Microbiology

"COMPARATIVE STUDY OF AEROBIC BACTERIAL ISOLATES IN SUPERFICIAL WOUND SWAB AND DEEP TISSUE BIOPSY CULTURES AMONG TYPE 2 DIABETES MELLITUS PATIENTS WITH FOOT ULCERS"

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ABSTRACT INTRODUCTION: Diabetes is a metabolic disorder of the endocrine system and a worldwide problem.

Diabetic foot lesions are major medical, social and economic problem and are the leading cause of hospitalization for patients with diabetes. 85% major leg amputation begins with a foot ulcer. Targeted treatment of infective organism requires accurate identification of pathogens to enable refinement of antibiotic protocols to improve the outcome and to reduce antibiotic resistance

AIMS AND OBJECTIVES OF THE STUDY: 1. To isolate, identify and compare the aerobic bacteria in superficial wound swabs and deep tissue biopsies.

2. To study the antibiotic susceptibility pattern of the isolated bacteria.

MATERIALS AND METHODS: Two hundred patients with diabetic ulcers were included, ulcer swabs and tissue specimens collected, immediately transported and organisms were identified by culture and biochemical reactions. Antibiotic susceptibility test was done by using Kirby Bauer's disc diffusion method on Mueller Hinton agar under CLSI guidelines.

RESULTS AND CONCLUSION: Out of 200 samples, aerobic bacteria isolated were 77% from swab and 76% from tissue biopsy. A total of 220 isolates were obtained from 154 positive swabs and 207 isolates were obtained from 152 positive tissue biopsy specimens. 61% from swab and 65.8% from tissue biopsy yielded monomicrobial growth. The organisms isolated predominantly were Staphylococcus aureus (including MRSA), Klebsiella spp, Escherichia coli, Pseudomonas aeruginosa, Acinetobacter spp, Proteus spp and Enterococcus spp.

Swab culture yielded more organisms than tissue biopsy culture. Calculated P value was insignificant (0.115, P value<0.05 significant). Gram positive organisms were more sensitive to linezolid, vancomycin, amikacin and cotrimoxazole. Gram negative organisms were more sensitive to imipenem, piperacillin-tazobactam, amikacin and aztreonam. Swab is preferable because it is more economical, technique is easy and more compliable for the patient.

KEYWORDS: Diabetic ulcer, Aerobic bacteria, Swab method, Tissue biopsy

INTRODUCTION

Diabetes is a metabolic disorder of the endocrine system and a worldwide problem.1 India has become the diabetic capital of the world; within next few years with its attendant complications it is going to burden the resources of the country.

Global prevalence of diabetes mellitus was 3%.²Global prevalence of diabetes in 2003 was 194 million. By 2030, predicted to rise to 366 million due to longer life expectancy and changing dietary habits. Prevalence in southern India for diabetic ulcer ranges from 13 to 18%. Diabetic foot lesions are leading cause of hospitalization.⁴ 85% major leg amputation begins with a foot ulcer.5

The most common pathogenic organisms in diabetic ulcers are colonizers which include Coagulase Negative Staphylococcus, Enterococcus, Alpha hemolytic Streptococcus, Diphtheroids, Beta hemolytic Streptococcus, Pseudomonas aeruginosa, Staphylococcus aureus, Prevotella, Peptostreptococcus.6 Targeted treatment of infective organisms require accurate identification of pathogens in order to enable refinement of antibiotic protocols to improve the outcome and reduce antibiotic resistance.

Swab culture method is universal, quick and easy but susceptible to collecting contaminants including high number of colonizers and often lack the true pathogens.8-1

Hence the present study has been undertaken to evaluate the isolation of aerobic bacteria and their antibiotic susceptibility pattern and also for comparison of isolates between superficial swab and deep tissue biopsy.

AIMS AND OBJECTIVES OF THE STUDY

- To isolate, identify and compare the aerobic bacteria in superficial 1. wound swabs and deep tissue biopsies.
- To study the antibiotic susceptibility pattern of the isolated 2. bacteria.

MATERIALS AND METHODS

The present cross sectional comparative study was conducted at a tertiary care centre from November 2015 to April 2017 after obtaining informed consent. Institutional ethical clearance was obtained.

A total of 200 Type 2 diabetic patients with diabetic ulcer from the surgery department (inpatient and out patients) were included. Ulcer

swab and tissue biopsy samples were collected, transported to the microbiology laboratory and processed.

INCLUSION CRITERIA:

Type 2 diabetes mellitus patients with Wagner's grade 1, 2 foot ulcers.

EXCLUSION CRITERIA:

- Type 1 diabetes patients presenting with foot ulcers.
- Patient already underwent surgical debridement.
- Patient with Wagner's grade 0,3,4,5 diabetic ulcer

WAGNER ULCER CLASSIFICATION SYSTEM¹³ Grade Description

- 0 Skin intact 1
 - Superficial ulcer
- 2 Deeper, full-thickness extension of ulcer
- 3 Deep abscess or osteomyelitis associated with ulcer
- 4 Partial forefoot gangrene with ulcer
- 5 Extensive foot gangrene with ulcer

METHODS OF COLLECTION OF SPECIMENS Swab collection

Two swabs were collected under aseptic precautions, one for Gram's staining and another for culture. After thoroughly cleaning the wound with normal saline, cotton tipped swab was rubbed over the wound surface 1 cm² or collected directly from the base of the ulcer with sterile swab which is in contact with wound for at least 5sec.

Tissue biopsy collection

Deep tissue sample was surgically excised, approximately 1 to 5grams by using a new set of sterile instruments: curette, forceps and scalpel, after surrounding area of the wound was cleaned with povidone iodine solution at depth of 5mm from ulcer base.

Specimen transport - Both swab and tissue specimen were transported and processed within 15 minutes.1

The samples were processed using standard protocols. Identification of aerobic bacteria was based on morphology, staining characteristics, cultural characteristics and biochemical reactions. Antimicrobial susceptibility testing of the obtained isolates was done on Mueller Hinton agar using Kirby Bauer's disc diffusion method under CLSI guidelines (2015).

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RESULTS

A total of 200 patients were enrolled for the study. A total of 220 isolates were obtained from 154 positive swabs and 207 isolates were obtained from 152 positive tissue biopsy specimens.

The study group comprised predominantly males 123(61.5%) and 77(38.5%) were females.

Table 1.age Distribution

| AGE IN YEARS | FREQUENCY | PERCENTAGE (%) |
|--------------|-----------|----------------|
| 30 - 40 | 13 | 6.5 |
| 40 - 50 | 38 | 19.0 |
| 50 - 60 | 69 | 34.5 |
| 60 - 70 | 49 | 24.5 |
| >70 | 31 | 15.5 |
| Total | 200 | 100.0 |

Table-2 Culture Outcome

| | Swab | Tissue biopsy |
|-----------|----------|---------------|
| Growth | 154(77%) | 152(76%) |
| No Growth | 46(23%) | 48(24%) |
| Total | 200 | 200 |

Table -3 Microbial Growths Obtained In Swab And Tissue Biopsy

| | SWAB | % | TISSUE | % |
|---------------|------|------|--------|------|
| Monomicrobial | 94 | 61 | 100 | 65.8 |
| Dimicrobial | 54 | 35.1 | 47 | 31 |
| Polymicrobial | 6 | 3.9 | 5 | 3.2 |
| Total | 154 | 100 | 152 | 100 |

Table-4 Organisms Obtained From Swab And Tissue Biopsy

A.GRAM POSITIVE ORGANISMS

| | SWAB | % | TISSUE | % |
|---------------------------------|------|-------|--------|-------|
| | | | BIOPSY | |
| 1. Staphylococcus aureus | 56 | 36.36 | 48 | 31.57 |
| 2.Enterococcus spp | 9 | 5.84 | 5 | 3.28 |
| 3. Staphylococcus epidermidis | 7 | 4.54 | 2 | 1.31 |
| 4.Streptococcus pneumonia | 1 | 0.64 | 1 | 0.65 |
| 5.Alpha haemolytic Streptococci | 1 | 0.64 | - | - |
| 6.Beta haemolytic Streptococci | 1 | 0.64 | - | - |
| Total | 75 | | 56 | |

B.GRAM NEGATIVE ORGANISMS

| | SWAB | % | TISSUE BIOPSY | % |
|--------------------------|------|-------|---------------|-------|
| 1.Klebsiella spp | 38 | 24.67 | 46 | 30.26 |
| 2.Escherichia coli | 35 | 22.72 | 37 | 24.34 |
| 3.Pseudomonas aeruginosa | 32 | 20.77 | 29 | 19.07 |
| 4.Acinetobacterspp | 25 | 16.23 | 23 | 15.13 |
| 5.Proteus spp | 11 | 7.14 | 11 | 7.2 |
| 6.Citrobacter freundii | 3 | 1.94 | 4 | 2.63 |
| 7.Enterobacterspp | 1 | 0.64 | 1 | 0.65 |
| Total | 145 | | 151 | |

8 isolates of *Staphylococcus aureus*, 3 *Pseudomonas aeruginosa* and 1 *Beta haemolytic Streptococci* were grown in swab but missed from tissue biopsy specimen. However 8 isolates of *Klebsiella spp*, 2 *Escherichia coli* and 1 *Citrobacter freundii* were grown in tissue biopsy but missed from swab.

Table-5 Concordances Between Swabs And Deep Tissue Cultures

| 12 INDIAN JOURNAL OF APPLIED RESEARCH | | | | |
|---|--------------------|--|--|--|
| Only growth in Tissue biopsy | 17/152 (11.038%) | | | |
| Only growth in Swab | 24/154 (15.584%) | | | |
| Isolates not grown in swabs but found in deep tissue | 3/154 (1.9480%) | | | |
| Tissue biopsy yielded all isolates found in Swab plus additional growth | 121/152(79.6%) | | | |
| Swabs that yielded all isolates found in Deep tissue plus additional growth | 130/154 (84.415%) | | | |
| Swabs that yielded all isolates cultured from Deep tissue | 121/154 (78.571%) | | | |
| Swaha that wielded all isolates cultured from | 121/154 (70 5710/) | | | |

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Table- 6 A. Antibiotic Susceptibility Pattern Of Gram Positive Organisms

| | SWAB | | TISSU | E BIOPSY |
|------------------------|------|------|-------|----------|
| ANTIBIOTICS TESTED | S | % | S | % |
| 1.linezolid(LZ) | 73 | 100 | 52 | 92.8 |
| 2.vancomycin(VA) | 62 | 82.6 | 47 | 83.9 |
| 3.amikacin(AK) | 46 | 62.1 | 38 | 67.8 |
| 4.cotrimoxazole(COT) | 46 | 61.3 | 35 | 62.5 |
| 5.erythromycin(E) | 38 | 58.4 | 33 | 60 |
| 6.ampicillin(AMP) | 38 | 50.6 | 32 | 58.1 |
| 7.penicillin(P) | 9 | 45.3 | 13 | 27 |
| 8.ciprofloxacin(CIP) | 30 | 44.1 | 25 | 44.6 |
| 9.gentamicin(GEN)/ HLG | 27 | 42.8 | 19 | 34.5 |
| 10.cefoxitin | 13 | 23.2 | 12 | 25 |

HLG-high level gentamicin

B. ANTIBIOTIC SUSCEPTIBILITY PATTERN OF GRAM NEGATIVE ORGANISMS

| | SWAB | | TISSUE | BIOPSY |
|------------------------------------|------|-------|--------|--------|
| ANTIBIOTIC | S | % | S | % |
| 1.piperacillin-tazobactam(PTZ) | 36 | 90 | 36 | 80 |
| 2.imipenem(IMP) | 122 | 84.7 | 130 | 88.4 |
| 3.amikacin(AK) | 80 | 55.17 | 86 | 56.9 |
| 4.aztreonam (AT) | 18 | 54.5 | 26 | 56.5 |
| 5.ceftazidime clavulinic acid(CAC) | 63 | 43.7 | 68 | 45.6 |
| 6.amoxicillin clavulinic acid(AMC) | 60 | 41.3 | 61 | 40.6 |
| 7.ceftriaxone(CTR) | 51 | 35.9 | 64 | 42.3 |
| 8.ciprofloxacin(CIP) | 47 | 32 | 58 | 38.4 |
| 9.cotrimoxazole(COT) | 43 | 29.6 | 58 | 38.6 |
| 10.ampicillin(AMP) | 41 | 28.4 | 58 | 38.4 |
| 11.ceftazidime(CAZ) | 38 | 26 | 50 | 33.1 |
| 12.gentamicin(GEN) | 26 | 25.7 | 51 | 34 |

Table -7 Mrsa And Esbl Strains With Percentage

| MRSA 43(76.8%) 36(75%) 79(75.96%) ESBL 104(71.7%) 97(64.2%) 201(67.9%) | | SWAB (%) | TISSUE BIOPSY (%) | Total (%) |
|--|------|------------|-------------------|------------|
| ESBL 104(71.7%) 97(64.2%) 201(67.9%) | MRSA | 43(76.8%) | 36(75%) | 79(75.96%) |
| | ESBL | 104(71.7%) | 97(64.2%) | 201(67.9%) |

MRSA - methicillin resistant *Staphylococcus aureus*, ESBL - extended spectrum β -lactamase

DISCUSSION

In the present study, a total 200 patients were enrolled over a period of 18 months. Most common age group with diabetic foot was between 50–60 years, probably due to poor glycemic control. In a study by Hena *et al*¹⁷ the common age group affected was 56-65 years with average of 58 yrs. In another study by Hafni AA*et al*¹⁸ the common age group affected was 51-60 years.

In the present study 61.5% were males and 38.5% were females. In a study conducted by Siham Sh *et al*¹⁹ in 2013, 68% were males and 32% were females. Bengalorkar G *et al*²⁰ showed 70% males and 30% females which are all in concordance with our study. Male preponderance was due to more exposure to injuries during their occupational and recreational activities.

SWAB V/S TISSUE BIOPSY Table-8 Comparison Of Culture Outcome

| | Swab | Tissue biopsy |
|------------------------|------|---------------|
| Present study | 77% | 76% |
| Bozkurt ²¹ | 68% | 52% |
| Nelson ⁷ | 70% | 86% |
| Haalboom ²² | 96% | 77% |

Determination of the microbiological cause of diabetic foot ulcer relies on the sampling method used. Study conducted by Bozkurt *et al* and Haalboom *et al* have shown that swab technique yielded better result and to be a better method. However Nelson *et al* had shown tissue biopsy to be a better method.

In our study out of 200 patients, pathogens reported was 77% from swabs and 76% from tissue biopsies.78.5% swabs yielded all organisms that were cultured from deep tissue biopsy. Organisms that

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were not grown in swab but grown in tissue biopsies were 1 isolate of Citrobacter species, 2 Escherichia coli and 8 Klebsiella species. Organisms that were not grown in tissue but grown in swab were 8 Staphylococcus aureus, 3 Pseudomonas aeruginosa and 1 Beta haemolytic Streptococci.

Swab culture yields more organisms than tissue biopsy culture. Calculated P value is not significant. Swab is preferable because it is more specific, economical, easy technique and more compliable from patients.

Table 9 Causative Organisms

| Study | Staphylococc | | | Pseudomo |
|----------------------------------|---------------|----------|---------|----------|
| | us aureus and | a spp(%) | coli(%) | nas |
| | MRSA(%) | | | spp(%) |
| Present study | 36 | 24.6 | 22 | 20 |
| Hena JV et al17 | 42.3 | 9 | 15.3 | 24.3 |
| Gadepalli R et al4 | 13.7 | 6.6 | 12 | 9.8 |
| Jain M <i>et al</i> ² | 12.7 | 22.29 | 16.56 | 30.57 |
| Anandi C et al ²³ | 13.6 | 13.6 | 27.7 | 11.3 |

Staphylococcus aureus was the most common organism isolated in our study which is similar to the studies conducted by Hena JV et al and Gadepalli R et al whereas Pseudomonas spp and E.coli were the common organisms which were isolated in the studies Jain M et al and Anandi C et al respectively.

ANTIBIOTIC SUSCEPTIBILITY

In our study Gram positive bacteria were more sensitive to linezolid, vancomycin, amikacin and cotrimoxazole Gram negative bacteria were more sensitive to piperacillin-tazobactam, imipenem, amikacin and aztreonam by swab and tissue culture.

DRUG RESISTANCE

Multidrug resistance is a worrying global health issue as infections caused by them are associated with higher morbidity and mortality. Our study shows 75.96% MRSA and 67.9% ESBL. They have limited therapeutic options to treat them which may result in poor clinical outcome. ESBL-producing organisms frequently exhibit resistance to other antimicrobial agents may be due to plasmid encoded/ chromosomal mediated. Therefore early detection of these bacteria is important to control and prevent nosocomial outbreaks in hospital settings.

CONCLUSION

This study gives us knowledge about the prevalence of diabetic foot ulcers in our hospital and an insight of the causative organisms of diabetic ulcer and their sensitivity pattern. Higher prevalence of multidrug resistance was observed in our study warranting prompt need of surveillance for the effective management of such MDR strains. It also discourages the indiscriminate use of antibiotics. Hence there is a need to select an appropriate technique to ensure continued surveillance in order to combat bacterial drug resistance.

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