

## Therapeutic Effect of Aqueous Extract of Vitis Vinifera L. Against Thyrotoxicosis Induced by Levothyroxine Sodium in Rabbit Females



### Chemistry

**KEYWORDS :** Vitis Vinifera L., Levo thyroxine sodium, LDH, ACP, ALP.

Taghreed U.Muhammd

Chemistry department, Colloge of Education for Pure Science – Ibn- Al-Haitham, Baghdad University

### ABSTRACT

To investigat the hepatoprotective potential of aqueous grape leaf extract on levothyroxine sodium induced liver toxicity in femals of rabbit. Liver injury was induced by levothyroxine sodium (5 µg) on the first day of experiment. Vitis vinifera L.leaf extract (100mg/ml) was administered orally daily for two months. The animals were sacrificed after two months and the hepatoprotective activity was assessed by using various biochemical parameters. In levothyroxine sodium treated femals of rabbit , the lactate dehydrogenase(LDH), acid phosphatase (ACP), and alkaline phosphatase (ALP) levels were significantly increased. The Vitis vinifera L.leaf extract administration therapy the toxic effect of levothyroxine sodium on the LDH, ACP, and ALP.

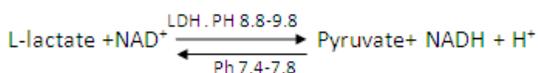
### Introduction

Vitis vinifera L. (Family: Vitaceae) is one of the most popular and the world's largest fruit crop[1]. Reviewing the current literature, it was found that Vitis vinifera L. contains many chemical constituents viz, phenolic acids, flavonoids, anthocyanins, proanthocyanidins, sugars, sterols, amino acids, and minerals [2]. Natural antioxidants have gained considerable interest in recent years for their role in preventing the auto oxidation of fats; Vitis vinifera L. plant is considered as a natural antioxidant source [3].

Vitis vinifera L. is considered as an economic plant, whose several cultivars which are grown for their edible fruits, the source of raisins, wines, sultanas and currents [4]. Grape are also used as demulcent, laxative, refrigerant, stomachic, diuretic and cooling. Moreover, it is useful in bilious dyspepsia, haemorrhage, dysuria, in chronic bronchitis heart disease and gout[5].

### Lactate dehydrogenase(LDH, EC 1.1.1.27):

is a hydrogen transfer enzyme that catalyses the oxidant of L-lactate to pyruvate with nicotinamide- dinucleotide (NAD)<sup>+</sup> as hydrogen acceptor; the final step in the metabolic chain of anaerobic glycolysis. The reaction is reversible and the reaction equilibrium strongly favours the revers reaction, namely the reduction of pyruvate(P) to lactate(L) [6]:



Acid and alkaline phosphatase: have been traditionally classified as being acid and alkaline, due to their optimum pH activity, above pH 7.0 or below pH 7.0 [7]. Acid phosphatase (EC 3.1.3.2) and alkaline phosphatase (EC 3.1.3.1) are enzymes that catalyze the removal of inorganic phosphate (orthophosphate) from organic phosphate esters, in acidic and alkaline media respectively. These phosphatase are ubiquitous in plants, animals and microorganisms[8].

### Thyroxine:

is the main hormone secreted into the bloodstream by the thyroid gland. It is inactive and most of it converted to an active form called triiodothyronine by organs such as the liver and kidneys. Thyroid hormones play vital roles in regulating the body's metabolic rate, heart and digestive functions, muscle control, brain development and maintenance of bones [9].

The aim of this study was to evaluate effect of aqueous extract of Vitis vinifera L. leaves on LDH, ACP and ALP levels in rabbit females have thyrotoxicosis induced by levothyroxine sodium.

### Materials and methods

Samples of whole dried Vitis vinifera L. were brought from Iraqi market in Baghdad, then leaves of the plant were isolated and kept in airtight glass containers till the time of the

experiment. Then the dried plant leaves were ground to fine powder. 25 gm of the powder, mixed with 250 mL of distilled water and were incubated for 3hrs at (60)°C then incubated overnight at room temperature. Suspension was then filtered. Water extracts were prepared daily just before administration orally to the experimental animals in a dose of (5 mL rabbit of (1.5-2)kg) .

### Preparation of levothyroxine sodium:

Fresh solution of levothyroxine sodium was prepared (tablet dissolve in water or food) just before feeding. For the animals given the levothyroxine sodium (50µg/each rabbit).

### Experimental animals: Eighteen

female oryctolagus cuniculus rabbit (1.5-2 Kg. each) were kindly supplied by city of Medicine, for the period from September 2012 to May 2013. And were used in this research. Rabbit were maintained with free access to water and diet (containing multivitamins, vegetables, and wheat). Experimental animals were divided into two groups (9 rabbit each):

- 1- Control group: rabbit were orally administered (using a feeding solution) with a daily dose of 5 mL distilled water for 2 months;
- 2- Plant- treated levothyroxine sodium group: : 50µg of levothyroxine sodium was orally administered daily to each rabbit in this group for 1 month, then 5 mL of the plant extract (100 mg/mL) was orally administered daily to each rabbit in this group for 2 month.

Blood sampling: Blood samples were collected from the heart of rabbit using heparinized capillary tubes. Serum was separated from blood samples, then frozen until used.

The activity of enzymes (LDH, ACP, and ALP) in the serum were determined according to method of King J(1965), Bergmeh H. U, et al(1974) and Roy A.V. (1970) respectively (10-12).

Statistical analysis: All statistical analysis of the study were done using SPSS version 15.0 for Windows (statistical Package for Social Science, Inc., Chicago, IL, USA).

Descriptive analysis was used to show the mean ± standard deviation of variables. The significance of difference between mean values was estimated by Student T- test. The probability  $p < 0.05$  = significant.

### Results and discussion

Results presented in table 1 indicate that the levels of diagnostic marker enzymes namely LDH, ACP, and ALP levels were significantly ( $p < 0.001$ ) increased in levothyroxine sodium treated femals of rabbit when compared with normal rabbit. However, treatment of rabbit female with Vitis vinifera L. significantly decreased ( $p < 0.001$ ) diagnostic marker enzymes like LDH, ACP, and ALP levels when compared to L. thyroxine sodium treated rabbit females.

**Table(1): Illustrate levels of LDH, ACP and ALP in blood rabbit females.**

	Control C	L.Thyroxine sodium G1	L.Thyroxine sodium +Plant extract 1 month G2	L.Thyroxine sodium +Plant extract 2 month G3	p-value				
	Mean±SEM	Mean±SEM	Mean±SEM	Mean±SEM	G1vs.C	G2vs.C	G3vs.C		G1 vs.G2
LDH U/L	147.8±2.02	173.5 ± 2.03	160.4± 0.44	150.3 ± 0.53	0.0001**	0.0001**		0.001*	0.0001**
ACP U/L	6.22±0.11	7.34± 0.11	6.52± 0.12	4.9 ± 0.18	0.0001**	N.S		0.0001**	0.001*
ALP U/L	17.10±0.53	22.2±0.49	20.1± 0.38	8.7±0.33	0.0001**	0.0001**		0.0001**	0.05*

SEM: Mean standard error, \*\* P=<0.0001 (highly significant), \*P = <0.05(significant), P=>0.05(N.S=No. significant)

The liver is the chief organ for metabolism of carbohydrates, lipids, proteins and detoxification of drugs and toxins. However, drugs affect the liver more commonly than any other organ and place the liver at a great risk of toxic damage[13]. After absorption of drugs by the intestine, it reaches the liver via the portal system. Liver disease remains as one of the serious health problems throughout the world[14].

The release of too much thyroxine in the bloodstream is known as thyrotoxicosis. This may be caused by over activity of the thyroid gland (hyperthyroidism), as in Graves' disease, inflammation of the thyroid or a benign tumour. Thyrotoxicosis can be recognized by a goiter which is a swelling of the neck due to enlargement of the thyroid gland. increased bowel movements, irregular menstrual cycle rapid or irregular heartbeat, palpitation, tiredness, irritability, tremor, hair loss and retraction of the eyelids resulting in a staring appearance[15]. Normal circulating levels of thyroid hormones are required for both, normal hepatic function and normal bilirubin metabolism[16].

The diagnosis of thyrotoxicosis is based on clinical symptoms, and characteristic pattern of changes in some serum enzymes such as LDH, ACP, and ALP[17]. LDH is the a cytosolic enzyme mainly present in periportal hepatocytes and released when the cells are lysed by hepatotoxin. The amount of enzyme released is proportional to the extent of damage caused to the cell [18]. Different forms of acid phosphatase are found in different organs such as spleen, kidney, pancreas, liver, bone, and prostate gland [19]. ALP is present in number of tissue including liver,

bone, intestine, and placenta [20]. The amount of enzymes released depends on the degrees or cellular damage, the intracellular concentration of the enzyme and the mass of affected tissue. The cause of the damage the enzyme released reflects the severity of the damage[21].

In other meaning the elevated levels of the LDH, ACP, and ALP in the rabbit females administered with levothyroxine sodium indicate the hepatocellular damage and alteration in the membrane permeability[22]. Our results are consistent with earlier studies, which strongly suggest that *Vitis vinifera* L. may protect the structural integrity of hepatocytes and prevent the leakage of cytosolic enzymes into bloodstream[23]. In addition *Vitis vinifera* L. leaf proved to be effective in reducing the extent of hepatocellular damage may be due to the presence of therapeutic phytochemicals such as proanthocyanidins and natural polyphenolic. Our finding suggest hyperthyroidism (also known as thyrotoxicosis overactive thyroid ) frequently leads to liver dysfunction[24].

#### Conclusion:

The present study concludes that the aqueous leaf extract of *Vitis vinifera* L. shows a therapeutic effect against levothyroxine sodium induced thyrotoxicosis in experimental female of rabbit and levels of LDH, ACP, and ALP are reduced which may be due to presence of bioactive compounds such as flavonoids, polyphenols, anthocyanins, proanthocyanidins, procyanidine and resveratrol.

## REFERENCE

- Almanza M. Pedro J. Quijano R. and et al. (2010). Physicochemical characterization of 'Point Noir' grapevine (*Vitis vinifera* L.) fruit during its growth and development under high altitude tropical conditions, *Agronomia Colombiana*, 38(2): 173-180. | 2- Mesely K. M. Slem A. and Talaat Z. (2012). Phytochemical and Biological Investigation of *Vitis vinifera* L. (Flame cultivar), Family Vitaceae Cultivated in Egypt, *Nature and Science*, 10(10): 48-59. | 3- Leelavinothan P. and Arumugam S. (2008). Effect of grape (*Vitis vinifera* L.) leaf extract on alcohol induced oxidative stress in rats, *Food and Chemical Toxicology*, 46(5): 1627-1634. | 4- Jean-Frederic T. Laurent B. Sarah I. Thierry P. Isabel F. and et al. (2010). Evolution and history of grapevine (*Vitis vinifera* L.) under domestication: new morphometric perspectives to understand seed domestication syndrome and reveal origins of ancient European cultivars, *Annals of Botany*, 105: 443-455. | 5- Mohammed K.Gh. Mojdeh N.H. and Akbar H. (2005). Vasorelaxatory Effect of *Vitis vinifera* Extract on Rat Aorta, *Iranian Journal of Pharmaceutical Research*, 2:93-99. | 6- Naik R. S. Mujumdar A. M. and Ghaskabdi S. (2004). Protection of liver cells from ethanol cytotoxicity by curcumin in liver slice culture in vitro, *Journal of Ethnopharmacology*, 95:31-37. | 7- Agoreyo B. O. (2010). Acid phosphatase and alkaline phosphatase activities in ripening fruit of *Musa Paradisiaca* L., *Omics Journal*, 3(3): 66-69. | 8- Ganjewala D. Nagaraja C. Nayak MR. and Devi SA. (2010). Effects of sodium nitroprusside on activity of acid and alkaline phosphatase and alkaline phosphatase in lemongrass (*Cymbopogon flexuosus* Steud) Wats, *International Journal of Plant Biology*, 1: 9-12. | 9- Djelic N. Nestic I. Stanimirovic Z. and Jovanovic S. (2007). Evaluation of the genotoxic effect of thyroxine using in vivo cytogenetic test on swiss albino mice, *Acta Veterinaria (Beograd)*, 57(5-6):487-495. | 10- Stentz R. et al. (2010). Controlled protein release from viable *Lactococcus lactis* cells. *Appl. Environ. Microbiol.* 76(9): 3026-3031. | 11-King J. The dehydrogenases or oxidoreductase. Lactate dehydrogenase. In: Nostrand, Van (Ed) *Partical Clinical Enzymology*, London,(1965); 106. | 12-Bergmeyer H.U. et al., in *Methods of Enzymatic Analysis*, Volume 1, 2nd ed., Academic Press, Inc.,(New York, NY: 1974) pp.495-496 | 13-Roy A.V. (1970). Rapid method for determining alkaline phosphatase activity in serum with thymolphthalein monophosphate, *Clin. Chem.* 16(5): 431-436. | 14-1. Kim, H.J. et al (2006). Glucocorticoids suppress bone formation via the Osteoclast, *J. Clin. Invest.* 116:2152-2160. | 15- Swapna S. Sowjanya P. Srinivasa R.N. Govinda G. and Giri BN. (2013). Effect of *Vitis vinifera* L. Seed Extract on Hepatic Marker Enzymes and Oxidative Stress against Acetaminophen Induced Hepatotoxicity in Rats, *International Journal Of Pharmaceutical And Chemical Sciences*, 2(2):2013. | 16- Salbab M. and Canelo R.(2011). An overview of evidence-based management of hepatocellular carcinoma: a metaanalysis, *Journal cancer Research Ther*, 7(4):463-475. | 17- NHS Choices, (2010). Thyroid, overactive [online]. Available from: www.nhs.uk/condition/Thyroid-overactive/pages/Introduction.aspx?url=Pages/What-is-it.aspx [Accessed 12 January 2010]. | 18- Cooper DS. (2001). Subclinical hypothyroidism, *N Engl J Journal Medical*, 1(345):260-5. | 19- Velavan S. Aegil Land Gokulakrishnan K. (2008). Protective effect of *Vitis vinifera* against Myocardial Ischemia Induced By Isoproterenol in Rats, *Pharmacologyonline*, 3: 958-967. | 20- Hazem M. M. (2012). Hepatoprotective Effect of Red Grape Seed Extracts Against Ethanol- Induced Cytotoxicity, *Global Journal of Biotechnology and Biochemistry*, 7(2): 30-37. | 21- Turner WL. and Plaxton WC. (2001). Purification and Characterization of banana fruit acid phosphatase. *Planta*, 214: 243-249. | 22- Hassan MQ. Tare R. Lee SH. and et al. (2007). HOXA 10: Controls Osteoblastogenesis by directly activating bone regulatory and phenolic genes, *Mol Cell Biol*, 27:3337-3352. | 23-Orhan DD. Orhan N. Ergun E. and Ergun F. (2007). Hepatoprotective effect of *Vitis vinifera* L. leaves on carbon tetrachloride- induced acute liver damage in rats, *Journal Ethnopharmacol*, 112(1): 145-151. | 24- Tenore GC. Manfra M. Stiuso P. Coppola L. Russo M. Gomez Monterrey IM and Campiglia P.(2012). Antioxidant profile and in vitro cardiac radical-scavenging versus pro-oxidant effects of commercial red grape juices(*Vitis vinifera* L. cv. Aglianico N.). *J Agric Food Chem*, 60(38):9680. | 25- Shin MO. And Moon JO. (2010). Effect of dietary supplementation of grape skin and seeds on liver fibrosis induced by dimethylnitrosamine in rats, *Nut Res Pract*, 4(5):369-374. | 26- Mirgam Ch. Christian M. Jarden P. Jean- Jacques S. and Peter R. H. (2004). Changes in Liver Function correlate with the Improvement of Lipid Profile after Restoration of Euthyroidism in Patients with Subclinical Hypothyroidism, *EXCLI Journal*, 3(1-9):1611-2156. |