



POLYCAPROLACTONE NANOPARTICLES AS DELIVERY VEHICLE IN COMBATING DISEASES

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ABSTRACT Conventional chemotherapy against various diseases causes severe side effects in the biological system. To overcome drug toxicity, insolubility and resistance, and to get greater therapeutic efficacies, vesicular biodegradable and biocompatible polymeric nanoparticles have attracted attention as drug delivery vehicle. Polycaprolactone (PCL) nanoparticles owing to their nanosizes, spherical shapes and easy surface functionalization capability with drug encapsulation characteristics are utilized as drug carrier for targeted biomedical applications. This review is mainly focused on the preparation of polycaprolactone nanoparticles with their surface functionalization and their usages in different targeted drug delivery applications.

KEYWORDS : Diseases; Chemotherapy; Side effects; Polycaprolactone nanoparticles; Targeted drug delivery applications

INTRODUCTION

Pharmaceutical drugs used as chemotherapeutics against various diseases, encounters several limitations such as biological barriers, non-specificity, instability, fast degradation, insolubility and resistance before transporting to the targeted site/s.^[1-3] To conquer these barriers, it is needed to administer high drug dosage for achieving the preferred concentration at the targeted zone/s leading to cytotoxicity and other unwanted pathological side effects.^[4] These adverse effects may be overcome through the employment of polymeric cargo delivery systems for getting desired concentration of cargo to the specific targeted area/s.^[1,5-8] FDA-approved biodegradable and biocompatible polycaprolactone nanoparticles (PCL NPs) have been attracted attention not only as delivery system but also as vaccine carrier for their nanosize, cell penetrating capability, non-immunogenic characteristics and sustained cargo-release property to the targeted site/s of interest.^[9-14] PCLs, the semi-crystalline hydrophobic polymers having relatively polar ester groups with five non-polar methylene groups in the repeating units, are produced by ring-opening polymerizations of ϵ -caprolactone monomers, while the high olefin contents provide poly-olefin-like features to PCL.^[13,15] By virtue of the high degree of hydrophobicity and crystallinity, PCLs generally degrade slowly and are less biocompatible with biological cells leading to restricted long term bioavailable treatment against diseases as drug delivery system or vaccine carrier.^[16,14] Therefore, surface-modification of PCL by copolymers and / or ligands may be suitable to get greater biological efficacies associated with their improved hydrophilicity, non-toxicity, non-antigenicity, non-immunogenicity and biodegradability.^[17-19] The synthetic biodegradable polymers are very suitable for biomedical application against diseases as they are biologically inert, having lot to lot homogeneity and their composition and structure are easily functionalized for specific application.^[20,21] The hydrophilic, non-toxic and non-immunogenic polyethylene glycol (PEG) has great effects on cargo adsorption, prevention and escape from capture of the reticuloendothelial system increasing blood circulation time of PEGylated PCL NPs to get maximum delivery to targeted region/s.^[22,23] In addition, PEG side chains further enable functionalization of NPs utilizing well designed specific bio-ligands to get greater targeting of the encapsulated cargo through the transportation to the zone/s of the intended liberation.^[24] Moreover, restricted smaller nanosized PCL particles coupled with targeting ligand can specifically bind to target cells after administration for internalizing the NPs to release cargos inside the cells with higher affinity and specificity.^[25-27] Furthermore, as traditional vaccines have susceptibility to degradation, short duration of action, side effects and inflammatory reactions, polymeric nanoparticles have attracted attention to overcome these barriers. PCL NPs as carrier / adjuvant may promote and induce strong cellular Th1, humoral Th2 and mucosal immune responses to enhance the vaccines immune efficacy and achieve controlled release and mucosal immunities.^[28-32] This review elucidates the therapeutic efficiencies of PCL NPs against various diseases to consider as favorable delivery carrier.

to prepare the PCL NPs with the requirements of organic phase (solvent phase) and aqueous phase (non-solvent phase).^[10,33,34] In brief, PCL is liquefied in a suitable organic solvent, while the organic phase is added gently into a surfactant containing watery phase under continual magnetic stirring. The organic solvent is then evaporated out at ambient temperature with continuous stirring or by using a rotary evaporator. After that, the solution is centrifuged and lyophilized to prepare PCL NPs.^[33,35-38] In general, aqueous phase containing various hydrophilic surfactants such as polysorbate 80 / tween®80 / polyvinyl alcohol is used for preventing the aggregation of the NPs. In this context, different operating conditions such as polymer molecular weight and amount, surfactant amount and stirring rate in this method may influence the size of the NPs.^[33] For encapsulation of cargos in the NPs, various hydrophobic drugs / bioactive molecules are dissolved in appropriate solvent and adjoined to the prepared PCL solution prior to the inclusion of the organic phase into the watery phase.^[39,10,38] On the other hand, several hydrophilic molecules / drugs may also be encapsulated effectively into the PEGylated PCL NPs by dissolving the cargos in an external watery phase or usages of co-solvent or suspension in minimum quantity of cargos in the organic phase.^[10,33,40,41] Coupling of PCL and the amino-aptamer is made with N-hydroxysuccinimide (NHS) and dicyclohexylcarbodiimide (DCC) reagents. In brief, an amount of 2.5 nmol S15-aptamer is amalgamated with 34.4 μ L 70 mM carbonate buffer (sodium carbonate and sodium hydrogen carbonate, pH 10). After that, 12.5 μ mol NHS and DCC, and 250 nmol PCL_{5.4} dissolved in 100 μ L di-methylsulfoxide (DMSO) at 70°C, is adjoined into the aptamer. After ultrasonication for 2 min, the reaction mixture is incubated at 500 rpm for 6 h at 20°C. Each 1 h new NHS and DCC (each 12.5 μ mol in 40 μ L DMSO) are adjoined following ultrasonication for 2 min. The solvents are then removed under reduced pressure to get solid conjugates, while the water-resuspended aptamer-PCL_{5.4} NPs are refined with reversed phase high performance liquid chromatography (RP-HPLC).

Determination of encapsulation efficiency and drug loading

The drug loading and encapsulation efficiency of nanoparticulated formulations are assayed quantitatively by RP-HPLC. The prepared drug loaded nanoparticles are dissolved in 2 mL methanol for 24 hrs at 4°C. The supernatant suspension containing drug is collected by centrifugation. The drug loading and encapsulation efficiency of PCL NPs are estimated by the following formulas:

Encapsulation efficiency (%) = $\frac{[\text{Total amount of formulated drug} - \text{Amount of supernatant drug}]}{\text{Total amount of formulated drug}} \times 100$.
Drug loading (%) = $\frac{[\text{Total amount of formulated drug} - \text{Amount of supernatant drug}]}{\text{Polymeric mass}} \times 100$.

Characterization of nanoparticles-conjugates

Transmission electron microscope and scanning electron microscope are used to determine the shape, size and surface morphology of the NPs. The average particle sizes and polydispersity index are measured by dynamic light scattering utilizing zetasizer nano-ZS. The zeta potential of the aqueous NPs dispersion is determined by zetasizer nano-ZS at 25°C. The interactions of drug and polymer are investigated by Fourier transform infrared spectrometer. The interactions among different ingredients in the NPs-conjugates are estimated by differential scanning calorimeter.

Preparation and functionalization of polycaprolactone nanoparticles

The nanoprecipitation / solvent displacement method may be utilized

Polycaprolactone nanoparticles in delivery applications

PCL NPs have been used in different drug delivery applications for achieving better therapeutic efficiencies with sustained targeting to specific site of interests.^[42-44] Curcumin-loaded PCL NPs have shown their high impact in therapeutic efficacies.^[45] Carboplatin-loaded PCL NPs have been utilized for treating glioma through nasal route having no severe damage on the nasal mucosal integrity.^[46] Hydrophobic tamoxifen-loaded polyethylene oxide -modified PCL NPs have shown their enhanced level of drug accumulation within tumor for anticancer therapy.^[47] Biodegradable PCL and non-biodegradable polycationic acrylic polymer blended NPs have been used to deliver orally insulin for the treatment of diabetes.^[48] Hydrophobic clonazepam -loaded poly(N-isopropylacrylamide)-b-poly(ϵ -caprolactone) NPs have been utilized to achieve desired drug release motif having additional diffusion barrier to the encapsulated drug by the poly(N-isopropylacrylamide) layer on the surfaces of NPs.^[49] Curcumin encapsulated polycaprolactone-montmorillonite nanoclay -functionalized n-cetyl-N,N,N-trimethylammonium bromide NPs have been delivered for enhancing their antitumor activities.^[50] Vinblastine-loaded PCL-grafted dextran NPs have been used to treat breast cancer cells for getting efficient cellular uptake of NPs.^[51] Mucoadhesive chitosan-PCL NPs have been utilized to combat H1N1 influenza virus as a vehicle for nasal vaccines for getting effective humoral and cellular immune responses and mucosal antibodies.^[30,31] Methoxy PEG-PCL has been used as ecotropic diblock copolymer for the hydrophobic drugs-deliveries.^[32] Quercetin-biapiogenin encapsulated PCL NPs have been applied as synergistic actions of antioxidants to get effective hepatoprotective effects.^[53] A lot of studies have indicated that mPEG-PCL NPs have been utilized as vehicles for protein, genes and vaccines cargos specifically in a sustained liberation drug delivery system for their specific targeting to cells to get higher biological effectiveness.^[53]

Biodegradation of polycaprolactone

Polycaprolactone is degraded through two discrete stages in physiological conditions: (i) Non-enzymatic hydrolysis of ester linkage of PCL attributing to haphazard chain scission and resulting reduced molecular weight, (ii) Intracellular degradation of highly crystalline and low molecular weight (<3000) -PCL.^[42,54-56] Generally, PCL is degraded from few months to several years depending on the starting molecular weight.^[42,21]

Conclusions and future perspectives

As biodegradation rate of PCL NPs is slower, it may be utilized to overcome biological barriers as drug delivery vehicle for long term treatment against chronic diseases. Moreover, PCL NPs may be copolymerized with PEG and / or conjugated with ligands and loaded with cargos to deliver drugs to targeted site/s with sustained release and accumulation for achieving higher efficacies at lower dosages against various diseases. However, this delivery system needs more explorations regarding their thorough systemic long term studies on biodistribution, pharmacokinetics, cytotoxicity, elimination, immune responses, appropriate size, shape and suitable surface functionalizations with surfactants, cargos, ligands and polymer coating, administrative routes specially oral and intravenous, and efficacies for the specific targeting to diseased cells before going to clinics to consider as suitable nanoparticulated delivery system for biomedical applications in combating different diseases.

Conflict of interest

None.

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REFERENCES

- Ponnappan N, Chugh A. Nanoparticle-mediated delivery of therapeutic drugs. *Pharmaceut Med* 2015; 29:155-167.
- Senapati S, Mahanta AK, Kumar S, Maiti P. Controlled drug delivery vehicles for cancer treatment and their performance. *Signal Trans Target Ther* 2018; 3:7.
- Mandal AK. Gallium nanoparticles as delivery system against infectious diseases and cancer. *Ind J Appl Res* 2020; 10(10):10.36106/ijar.
- Torchilin VP. Drug targeting. *Eur J Pharm Sci* 2000; 11:S81-S91.
- Ibrahim KE, Bakhtiet AO, Khan A, Khan HA. Recent trends in biomedical applications of nanomaterials. *Biosci Biotechnol Res Asia* 2018; 15:235-243.
- Mandal AK. Dendrimers in targeted drug delivery applications: A review of diseases and cancer. *Int J Polym Mater Polym Biomater* 2021; 70(4):287-297.
- Mandal AK. Poly alkyl cyanoacrylate nanoparticles as delivery vehicle in combating diseases. *Int J Sci Res* 2020; 9(11):10.36106/ijrs.
- Sana S, Ghosh S, Das N, Sarkar S, Mandal AK. Vesicular melatonin efficiently downregulates sodium fluoride-induced rat hepato and broncho TNF- α , TGF- β expressions and associated oxidative injury: A comparative study of liposomal and

- nanocapsulated forms. *Int J Nanomed* 2017; 12:4059-4071.
- Rejman J, Oberle V, Zuhorn IS, Hoekstra D. Size-dependent internalization of particles via the pathways of clathrin and caveolae-mediated endocytosis. *Biochem J* 2004; 377:159-169.
- Kumari A, Yadav SK, Yadav SC. Biodegradable polymeric nanoparticles based drug delivery systems. *Colloids surf B Biointerfaces* 2010; 75:1-18.
- Modrejewski J, Walter JG, Kretschmer I, Komal E, Green M, Belhadi H, et al. Aptamer-modified polymer nanoparticles for targeted drug delivery. *BioNanoMater* 2016; 17(1-2):43-51.
- Kim SY, Lee YM. Taxal-loaded block copolymer nanospheres composed of methoxy poly(ethylene glycol) and poly(ϵ -caprolactone) as novel anticancer drug carriers. *Biomater* 2001; 22:1697-1704.
- Mei L, Zhang Y, Zheng Y, Tian G, Song C, Yang D, et al. A novel paclitaxel-loaded poly(ϵ -caprolactone) / pluronic F68 nanoparticle overcoming multidrug resistance for breast cancer treatment. *Nanoscale Res Lett* 2009; 4:1530-1539.
- Guo S, Fu D, Utupova A, Sun D, Zhou M, Jin Z, et al. Applications of polymer-based nanoparticles in vaccine field. *Nanotechnol Rev* 2019; 8:143-155.
- Kim J, Bae Y. Albumin loaded microsphere of amphiphilic poly(ethylene glycol) / poly(α -ester) multiblock copolymer. *Eur J Pharmaceut Sci* 2004; 23:245-251.
- Jia WJ, Gu YC, Gou ML, Dai M, Li XY, Kan B, et al. Preparation of biodegradable polycaprolactone / poly(ethylene glycol) / polycaprolactone (PCEC) nanoparticles. *Drug Deliv* 2008; 15:409-416.
- Chen DR, Bei JZ, Wang SG. Polycaprolactone microparticles and their biodegradation. *Polym Degrad Stab* 2000; 67:455-459.
- Huang MH, Li S, Hulmacker DW, Schantz JT, Vacanti CA, Braud C, et al. Degradation and cell culture studies on block copolymers prepared by ring opening polymerization of ϵ -caprolactone in the presence of poly(ethylene glycol). *J Biomed Mater Res* 2004; 69:417-427.
- Moon HT, Lee YK, Han JK, Byun YJ. Improved blood compatibility by sustained release of heparin deoxycholic acid conjugates in a PCL-PEG multiblock copolymer matrix. *J Biomater Sci, Polym Edition* 2002; 13:817-828.
- Nair LS, Laurencin CT. Biodegradable polymers as biomaterials. *Prog Polym Sci* 2007; 32:762-798.
- Gunatillake PA, Adhikari R. Biodegradable synthetic polymers for tissue engineering. *Eur Cells Mater* 2003; 5:1-16.
- Ryu JG, Jeong YI, Kim YH, Kim IS, Kim DH, Kim SH. Clonazepam release from core-shell typen nanoparticles of poly(ϵ -caprolactone)-poly(ethylene glycol)-poly(ϵ -caprolactone) triblock copolymers. *Bull Korean Chem Soc* 2001; 22(5):467-475.
- Zhang Y, Zhuo R. Synthesis and in vitro drug release behavior of amphiphilic triblock copolymers nanoparticles based on poly(ethylene glycol) and polycaprolactone. *Biomater* 2005; 26:6736-6742.
- Venuta A, Moret F, Dal Poggetto G, Esposito D, Fraix A, Avitabile C, et al. Shedding light on surface exposition of poly(ethylene glycol) and folate targeting units on nanoparticles of poly(ϵ -caprolactone) diblock copolymers: Beyond a paradigm. *Eur J Pharm Sci* 2018; 111:177-185.
- Zhang H, Zhou L, Zhu Z, Yang C. Recent progress in aptamer-based functional probes for bioanalysis and biomedicine. *Chem* 2016; 22:9886-9900.
- Tuerk C, Gold L. Systematic evolution of ligands by exponential enrichment: RNA ligands to bacteriophage T4 DNA polymerase. *Sci* 1990; 249:505-510.
- Zhang Y, Chen Y, Han D, Ocsosy I, Tan W. Aptamers selected by cell-SELEX for application in cancer studies. *Bioanal* 2010; 2(5):907-918.
- Vijaya BJ, Sean MG, Aliasger KS. Biodegradable particles as vaccine antigen delivery systems for stimulating cellular immune responses. *Hum Vacc Immunother* 2013; 9:2584-2590.
- Haas J, Kumar MNR, Borchard G, Bakowsky U, Lehr CM. Preparation and characterization of chitosan and trimethyl-chitosan-modified poly(ϵ -caprolactone) nanoparticles as DNA carriers. *AAPS Pharm Sci Tech* 2005; 6:E22-E30.
- Gupta NK, Tomar P, Sharma V, Dixit VK. Development and characterization of chitosan coated poly(ϵ -caprolactone) nanoparticulate system for effective immunization against influenza. *Vacc* 2011; 29:9026-9037.
- Jesus S, Soares E, Costa J, Borchard G, Borges O. Immune response elicited by an intranasally delivered HBSAg low-dose adsorbed to poly- ϵ -sialon-caprolactone based nanoparticles. *Int J Pharm* 2016; 504:59-69.
- Mehdi F, Ali H, Kobra R, Soghrat F. Efficiency of flubendazole-loaded mPEG-PCL nanoparticles: A promising formulation against the protozoocysts and cysts of *Schincococcus granulosis*. *Acta Trop* 2018; 187:190-200.
- Miladi K, Sfar S, Fessi H, Elaissari A. Nanoprecipitation process: From particle preparation to in vivo applications. In: *Polymer nanoparticles for nanomedicines: A guide for their design, preparation and development* (Vauthier C, Ponchel G. ed.). Springer International Publishing 2016; pp 17-53.
- Badri W, Miladi K, Robin S, Viennet C, Nazari QA, Agusti G, et al. Polycaprolactone based nanoparticles loaded with indomethacin for anti-inflammatory therapy: From preparation to ex vivo study. *Pharm Res* 2017; 34:1773-1783.
- Sinha VR, Bansal K, Kaushik R, Kumria R, Trehan A. Poly- ϵ -sialon-caprolactone microspheres and nanospheres: An overview. *Int J Pharm* 2004; 278:1-23.
- Bilati U, Allemann E, Doelker E. Development of a nanoprecipitation method intended for the entrapment of hydrophilic drugs into nanoparticles. *Eur J Pharm Sci* 2005; 24:67-75.
- Savant K, Pandey A, Patel S. Aripiprazole loaded poly(caprolactone) nanoparticles: Optimization and in vivo pharmacokinetics. *Mater Sci Eng C Mater Biol Appl* 2016; 66:230-243.
- Badri W, ElAsbahani A, Miladi K, Barakat A, Agusti G, Nazari QA, et al. Poly(μ -caprolactone) nanoparticles loaded with indomethacin and Nigella Sativa L. essential oil for the topical treatment of inflammation. *J Drug Deliv Sci Technol* 2018; 46:234-242.
- Fessi H, Puisieux F, Devissaguet JP, Ammoury N, Benita S. Nanocapsule formation by interfacial polymer deposition following solvent displacement. *Int J Pharm* 1989; 55:R1-R4.
- Azzech M, Szczepanowicz K, Jantas D, Piotrowski M, Kida A, Lason W, et al. Neuroprotective action of undecylenic acid (UDA) encapsulated into PCL nanocarriers. *Colloids Surf Phys Eng Asp* 2017; 532:41-47.
- Rafei P, Haddadi A. Docetaxel-loaded PLGA and PLGA-PEG nanoparticles for intravenous application: Pharmacokinetics and biodistribution profile. *Int J Nanomed* 2017; 12:935-947.
- Woodruff MA, Hutmacher DW. The return of a forgotten polymer-polycaprolactone in the 21st century. *Prog Polym Sci* 2010; 35:1217-1256.
- Merkli A, Tabatabay C, Gurmy R, Heller J. Biodegradable polymers for the controlled release of ocular drugs. *Prog Polym Sci* 1998; 23:563-580.
- Malikmammadov E, Tanir TE, Kizilay A, Hasirci V, Hasirci N. PCL and PCL-based materials in biomedical applications. *J Biomater Sci Polym Ed* 2018; 29:863-893.
- Kasinathan N, Amirthalingam M, Reddy ND, Jagani HV, Volety SM, Rao JV. In-situ implant containing PCL-curcumin nanoparticles developed using design of experiments. *Drug Deliv* 2016; 23:1007-1015.
- Alex AT, Joseph A, Shavi G, Rao JV, Udupa N. Development and evaluation of carboplatin-loaded PCL nanoparticles for intranasal delivery. *Drug Deliv* 2016;

- 23:1244-1253.
47. Shenoy DB, Amiji MM. Poly(ethylene oxide)-modified poly(ϵ -caprolactone) nanoparticles for targeted delivery of tamoxifen in breast cancer. *Int J Pharm* 2005; 293:261-270.
 48. Damge C, Maincent P, Ubrich N. Oral delivery of insulin associated to polymeric nanoparticles in diabetic rats. *J Control Rel* 2007; 117:163-170.
 49. Choi C, Chae SY, Nah JW. Thermosensitive poly(N-isopropylacrylamide)-b-poly(μ -caprolactone) nanoparticles for efficient drug delivery system. *Polymer(Guildf)* 2006; 47:4571-4580.
 50. Bakre LG, Sarvaiya JJ, Agarwal YK. Synthesis, characterization and study of drug release properties of curcumin from polycaprolactone / organomodified montmorillonite nanocomposite. *J Pharm Innov* 2016; 11:300-307.
 51. Prabu P, Chaudhari AA, Dharmaraj N, Khil MS, Park SY, Kim HY. Preparation, characterization, in-vitro drug release and cellular uptake of poly(caprolactone) grafted dextran copolymeric nanoparticles loaded-with anticancer drug. *J Biomed Mater Res Part A* 2008; 90A:1128-1136.
 52. Oliveira AL, Pinho C, Fonte P, Sarmiento B, Dias ACP. Development, characterization, antioxidant and hepatoprotective properties of poly(ϵ -caprolactone) nanoparticles loaded with a neuroprotective fraction of *Hypericum perforatum*. *Int J Biol Macromol* 2018; 110:185-196.
 53. Molina J, Urbina J, Gref R, Brener Z, Junior JMR. Cure of experimental Chagas' disease by the bis-triazole DO870 incorporated into 'stealth' polyethyleneglycol-poly lactide nanospheres. *J Antimicrob Chemother* 2001; 47:101-104.
 54. Azimi B, Nourpanah P, Rabiee M, Arbab S. Poly(μ -caprolactone) fiber: An overview. *J Eng Fiber Fabr* 2014; 9:74-90.
 55. Abedalwafa M, Wang F, Wang L, Li C. Biodegradable poly-epsilon-caprolactone (PCL) for tissue engineering applications: A review. *Rev Adv Mater Sci* 2013; 34:123-140.
 56. Woodward SC, Brewer PS, Moatamed F, Schindler A, Pitt CG. The intracellular degradation of poly(μ -caprolactone). *J Biomed Mater Res* 1985; 19:437-444.