

## An Investigation on a New Tapeworm *Staphylocystis myotisae* n. sp, in Bat intestine from Aurangabad District, Maharashtra, India



### Zoology

**KEYWORDS :** *Staphylocystis myotisae* n. sp, Aurangabad, Bat intestine, Cestode, Parasite.

B.W. Sawarkar

Department of Zoology, G. S. Science, Arts and Commerce College, Khamgaon Dist. Buldana 444 312(M.S.)

### ABSTRACT

Seven worms were collected from intestine of a Bat, *Myotis mystacinus* near Aurangabad, Maharashtra, India. The worms are of small size, short, with scolex neck, immature, mature and gravid proglottid. The scolex is large globular in shape and measures  $0.234 \times 0.221 - 0.240$  in length and breadth. It is wider than neck region. It bears a rostellum; which is cap shaped broader at the anterior end and tapering at the base and measures  $0.141 \times 0.035 - 0.069$  in length and breadth. Rostellum is armed with 28 hooks in a single circle each row with 14 hooks, short in length, bifurcated; the small hooks, measure  $0.024 \times 0.002$  in length and breadth, while handle measures  $0.012 \times 0.002 - 0.003$  in length and breadth. The four suckers are of medium size; slightly overlap on each other, round and oval in shape and measures  $0.069 - 0.073$  in diameter and  $0.053 \times 0.050$  in length and breadth. The worm is fully described in this paper.

### Description

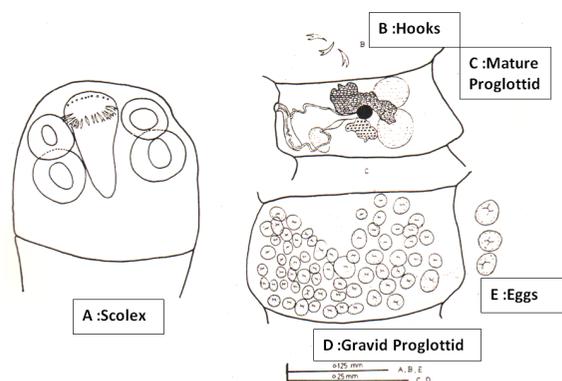
Seven worms were collected from intestine of a Bat, *Myotis mystacinus* in the month of April, 1984 at Aurangabad, Maharashtra, India. The worms are of small size, short, with scolex neck, immature, mature and gravid proglottid.

The scolex is large globular in shape and measures  $0.234 \times 0.221 - 0.240$  in length and breadth. It is wider than neck region. It bears a rostellum; which is cap shaped broader at the anterior end and tapering at the base and measures  $0.141 \times 0.035 - 0.069$  in length and breadth. Rostellum is armed with 28 hooks in a single circle each row with 14 hooks, short in length, bifurcated; the small hooks, measure  $0.024 \times 0.002$  in length and breadth, while handle measures  $0.012 \times 0.002 - 0.003$  in length and breadth. The four suckers are of medium size; slightly overlap on each other, round and oval in shape and measures  $0.069 - 0.073$  in diameter and  $0.053 \times 0.050$  in length and breadth.

The neck is short and measures  $0.071$  in length and  $0.206$  in breadth.

The mature proglottids are broader than long. Almost two times broader than long, narrow at the anterior end and broader towards the posterior side and measure  $0.148 - 0.172$  in length and  $0.316 - 0.37$  in breadth. Testes are three in number, almost round and oval in shape, two aporal in position, one poral in position and measure  $0.072$  in length and breadth. The two aporal testes are lateral to ovary while poral one is posterior in position and measure  $0.072$  in diameter and  $0.058 \times 0.036$  to  $0.074 \times 0.072$  in length and breadth. The two aporal testes are lateral to ovary, while poral one is posterior to ovary. The cirrus pouch is cylindrical, obliquely placed and narrow anteriorly and broader posteriorly and measures  $0.110 \times 0.012 - 0.024$  in length and breadth. It is situated at the lateral side of the proglottid and parallel to the margin of the same. It extends upto the posterior margin of the proglottid. The vas deferens is long, thin, coiled and measures  $0.185 \times 0.003$  in length and breadth. The vas deferens internally forms internal seminal vesicle, which is spindle shaped, transversely placed and measures  $0.041 \times 0.013$  in length and breadth. The cirrus is spiral coiled and measures  $0.129 \times 0.003$  in length and breadth.

The ovary is bilobed, having lobed margin situated in anterior half and near the anterior margin of the proglottid and measures  $0.147 \times 0.03 - 0.069$  in length and breadth. The vagina is a thin coiled tube, anterior to cirrus pouch and posterior to ovary which measures  $0.249 \times 0.003$  in length and breadth. The vagina opens in to a round, small, middle and posterior to ovary placed ootype and measures  $0.032$  in diameter. The vitelline gland is irregular, lobed, situated posterior to the ootype and posterior side of the proglottid and measures  $0.039 \times 0.060$  in length and breadth



**Fig.1. *Staphylocystis myotisae* n. sp. (A: Scolex B: Hooks, C: Mature segment D: Gravid segment, E: Eggs)**

The genital pores are unilateral, at one third from anterior lateral margin of the proglottid, small in size, oval in shape and measure  $0.018 \times 0.006$  in length and breadth.

The gravid segment are broader than long, almost two times and measure  $0.266 \times 0.0369 - 0.0443$  in length and breadth. The uterus breaks up in to 70-75 egg capsules. The egg capsules are small and measures  $0.026 \times 0.025$  to  $0.034 \times 0.026$  in length and breadth respectively.

### DISCUSSION

The genus *Staphylocystis* is established by Villot in 1877, with its type species *S. pistillum*. Later on 20 species are added to this genus. Out of them *Staphylocystis acuta* (Rud, 1819) and *S. bacillaris* (Goeze, 1782) are transferred to the genus *Vampirolepis* Spassky, 1950, because of the large number of characteristically shaped, numerous (upto 50) Y-shaped rostellar hooks, with the handle comparatively long and the guard broad and equal to or slightly more or less as long as the blade and in the large size of strobula. Prokopic (1967) in his studies on *Staphylocystis muris-sylvatici* (Rud, 1819) from apodemus considered it a synonyms of *Crenata* (Goeae, 1982) *Hymenolepis asketus* Brooks and Mays, 1977 is transferred to *S. (staphylocystoides)*.

The genus *staphylocystis* was proposed by Villot (1877) for grape like clusters of *Cysticercoides* found in the millipede, *Glomeris limbata* Latreille. The *cysticercooids* are formed by successive branching and external proliferation of secondary ones. There is no external membrane enclosing the cluster (Ramsom, 1904, Wardle and McLeod, 1952) as distinct from *Coenurus* and *Echinococcus* with internal budding and external enclosing membrane. From these clusters of *cysticercooids*, Villot (1877 a) described *S. bilarius* from the malphigian tubules and later (1877 b) *S. micracanthus* from the fat bodies. In a subsequent paper (1877 c), he recognized *S. bacillaris* as the *cysticercooids* of

*Taenia scutigera* Dujardin, 1845 and *S.micracanthus* as *Taenia pastille* Desjardin, 1845.

Joyeux and Baer (1835) demonstrated experimentally that *S.micracanthus* Violet, 1877, develops in to *Hymenolepis* contained 320 recognised species. He (1941) prepared a key, acknowledged as not entirely reliable together with figures from the structure of the rostellar hooks of the species. Skrijabin and Mathevosyan (1940 reported from the original literature specific descriptions and figures of the species of *Hymenolepididae*, reported from Mammals.

Spassky (1950) pointed out that in as much as *Staphylocystis micracanthus* Villot 1877 is a synonyms of *Taenia pistillum* Dujardin, 1845, it shall be regarded as the genotype of *Staphylocystis* i.e. *S.pistillum* (Dujardin, 1845).

In this classification of the hymenolepidid cestodes of mammals, Spassky (1945) provided a description of the genus *Staphylocystis*. In it, he pointed out that the species are small tapeworms, but gave no indication of the number of proglottids, that might be considered in the category of small, as he used it.

Yamaguti (1959) divided the genus *Staphylocystis* villot, 1877 in the two sub-genera. The *Hymenolepis pistillum* (Dujardin, 1845) as type, contains 16 species, 19 of the same ones included by spassky (1945). The members of this subgenus include those species of the genus, with one testis poral to the vitelline gland and two aporal. The species listed by Yamaguti range from 0.6 long with as a few as 10-12 long and probably with proglottids *S.(S.) pistillum* (Dujardin, 1845) to 150 long and probably with hundreds of proglottids *S. (S.) bacillaris* (Goeze, 1782).

The subgenus *Staphylocystoides* Yamaguti, 1959 with *Hymenolepis sphenomorphus* Locker and Rausch 1952 as a type contain six species. The subgenus is characterized by two testes being poral to the vitelline gland and one aporal. They range in size from 0.475 long with 7-10 proglottids *S.(S.) parvissima* (Voge, 1953) to 20 long with 158 recognisable proglottids *S. (S.) sengeri* (Nailand, 1953). Yamaguti's (1958) description of the genus varies from that of the Spassky (1959) description of the genus varies from that of the Spassky (1954) in such a manner as to be in consistent with the species that Yamaguti included and stated that the species are the "small worms", as did Spassky, but added "proglottids not numerous" which statement Spassky's description does not included. These two statements by Yamaguti are contradictory, when *S.(S.) pastillum* and *S.(S.) bacillaris* are examined, with intermediaries of short and long length, are included in Yamaguti's description of the gravid uterus is "horse shoe shaped on a simple sac". Since both authors included some species with a horse shoe shaped uterus and others with sac shaped uterus, this change by Yamaguti appears long, Later on the following species are added to this genus which are valid and are as such-

1. *S.(S.) chrysochloridis* (Janicki, 1904) Spassky, 1950.
2. *S.(S.) dodecocanth* (Baer, 1925) Spassky, 1950.
3. *S.(S.) furcata* (Stied, 1862) Spassky, 1950.
4. *S.(S.) fullebani* (Hilmy, 1936) Spassky, 1950.
5. *S.(S.) loossi* (Hilmy, 1936) Hubscher, 1937.
6. *S.(S.) minutissima* (Meggitt, 1927) Hubscher, 1937.
7. *S.(S.) Scalaris* (Dujardin, 1845) Blanchard, 1893.
8. *S.(S.) Solitaria* Meggitt, 1927.
9. *S.(S.) Syrdariensis* (Skarbilovitsch, 1945) Spassky, 1950.
10. *S.(S.) tiara* (Dujardin, 1845) Spassky, 1950.
11. *S.(S.) toxometra* (Baer, 1932) Skrijabin and Mathev, 1948.
12. *S.(S.) evansi* (Skrj et Mathev, 1942) Evans, 1940.
13. *S.(S.) longi* Oswald, 1951.
14. *S.(S.) parvissima* Voge, 1951.
15. *S.(S.) sengeri* Neiland, 1953.
16. *S.(S.) serrula* Oswald, 1951.
17. *S.(S.) sphenomorphus* Locker et Rausch, 1952.
18. *S.(S.) sanchorensis* Nama and khichi, 1975.
19. *S.(S.) shindensis*, Nama, 1976.
20. *S.(S.) suncusensis* Olsen and Kuntz, 1978.

In having testes in triangular arrangement and 28 hooks in a single circle, each row with 14 hooks, the worm under discussion comes closer to *S.(S.) sanchorensis* Nama and Khichi, 1975; *S. (S.) pistillum* Dujardin (1945); *S.(S.) syrdariensis*, Skarbilovitsch, 1946 and *S.(S.) suncusensis* Olsen and Kuntz, 1978.

1. The present cestode in having all the characters described earlier, differs from *S.(S.)sanchorensis* which is having 30 rostellar hooks, 15-17 long, strobula upto 10.77 long and in *Suncus*.
2. The present worm differs form *S.(S.) pistillum* which is having rostellar hooks 10 long, 14-22 in number, wrench shaped, strobula 0.02-2 and 0.65-2 long, scolex 120-125, uterus horse shoe shaped, eggs 51 x 46 and 57-46 and scolex.
3. The present worm differs from *S.(S.) syrdariensis* which is having rostellar hooks 20-30 in number, 20 long, strobula 15 long and eggs 43 x 32 and in *pidisprellus*.
4. The present worm differs from *S.(S.) suncusensis* which is having rostellar hooks 11-14 in number, 16-18 long, strobula 6.4-18.2 long, scolex 197-239 in diameter and in *Suncus*.

The above noted characters necessitate the erection of a new species to accommodate the present form and the name *S.(S.) myotisae* n.sp. is proposed after the generic name of the host.

## REFERENCE

- Binkienė, R., Kontrimavichus, V. & Hoberg, E.P. (2011) Overview of the cestode fauna of European shrews of the genus *Sorex* with comments on the fauna in *Neomys* and *Crocidura* and an exploration of historical processes in post-glacial Europe. *Helminthologia*, 48, 207–228. | <http://dx.doi.org/10.2478/s11687-011-0031-5> | Cheke A. (2011) Sonnerat's shrew - evidence for a new and possibly extinct species in an early 19th century manuscript (Mammalia: Soricidae). *Journal of the Bombay Natural History Society*, 108, 95–97. | Dubey, S., Salamin, N., Ruedi, M., Barrière, P., Colyn M. & Vogel, P. (2008) Biogeographic origin and radiation of the Old World crocidurine shrews (Mammalia: Soricidae) inferred from mitochondrial and nuclear genes. *Molecular Phylogenetics and Evolution*, 48, 953–963. | <http://dx.doi.org/10.1016/j.ympev.2008.07.002> | Eltyshov, Y.A. (1975) [Helminth fauna of mammals in the Barguzin basin and its geographical analysis. I. Systematic survey of helminths]. In: Parasitic organisms of north-east of Asia. Publishing house of the Far-Eastern scientific center, Vladivostok, pp. 135–167. (In Russian) | Genov, T. (1984) [Helminths of insectivores and rodents in Bulgaria]. *Izdatelstvo na Bulgarskata Akademiya na Naukite*, Sofia, 348 pp. (In Bulgarian). | Greiman, S.E. & Tkach, V.V. (2012) Description and phylogenetic relationships of *Rodentolepis gnoskei* n. sp. (Cyclophyllidae: Hymenolepididae) from a shrew *Suncus varilla minor* in Malawi. *Parasitology International*, 61, 343–350. | <http://dx.doi.org/10.1016/j.parint.2012.01.003> | Gulyaev, V.D., Dokuchaev, N.E. & Kornienko, S.A. (2007) [The cestodes of the genus *Staphylocystis* Yamaguti, 1959 (Cestoda, Hymenolepididae) in shrews of Beringia]. *Vestnik Severo-Vostochnogo Nauchnogo Tsentra Dal'ne-Vostochnogo Otdeleniya RAN*, issue 4, 75–84. (In Russian) | Gulyaev, V.D., Dokuchaev, N.E., & Lykova, K.A. (2010) [Description of *Spasskylepis rauschi* sp. n. (Cestoda, Hymenolepididae) from shrews *Sorex* in Alaska]. *Vestnik Severo-Vostochnogo Nauchnogo Tsentra Dal'ne-Vostochnogo Otdeleniya RAN*, issue 2, 75–84. (In Russian) | Gulyaev, V.D. & Shakhmatova, V.I. (1990) [On the morphology of cestode *Staphylocystis sibirica* (Morozov, 1957) (Hymenolepididae)]. In: G. S. Zolotareno (Ed.), *Taxonomy of insects and helminths*. Nauka, Novosibirsk, pp. 8–11 (In Russian) | Hall, T.A. (1999) BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symposium Series*, 41, 95–98. | Haukiasalmi, V., Hardman, L.M., Foronda, P., Feliu, C., Laakkonen, J., Niemimaa, J., Lehtonen, J.T. & Henttonen, H. (2010) Systematic relationships of hymenolepidid cestodes of rodents and shrews inferred from sequences of 28S ribosomal RNA. *Zoologica Scripta*, 39, 6, 631–641. | <http://dx.doi.org/10.1111/j.1463-6409.2010.00444.x> | Hoberg, E.P., Galbreath, K.E., Cook, J.A., Kutz, S.J. & Polley, L. (2012) Northern host-parasite assemblages: History and biogeography on the borderlands of episodic climate and environmental transition. *Advances in Parasitology*, 79, 1–97. | Karpenko, S.V. (1984) [Two new species of hymenolepidids (Cestoda: Hymenolepididae) from the shrews of the Khabarovsk krai]. In: *Izvestiya Sibirskogo otdeleniya Akademii Nauk SSSR, seriya biologicheskikh nauk*, vypusk 3, Nauka, Novosibirsk, pp. 117–124. | Karpenko, S.V. (2004) Helminths of shrews in the Khabarovsk krai. Materials of the All-Russian scientific-practical conference dedicated to the 65th anniversary of the Khabarovsk krai. Khabarovsk, pp. 80–86. | Kinsella, J.M. (2007) Helminths of the vagrant shrew, *Sorex vagrans*, from western Montana, USA. *Acta Parasitologica*, 52, 151–155. | <http://dx.doi.org/10.2478/s11686-007-0021-4> | Kinsella, J.M. & Tkach, V.V. (2009) Checklist of helminth parasites of Soricomorpha (=Insectivora) of North America north of Mexico. *Zootaxa*, 1969, 36–58. | Kornienko, S.A., Zubova, O.A., Gulyaev, V.D. & Dokuchaev, N.E. (2008) [Cestodes of shrews on Kunashir Island]. In: K.V. Galaktionov & A.A. Dobrovolskij (Eds.), *Proceedings of the IV Congress of the Russian Society of Parasitologists*, Russian Academy of Sciences, Vol. 2, Lema, St. Petersburg, pp. 75–77. (In Russian) | Littlewood, D.T.J., Waeschenbach, A. & Nikolov, P.N. (2008). In search of mitochondrial markers for resolving the phylogeny of cyclophyllidean tapeworms (Platyhelminthes, Cestoda)—a test study with Davaineidae. *Acta Parasitologica*, 53, 133–144. | <http://dx.doi.org/10.2478/s11686-008-0029-4> | Locker, B. & Rausch, R. (1952) Some cestodes from Oregon shrews, with descriptions of four new species of *Hymenolepis* Weinland, 1858. *Journal of the Washington Academy of Sciences*, 42, 26–31. | Makarikov, A.A. & Gulyaev, V.D. (2009). [Pararodentolepis gen. n., - a new cestode genus from rodents and the description of *P. sinistra* sp. n. (Cyclophyllidae: Hymenolepididae)]. *Parazitologiya*, 43, 454–459. (In Russian) | Morozov, Y.F. (1957) [Three new hymenolepidids from pygmy shrew]. *Uchenie zapiski Gor'kovskogo Gosuderstvennogo Pedagogicheskogo Instituta*, 19, 35–42. (In Russian) | Novikov, M.V. (1995) Cestodes of shrews (Insectivora, Soricidae) from the Magadan region, north-east Siberia. *Acta Parasitologica*, 40, 37–42 | Rausch R. & Kuns, M.L. (1950) Studies on some North American shrew cestodes. *Journal of Parasitology*, 36, 433–438. | <http://dx.doi.org/10.2307/3273168> | Nolan, M.J. & T.H. Cribb. (2005) The use and implications of ribosomal DNA sequencing for the discrimination of digenean species. *Advances in Parasitology*, 60, 101–163. | [http://dx.doi.org/10.1016/s0065-308x\(05\)60002-4](http://dx.doi.org/10.1016/s0065-308x(05)60002-4) | Olson, P.D. & Tkach, V.V. (2005) Advances and trends in the molecular systematics of the parasitic platyhelminthes. *Advances in Parasitology*, 60, 165–243. | [http://dx.doi.org/10.1016/s0065-308x\(05\)60003-6](http://dx.doi.org/10.1016/s0065-308x(05)60003-6) | Sawada, I. & Kobayashi, S. (1994) Cestode parasites of some micromammals (Insectivora) from the adjacent area of Akademgorodok City, Southern Central Siberia and Northern Teletskoye Lake, Altai Region, Russia. *Proceedings of the Japanese Society of Systematic Zoology*, 52, 14–33. | Senger, C.M. (1955) Observations on cestodes of the genus *Hymenolepis* in North American shrews. *Journal of Parasitology*, 41, 167–170. | <http://dx.doi.org/10.2307/3273786> | Spassky, A.A. (1954) [Classification of Hymenolepididae from mammals]. *Trudy Gel'mintologicheskoy Laboratorii Akademii Nauk*, 7, 120–167. (In Russian) | Spasskii, A.A. (1959) [A more precise definition of the types of relative positions of the genitalia in the Hymenolepididae]. *Zoologicheskii Zhurnal*, 38, 31–37. (In Russian) | Stieda, L. (1862) Ein Beitrag zur Kenntniss der Tánien. *Archiv für Naturgeschichte*, Berlin, 1, 200–209. | Stiles, C.W. (1906) Illustrated key to the cestode parasites of man. *Bulletin of the Hygienic Laboratory of the U.S. Public Health and Marine-Hospital Service*, 25, 1–104. | Tkach, V. & Pawlowski, J. (1999) A new method of DNA extraction from the ethanol-fixed parasitic worms. *Acta Parasitologica*, 44, 147–148. | Tkach, V.V. & Velikanov, V.P. (1990) [A new cestode species (Cestoda, Hymenolepididae) from desert shrew]. In: *Novosti faunistiki i sistematiki*, Naukova dumka, Kiev, pp. 7–10 (In Russian). | Tkach, V.V. & Velikanov, V.P. (1991) *Pseudhymenolepis turkestanica* sp. n. (Cestoda: Hymenolepididae), a new cestode from shrews. *Annales de Parasitologie Humaine et Comparée*, 66, 54–56. | Vaucher, C. (1971) Les cestodes parasites des Soricidae d'Europe. *Etude anatomique, révision taxonomique et biologie*. *Revue Suisse de Zoologie*, 78, 1–113. | Velikanov, V.P. & Tkach, V.V. (1993) [New cestode species (Cestoda, Hymenolepididae) from desert shrew]. *Vestnik zoologii*, N 5, 3–11 (In Russian). | Villot, F.A. (1877) *Classification du règne animal*. *Revue des Sciences Naturelles*, Montpellier, 47, No. 6. | Voge, M. & Rausch, R. (1955) Occurrence and distribution of hymenolepidid cestodes in shrews. *Journal of Parasitology*, 41, 566–574. | <http://dx.doi.org/10.2307/3274136> | Yamaguti, S. (1959) *Systema Helminthum*, Vol. 2. The Cestodes of Vertebrates. Interscience Publishers, Inc., New York, 860 pp. |