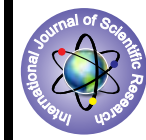


Prevalence and Antimicrobial Susceptibility of Pyogenic Microorganisms Isolated from Clinical Specimens of Patients of Lower Respiratory Infection



Medical Science

KEYWORDS : Antimicrobial susceptibility, Metallo- β -lactamase, Methicillin resistance

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ABSTRACT

Background : New emerging pathogens and associated antimicrobial resistance mechanisms have been observed in the respiratory tract of patients suffering from pyogenic infection in the last decade. Prevalence rate of metallo- β -lactamase (MBL)-producing *Pseudomonas aeruginosa* strains, Carbamapenase producing enterobacteriaceae, is growing, apart from prevalent ESBL positive *E.coli* & *Klebsiella pneumoniae*. **Methods :** In this study, 250 respiratory samples (Sputum) of were collected during a 24 months period. Microbiological cultures and antimicrobial susceptibility tests of the most frequently isolated bacteria were performed by standard disk diffusion method as per CLSI guidelines. **Results :** Out of 250 sputum samples, 142 pathogenic bacteria were isolated and characterized. Among the bacteria, *Streptococcus pneumoniae*, *Staphylococcus aureus* (both MSSA & MRSA) & *P. aeruginosa* (Both MBL producing & Non MBL producing) showed highest prevalence. **Conclusions :** The detection of MBL-producing *P. aeruginosa* and MRSA in patients confirms that antimicrobial resistance patterns should be always kept under surveillance. Moreover hygiene regulations in pulmonary clinics should prevent a further spread of resistant bacterial strains.

Introduction

The main causes for the high morbidity and mortality in patients with recurrent respiratory infections are predisposing conditions like smoking, viral infection in throat, extreme of age etc. Impairment of the muco-ciliary transport and thus the cleaning function of the upper airway predisposes bacterial colonization which turn into invasion [1]. Colonization is often initiated by respiratory commensal & Later on, becomes more complex when *Pseudomonas aeruginosa* and other gram-negative bacilli invade [2], [3] and [4]. During the course of therapy, these gram negative bacilli develop resistant mutant to ongoing antibiotic therapy, which is more difficult to eradicate [1], [5], [6] and [7]. The rate of methicillin resistance in *S. aureus* is rising [10] and the proportion of ESBL positive Enterobacteriaceae & metallo- β -lactamase-producing *P. aeruginosa* strains is growing [11], [12], [13], [14] and [15]. A continuous microbiological surveillance of the pathogens and a monitoring of the resistance situation is therefore of utmost importance. The aim of this study was to describe the prevalence of bacterial microorganisms in different age groups and to evaluate the antimicrobial susceptibility patterns of the common bacterial pathogens in respiratory illness during a two-year period.

Materials and methods

Patients and samples

Total 250 sputum samples (173 male patients and 77 female patients) were collected at tertiary care hospital affiliated with medical college in western India. The median age of the patients was 18 years, the range of age 6–71 years.

Microbiological cultures

Early morning Sputum was collected from each patient after vigorous gargling with drinking water to wash out oral commensal in sterile universal container. Each patient was instructed to take due possible care that not mix saliva with sputum & expectorate sputum only after deep breathing followed by coughing. Each sample transported immediately to Bacteriology laboratory. In laboratory, each sputum was primarily assessed for its quality by physical examination. Sputum mixed with saliva or pure salivary samples were rejected. Only mucoid or muco-purulent or purulent sputum were accepted for further investigation, rest of the sputum were rejected & asked patients

to collect sputum on next day morning. Each sample were inoculated primarily on Nutrient agar, Blood agar, Chocolate agar & Mac conkey agar & incubated for overnight incubation at 37^o C. If any growth was observed on next day, identification done by biochemical reaction & Antibiotic susceptibility by disc diffusion method according to the CLSI (formerly NCCLS) criteria were performed. If growth was not observed on next day, further another overnight incubation done at 37^o C. In order to ruled out mycobacterial infection, each sputum were screened by ZN stain.

Results

A total of 250 sputum samples were collected during a 24-months period. Out of which, 142 strains were isolated and characterized. A sum total of 108 sputum samples showed growth of mix commensal flora of oral cavity. They were considered as a non-pathogenic organism for lower respiratory tract infection. With regard to the prevalence of bacteria in the cohort under study, 36.6% of the isolates were *Strep. pneumoniae*, 27.4% *Staph. aureus*, 20.5 % *P. aeruginosa*, 9.8 % *Klebsiella pneumoniae*, 2.9% *H. influenzae*, 2.9% *E.coli*. Out of 29 the *P. aeruginosa*, 9 isolates (6.4%) showed a mucoid phenotype, which are mainly from patients with cystic fibrosis. With regard to the small-colony variant phenotype (SCV), 2 out of 20 non mucoid strain of *P. aeruginosa* (10%) isolates had SCVs., in contrast no SCVs of *Staph. aureus* were detected (**Table 1**).

Table 1. Prevalence of bacterial species in sputum sample

Bacterial species	No. of isolates (%)
<i>Staphylococcus aureus</i>	39 (27.4%)
<i>Streptococcus pneumoniae</i>	52 (36.6 %)
<i>klebsiella pneumoniae</i>	14 (9.8 %)
<i>Pseudomonas aeruginosa</i> (Non mucoid isolates)	20 (14.1 %)
<i>Pseudomonas aeruginosa</i> (Mucoid isolates)	9 (6.4 %)
<i>Escherichia coli</i>	4 (2.8 %)
<i>Haemophilus influenzae</i>	4 (2.8 %)
Total	142

An analysis of the prevalence of the various bacterial species in different age groups showed a constant increase of rate for mucoid and non mucoid *P. aeruginosa*. No mucoid isolates were detected in patients in the age group of 6–10 years. In contrast, the highest rate for *H. influenzae* and *S. aureus* was observed in patients in the age groups of 6–10 years and 11–15 years, respectively. *Strep. Pneumonia* were observed mainly in middle

age group (20 – 50 years), considering them as community acquired pathogen. In our study, Mucoid *P. aeruginosa* isolates exhibited a much higher antimicrobial susceptibility than the non mucoid strains. There is no predilection of age for isolated *Klebsiella pneumoniae*. *E.coli* were commonly isolated in age group of 11 – 20 years. (Table 2)

Table 2. Bacterial isolates in different age groups

Age (years)	Non mucoid <i>P. aeruginosa</i>	Mucoid <i>P. aeruginosa</i>	<i>S. aureus</i>	<i>H. influenzae</i>	<i>Strep. pneumoniae</i>	<i>E.coli</i>	<i>Klebsiella pneumoniae</i>	TOTAL
6–10	0	0	3	4	3	1	3	14
11–20	0	0	20	0	9	3	4	36
21–50	5	3	10	0	27	0	2	47
51–71	15	6	6	0	13	0	5	45
TOTAL	20	9	39	4	52	4	14	142

Table 3. Rate of susceptibility (%) of isolates against different antibacterial agents

Antibiotics	Non mucoid <i>P. aeruginosa</i> (n = 20)	Mucoid <i>P. aeruginosa</i> (n = 9)	<i>S. aureus</i> (n = 39)	<i>H. influenzae</i> (n = 4)	<i>Strep. Pneumonia</i> (n = 52)	<i>E.coli</i> (n = 4)	<i>Klebsiella pneumoniae</i> (n = 14)
Piperacillin	0	3	19	3	21	0	1
Ceftazidime	12	9	4	Not tested	5	0	2
Cefotaxime	2	6	6	4	45	0	2
Meropenem	18	9	32	4	52	4	13
Colistin Sulphate	20	9	Not tested	4	Not tested	4	14
Penicillin G	Not tested	Not tested	4	0	6	Not tested	Not tested
Cefazoline	Not tested	Not tested	31	2	39	0	0
Amikacin	15	8	29	4	26	3	11
Ciprofloxacin	6	3	10	3	16	0	4
Vancomycin	Not tested	Not tested	39	Not tested	52	Not tested	Not tested
Amoxicillin + Clavulanic acid	Not tested	Not tested	30	4	46	4	10
Oxacillin	Not tested	Not tested	29	0	27	Not tested	Not tested

Discussion

Prevalence and antimicrobial susceptibility of microorganisms isolated from respiratory samples of patients of LRTI have been described in several studies [3],[17],[18],[19],[20]. Similar to results reported by others, chronic respiratory tract infection of patients is caused mainly by bacteria such as *Staph. aureus*, *P. aeruginosa* and *H. influenzae*. In concordance with a previously published report [4], we observed that during the first decade of life, *Staph. aureus* and *H. influenzae* are the most commonly isolated bacteria from sputum, whereas in the second and third decade these species become less frequent, concomitant with a constant increase rate for gram negative bacilli infection.

With regard to the antimicrobial susceptibility, methicillin resistant *Staph. aureus* (MRSA) occurred in 10 patients (25.4%) of our study. In contrast, MRSA prevalence is 19.8% [3]; 25.9% [17], 18% [19] and 6% [18] in various parts of the world. Presumably the differences in the MRSA rates in patients correlate with the general nosocomial prevalence of MRSA of various parts of the world.

Nosocomial outbreaks of metallo- β -lactamase (MBL)-producing *P. aeruginosa* have been already described in Japan [11],[14], in Italy [12], in Brasil [13] and in Canada [15]. However, to the best of our knowledge, we report for the first time the prevalence rate of MBL-producing *P. aeruginosa* in patients. A systematic screening of these multi resistant strains is very important for a correct antimicrobial treatment, since MBL-producing strains are resistant to β -lactam-antibiotics. In our study, only 33% of MBL-producing *P. aeruginosa* were susceptible to ciprofloxacin but the All of them were still susceptible to colistin, a resurrected second-line therapeutic agent. Meropenem was also resistant in 10 % of non mucoid strain of *P. aeruginosa*. One strain

of *Klebsiella pneumoniae* was also resistant to carbapenem. All strains of *E.coli* (4 out of 4) & 12 out of 14 strain of *Klebsiella pneumoniae* were resistant to 3rd generation of Cephalosporin which may be due to ESBL production. We had not observed vancomycin resistance in *Staph. aureus*. Total 35 strain of *staph. aureus* out of 39 strain, were resistant to penicillin G, which is most likely due to β -lactamase enzyme production by *staph. aureus*. Amikacin had shown mild to moderate degree of resistance in both gram positive & gram negative bacteria. So, amikacin is the best choice for institution of empirical antibiotic when clinician is not sure that probable pathogen is gram positive or gram negative.

In our study, mucoid *P. aeruginosa* isolates exhibited a much higher antimicrobial susceptibility than the non mucoid ones. Independently from the antimicrobial susceptibility, mucoid strains in respiratory samples represent a non favourable prognostic factor, since they produce exopolysaccharide/alginate and consequently are resistant to phagocytosis. Furthermore they play a much greater role in lung disease progression than non mucoid *P. aeruginosa* [1], [5], [6] and [7].

Conclusions

The prevalence of different phenotypes of microorganisms in sputum of patients with respiratory illness should be closely monitored. Moreover, antimicrobial resistance patterns should be kept under surveillance, especially the occurrence of methicillin resistance in *S. aureus*, ESBL production in Enterobacteriaceae and MBL production *P. aeruginosa* respectively. Hygiene regulations in pulmonary clinics should prevent a further spread of resistant bacterial strains, as antibacterial treatment options are limited in these patients.

REFERENCE

1. J.R. Govan, V. Deretic Microbial pathogenesis in cystic fibrosis: mucoid *Pseudomonas aeruginosa* and *Burkholderia cepacia* *Microbiol Rev*, 60 (1996), pp. 539–574 | | 2. L. Saiman, J. Siegel Infection control in cystic fibrosis *Clin Microbiol Rev*, 17 (2004), pp. 57–71 | | 3. J.L. Burns, J. Emerson, J.R. Stapp et al. Microbiology of sputum from patients at cystic fibrosis centers in the United States *Clin Infect Dis*, 27 (1998), pp. 158–163 | | 4. S. Rajan, L. Saiman Pulmonary infections in patients with cystic fibrosis *Semin Respir Infect*, 17 (2002), pp. 47–56 | | 5. P.S. Stewart, J.W. Costerton Antibiotic resistance of bacteria in biofilms *Lancet*, 358 (2001), pp. 135–138 | | 6. J.R. Govan Multidrug-resistant pulmonary infection in cystic fibrosis—what does ‘resistant’ mean *J Med Microbiol*, 55 (2006), pp. 1615–1617 | | 7. Z. Li, M.R. Kosorok, P.M. Farrell et al. Longitudinal development of mucoid *Pseudomonas aeruginosa* infection and lung disease progression in children with cystic fibrosis *JAMA*, 293 (2005), pp. 581–588 | | 8. S. Besier, C. Smaczny, C. von Mallinckrodt et al. Prevalence and clinical significance of *Staphylococcus aureus* small-colony variants in cystic fibrosis lung disease *J Clin Microbiol*, 45 (2007), pp. 168–172 | | 9. S. Haussler, I. Ziegler, A. Lottel et al. Highly adherent small-colony variants of *Pseudomonas aeruginosa* in cystic fibrosis lung infection *J Med Microbiol*, 52 (2003), pp. 295–301 | | 10. K. Nadesalingam, S.P. Conway, M. Denton Risk factors for acquisition of methicillin-resistant *Staphylococcus aureus* (MRSA) by patients with cystic fibrosis *J Cyst Fibros*, 4 (2005), pp. 49–52 | | 11. K. Senda, Y. Arakawa, S. Ichiyama et al. PCR detection of metallo-beta-lactamase gene (bla_{IMP}) in gram-negative rods resistant to broad-spectrum beta-lactams *J Clin Microbiol*, 34 (1996), pp. 2909–2913 | | 12. L. Lauretti, M.L. Riccio, A. Mazzariol et al. Cloning and characterization of bla_{VIM}, a new integron-borne metallo-beta-lactamase gene from a *Pseudomonas aeruginosa* clinical isolate *Antimicrob Agents Chemother*, 43 (1999), pp. 1584–1590 | | 13. A.C. Gales, L.C. Menezes, S. Silbert, H.S. Sader Dissemination in distinct Brazilian regions of an epidemic carbapenem-resistant *Pseudomonas aeruginosa* producing SPM metallo-beta-lactamase *J Antimicrob Chemother*, 52 (2003), pp. 699–702 | | 14. H. Nishio, M. Komatsu, N. Shibata et al. Metallo-beta-lactamase-producing gram-negative bacilli: laboratory-based surveillance in cooperation with 13 clinical laboratories in the Kinki region of Japan *J Clin Microbiol*, 42 (2004), pp. 5256–5263 | | 15. J.D. Pitout, B.L. Chow, D.B. Gregson, K.B. Laupland, S. Elsayed, D.L. Church Molecular epidemiology of metallo-(beta)-lactamase-producing *Pseudomonas aeruginosa* in the Calgary Health Region: emergence of VIM-2-producing isolates *J Clin Microbiol*, 45 (2007), pp. 294–298 | | 16. K. Lee, D. Yong, J.H. Yum et al. Evaluation of Etest MBL for detection of bla_{IMP}-1 and bla_{VIM}-2 allele-positive clinical isolates of *Pseudomonas* spp. and *Acinetobacter* spp *J Clin Microbiol*, 43 (2005), pp. 942–944 | | 17. M.M. Anzaudo, N.P. Busquets, S. Ronchi, C. Mayoral Isolated pathogen microorganisms in respiratory samples from children with cystic fibrosis *Rev Argent Microbiol*, 37 (2005), pp. 129–134 | | 18. K. Semczuk, H. Dmenska, D. Dzierzanowska, M. Kolodziejczyk, E. Gabinska, H. Zareba The analysis of the isolated microorganisms from the respiratory tract of cystic fibrosis patients treated in Children’s Memorial Health Institute 1999–2002 *Pneumonol Alergol Pol*, 73 (2005), pp. 41–47 | | 19. A.D. Garcia, A. Ibarra, F.C. Rodriguez, M. Casal Antimicrobial susceptibility of bacterial isolates from patients with cystic fibrosis *Rev Esp Quimioter*, 17 (2004), pp. 332–335 | | 20. Lambiase, V. Raia, M. Del Pezzo, A. Sepe, V. Carnovale, F. Rossano | Microbiology of airway disease in a cohort of patients with cystic fibrosis *BMC Infect Dis*, 6 (4) (2006) |