, Masaki T, et al. Adoptive transfer studies demonstrate that 23

INDIAN JOURNAL OF APPLIED RESEARCH 12

- macrophages can induce proteinuria and mesangial cell proliferation. Kidney Int 2003; 63: 83-95 24.
- Trpkov K, Abdulkareem F, Jim K, Solez K. Recurrence of anti-GBM antibody disease twelve years after transplantation associated with de novo IgA nephropathy. Clin Nephrol 1998; 49: 124-8. Yamaguchi H, Takizawa H, Ogawa Y, Takada T, Yamaji I, Ura N. A case report of the 25.
- anti-glomerular basement membrane glomerulonephritis with mesangial IgA deposition. CEN Case Rep 2013; 2: 6-10.
- Troxell ML, Houghton DC. Atypical anti-glomerular basement membrane disease. Clin Kidney J 2016; 9: 211-21. 26.
- Ge YT, Liao JL, Liang W, Xiong ZY. Anti-glomerular basement membrane disease combined with IgA nephropathy complicated with reversible posterior leukoencephalopathy syndrome: an unusual case. Am J Case Rep 2015; 16: 849-53
- Xu D, Wu J, Wu J, et al. Novel therapy for anti-glomerular basement membrane disease 28.
- Xu D, Wu Y, Wu Y, Wu Y, Yu Y, Yu Y, Wu Y, Yu 29 402. doi: 10.4132/jptm.2019.08.05. Epub 2019 Sep 16. PMID: 31525832; PMCID: PMC6877440
- Dammacco F, Battaglia S, Gesualdo L et al. Goodpasture's disease: a report of ten cases and a review of the literature. Autoimmun Rev 2013; 12: 1101–1108 30
- Yang R, Hellmark T, Zhao J et al. Levels of epitope-specific autoantibodies correlate with renal damage in anti-GBM disease. Nephrol Dial Transplant 2009; 24: 1838–1844 31.
- Kamimura H, Honda K, Nitta K, et al. Glomerular expression of alpha2(IV) and alpha5(IV) chains of type IV collagen in patients with IgA nephropathy. Nephron 2002; 32 91.43-50
- Donadio JV, Grande JP. IgA nephropathy. N Engl J Med 2002; 347: 738–748 Tomana M, Novak J, Julian B, et al. Circulating immune complexes in IgA nephropathy consist of IgA1 with galactose-deficient hinge region and antiglycan antibodies. J Clin 34
- Consist of rgA1 win galactocencreat imge region and analyzed antibodies. J Chin Invest 1999; 104: 73–81 Wang A, Wang Y, Wang G, Zhou Z, Xun Z, Tan X. Mesangial IgA deposits indicate pathogenesis of anti-glomerular basement membrane disease. Mol Med Rep 2012;5:1212-4. 35.
- Katz A et al. (1992) IgA nephritis in HIV-positive patients: a new HIV-36 37
- associated nephropathy? *Clin Nephrol* **38**: 61–68 Kenouch S and Mery JP (1992) AIDS and IgA nephropathy. *Nephron* **61**: 473 38.