Original Resea	Volume - 11   Issue - 10   October - 2021   PRINT ISSN No. 2249 - 555X   DOI : 10.36106/ijar
al Of Appli	Biochemistry
CLEASE HOLD	PHYTOCHEMICAL PROFILING OF OXALIS CORNICULATA LEAF EXTRACT
Remya A. S.	Department of Biochemistry, University of Kerala, Kariavattom, Thiruvananthapuram- 695581, Kerala, India. Sree Narayana College For Women (Affiliated to University of Kerala), Kollam-691001, Kerala, India.
Annie Abraham*	UGC-BSR Faculty Fellow, Department of Biochemistry, University of Kerala, Kariavattom, Thiruvananthapuram-695581, Kerala, India. *Corresponding Author
from me to analyze plant extracts as this Fourier transform infrared (FTII spectrum provides information	tion of the phytochemical composition of plant extract is an essential step in developing novel therapeutic agents edicinal plants. Mass spectrometry, coupled with chromatographic separations (GC/MS), is increasingly applied technique has proved to be a valuable method for the analysis of nonpolar volatile components and alkaloids. R) is being used for the identification of characteristic functional groups present in the plant extract. The absorption about the structure of the molecules present in the phytoextract. FTIR analysis revealed the presence of phenols, arcmatic components alcohol and brow alkage compounds in methanolic leaf extract of <i>Cralic corriculata and</i>

GC/MS analysis provide different peaks determining the presence of twenty three phytochemical compounds. KEYWORDS : Phytochemicals, Oxalis corniculata

# INTRODUCTION

The plant Oxalis corniculata Linn. belonging to the family Oxalidaceae is commonly known as creeping wood sorrel, is a delicate-appearing, low growing, herbaceous medicinally important plant with abundant distribution in damp shady places, roadsides, plantations and lawns. Various traditional medicinal systems use this plant for the treatment of varied ailments of humans. Oxalis corniculata has hypoglycemic, antihypertensive, antipsychotic, nervous system stimulant, chronotropic, and inotropic effects [Achola et al., 1995; Raghavendra et al., 2006]. Antioxidant and toxicity evaluation of different extracts of Oxalis corniculata was also investigated [Alam et al., 2011]. The plant also has antifungal and antibacterial activities [Satish et al., 2008]. The plant offers protective effects against nephrotoxicity in CCl4 induced rats [Khan and Zehra, 2013] and also offers protection against lung injuries induced by CCl<sub>4</sub> [Ahmad et al., 2015]. Results of the in vitro and in vivo studies using aqueous extract of Oxalis corniculata indicate that, by virtue of its potent antioxidant property, it can effectively scavenge free radicals and thereby reduce the risk of isoproterenol-induced oxidative stress and myocardial infarction in rats [Abhilash et al., 2011]. Research studies carried out on the plant so far have also been reviewed [Badwaik et al., 2011; Kathiriya et al., 2010], highlighting its diverse medicinal applications.

In view of extensive ethnomedicinal applications of the edible folkloric medicinal plant *Oxalis corniculata*, the study was undertaken to identify the flavonoids and volatile phytochemicals present in the flavonoid rich fraction of *Oxalis corniculata* leaf extract.

Infrared spectroscopy (IR) has the potential to provide biochemical information without disturbing the biological sample. Fourier transform infrared spectroscopy [FTIR], due to its high precision in absorbance and wavenumber measurements, use infrared spectra for the identification of biomolecules [Pillai and Nair, 2014]. It measures the vibrations of bonds predominantly within chemical functional groups and generates a spectrum that can be regarded as a biochemical or metabolic "fingerprint" of the sample. By FTIR, it is possible to identify concrete structures of plant secondary metabolites and offers a rapid and non-destructive investigation to fingerprint herbal extracts or powders [Yang and Yen, 2002 and Ivanova and Singh, 2003].

Gas Chromatography-Mass Spectroscopy [GC-MS] is a very compatible technique used to screen active components from medicinal plants that have efficient protection and treatment role against various diseases and is most commonly used for identification and quantification purpose [Muthuraj *et al.*, 2015]. The unknown organic compounds in a complex mixture can be determined by interpretation and also by matching the spectra with reference spectra [Hites, 1997].

## MATERIALS AND METHODS

20

The shade dried leaves (1 kg) of *Oxalis corniculata* were refluxed in 80% methanol at 65 °C for 24 h. The supernatant was collected and the

extraction was repeated twice. The extract was decanted, filtered and concentrated to remove the solvent in a rotary evaporator. The aqueous concentrate was extracted successively with petroleum ether (60–80  $^{\circ}$ C) and ethyl acetate.

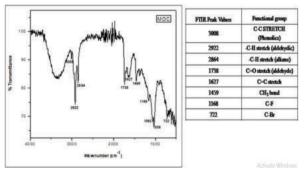
Ethyl acetate fraction gave positive Shinoda test for flavonoids and was used for further phytochemical analysis. The ethyl acetate extract was concentrated and subjected to phytochemical analysis. FTIR analysis was performed using the Perkin Elmer spectrophotometer system which was used to detect the characteristic peaks and their functional groups. The peak values of the UV-VIS and FTIR were recorded.

The GC-MS was performed by using Shimadzu QP2010 Ultra Model gas chromatograph interfaced to a mass spectrometer instrument employing the following conditions: The fused silica column was packed with Rxi-5Sil MS (5% Phenyl 95% dimethylpolysiloxane, 30m x 250µm) fused silica capillary column (30 x 0.25mm ID x 1µMdf, composed of 100% Dimethyl polysiloxane), operating in electron impact mode at 70 eV; Helium gas (99.999%) was used as carrier gas at a constant flow of 1 ml/min and an injection volume of 0.5 µl was employed (split ratio of 20:1) injector temperature 260 °C; ion-source temperature 200 °C.

The oven temperature was programmed from 110 °C (isothermal for 2 min), with an increase of 10 °C/min, to 200°C, then 5°C/min to 280°C, ending with a 9 min isothermal at 280°C. Mass spectra were taken at 70eV, a scan-interval of 0.5 seconds, and fragments from 40 to 450 Da. The total GC running time is 50 min.

The relative percentage amount of each component was calculated by comparing its average peak area to the total areas. The identification of components was based on Willey and NIST libraries as well as the comparison of their retention indices.

## RESULTS





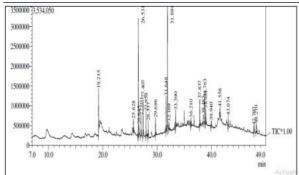


Figure 2: GC-MS Chromatogram of Oxalis corniculata leaf extract

Table 2: GC-MS analysis of phytochemical components
---

S1.	Retention	Name of compound	Peak area
No.	Time		(%)
1	19.215	2,4-Di-tert-butylphenol	9.77
2	25.628	9-eicosene, (E)-	1.65
3	26.531	Phytol, acetate	15.79
4	26.645	6,10,14-Trimethyl-pentadecan-2-Ol	1.73
5	27.031	3,7,11,15-Tetramethyl-2-hexadecen-1-ol	2.78
6	27.407	Neophytadiene	5.01
7	27.958	7,9-Di-tert-butyl-1-oxaspiro(4,5)deca-	3.72
		6,9-diene-2,8-dione	
8	28.337	Methyl palmitate	2.18
9	29.686	1-Nonadecene	2.43
10	31.648	Linolenic acid, methyl ester	5.62
11	31.884	Phytol	18.59
12	32.169	Methyl stearate	0.87
13	33.390	1-Tricosene	2.47
14	36.210	Butyl 9,12,15-octadecatrienoate	1.08
15	37.837	3-Cyclopentylpropionic acid, 2-	4.28
		dimethylaminoethyl ester	
16	38.483	Decane, 1,9-bis[(trimethylsilyl)oxy]-	4.04
17	38.763	2-Monopalmitin	7.69
18	38.951	1,2-Benzenedicarboxylic acid	2.36
19	39.940	Octadecyltrifluoroacetate	0.77
20	41.558	2,2-Dimethyl-3-vinyl bicyclo[2.2.1]heptane	1.16
21	43.074	Squalene	1.62
22	47.797	N-hentriacontanol-1	1.33
23	48.210	Tocopherol	3.07

### DISCUSSION

The FTIR spectrum helps to identify the functional groups of the active components present in the extract sample based on the peaks values in the region of IR radiation. The broadband at 3008 cm-1 corresponds to O-H stretching H-bonded alcohols and phenolic compounds in the leaf extract. The more intense bands occurring at 2922 cm<sup>-1</sup> stretch indicate the presence of carbohydrates. The absorption band at 2922 cm<sup>-1</sup> and 2864 cm<sup>-1</sup> indicates C-H stretch in methyl (-CH<sub>3</sub>) and methylene (-CH<sub>2</sub>) aliphatic saturated (C-H) sharp asymmetric and symmetric stretching. The C=O stretching observed at 1738 cm<sup>-1</sup> is due to the presence of the carbonyl group. The band at 1627 cm<sup>-1</sup> shows the presence of aromatic ring (C=C). The peak at 1460 cm<sup>-1</sup> indicates CH<sub>2</sub> bending vibration of alkane. Peaks at 1168 cm<sup>-1</sup> confirm the presence of C=O polysaccharide. The peaks 1168, 1038 and 722 cm<sup>-1</sup> indicate the presence of organic halogen compounds in Oxalis corniculata leaf extract. The above infrared functional group characteristics were cited in the literature [Ranjana and Mendhulkar 2015; John C, 2000; Saranya and Sekar, 2016; Liton and Islam, 2006]. The results of the present study confirm the presence of phenol, alkane, alkene, carboxylic acid, aromatic compound, alcohol, and Bromo alkane compounds in methanolic leaf extract of Oxalis corniculata.

Figure 2 shows the results of the GC-MS analysis of the flavonoid rich fraction of Oxalis corniculata leaf extract. The twenty three major peaks are tabulated in Table-2. The most phytochemical peak area were depicted by phytol (18.59%), 2-Monopalmitin, linoleic acid and neophytadiene. Phytols are diterpenes member of the long-chain unsaturated acyclic alcohols having a wide spectrum of biological effects. 2-Monopalmitin and linoleic acid are the major fatty acid

constituents of the leaf extract. The compound neophytadiene detected is a diterpene with anti-inflammatory properties [Bhardwa et al., 2020].

#### CONCLUSION

The FTIR analysis clearly indicates that the leaf extract is high in antioxidant phenolics and flavonoids. The GC-MS study also shows many phytochemicals like tocopherols, 2,4-Di-tert-butyl phenol, ethyl ester, Phytol, Linolenic acid, 1,2-Benzenedicarboxylic acid, 2-Monopalmitin, Squalene, etc., all of which contributes the activities like antimicrobial, antioxidant, anticancer and other pharmacological activities.

## REFERENCES

- Achola, K. J., Mwangi, J. W., & Munenge, R. W. (1995). Pharmacological activity of 1 Oxalis corniculata. International journal of pharmacognosy, 33(3), 247-249. Raghavendra, M. P., Satish, S., & Raveesha, K. A. (2006). Phytochemical analysis and 2
- antibacterial activity of Oxalis corniculata; a known medicinal plant. My science, 1(1), 72-78.
- Alam, M. B., Hossain, M. S., Chowdhury, N. S., Mazumder, M., Haque, M. E., & Islam, 3. A. (2011). In vitro and in vivo antioxidant and toxicity evaluation of different fractions of Oxalis corniculata Linr
- Satish, S., Raghavendra, M. P., & Raveesha, K. A. (2008). Evaluation of the antibacterial Δ potential of some plants against human pathogenic bacteria. Advances in biological research, 2(3-4), 44-48.
- Khan, M. R., & Zehra, H. (2013). Amelioration of CCl4-induced nephrotoxicity by 5. Oxalis corniculata in rat. Experimental and toxicologic pathology, 65(3), 327-334.
- Ahmad, B., Khan, M. R., & Shah, N. A. (2015). Amelioration of carbon tetrachloride 6. induced pulmonary toxicity with Oxalis corniculata. Toxicology and industrial health, 31(12), 1243-1251
- 7. Abhilash, P. A., Nisha, P., Prathapan, A., Nampoothiri, S. V., Cherian, O. L., Sunitha, T. K., & Raghu, K. G. (2011). Cardioprotective effects of aqueous extract of Oxalis corniculata in experimental myocardial infarction. Experimental and toxicologic pathology, 63(6), 535-540.
- 8 Badwaik, H., Singh, M. K., Thakur, D., Giri, T. K., & Tripathi, D. K. (2011). The botany, Schemistry, pharmacological and therapeutic application of Oxalis corniculata Linn-a review. *International Journal of Phytomedicine*, 3(1),01.Kathiriya, A., Das, K., Kumar, E. P., & Mathai, K. B. (2010). Evaluation of antitumor
- 9. and antioxidant activity of Oxalis corniculata Linn. against Ehrlich ascites carcinoma on mice
- 10. Pillai, L. S., & Nair, B. R. (2014). Functional group analysis of Cleome viscosa L. and C. burmanni W. & A.(Cleomaceae) extracts by FT-IR. Journal of Pharmacognosy and Phytochemistry, 2(6).
- 11. Yang, J., & Yen, H. E. (2002). Early salt stress effects on the changes in chemical composition in leaves of ice plant and Arabidopsis. A Fourier transform infrared spectroscopy study. *Plant physiology*, 130(2), 1032-1042.
- 12. Ivanova, D. G., & Singh, B. R. (2003). Nondestructive FTIR monitoring of leaf senescence and elicitin induced changes in plant leaves. Biopolymers: Original
- senescence and elicitin.induced changes in plant leaves. *Disponents. Crig.net. Research on Biomolecules*, 72(2), 79-85. Muthuraj, K., Shalimol, A., Sivapriya, K. T., & Nagarajan, N. (2015). Screening of active phytocompounds by GC-MS analysis and in vitro antibacterial activity of endemic plant Pogostemon mollis benth. *Inter J Rec Adv Multi Res*, 2(7), 534-9. Hites, R. A. (1997). Gas characterized by Sattle F. 13.
- 14 for Analytical Chemistry. edn. Edited by Settle F. Ranjana, S., & Mendhulkar, V. D. (2015). FTIR studies and spectrophotometric analysis
- 15 of natural antioxidants, polyphenols and flavonoids in Abutilon indicum (Linn) sweet leaf extract. Journal of Chemical and Pharmaceutical Research, 7(6), 205-211.
- John, C. (2000). Interpretation of infrared spectra, a practical approach. Encyclopedia of 16. analytical chemistry, 12, 10815-10837. Saranya, D., & Sekar, J. (2016). GC-MS and FT-IR Analyses of Ethylacetate Leaf
- 17. extract of Abutilon indicum (L.) Sweet. Int. J. Adv. Res. Biol. Sci, 3(2), 193-197.
- 18 Liton, A. K., & Islam, M. R. (2006). Condensation reaction of benzil with resorcinol and the establishment of the spectral data as well as cytotoxicity study. Bangladesh Journal of Pharmacology, 1(2), 58-63.

21