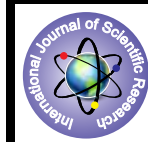


Down Syndrome: Association of Oxidative Stress and Thyroid Function With Haematologic Profile



Medical Science

KEYWORDS : Down syndrome(DS), Malondialdehyde(MDA), Catalase(CAT), Glutathione peroxidase(GPX), Uric acid (UA), Superoxide dismutase(SOD).

* DR.RESHMI.R

ASSISTANT PROFESSOR, DEPARTMENT OF PHYSIOLOGY, GOVERNMENT TD MEDICAL COLLEGE, VANDANAM, ALAPPUZHA, KERALA-688005
*CORRESPONDING AUTHOR

DR.VIJAYALEKSHMI
M.T

PROFESSOR AND HEAD, DEPARTMENT OF PHYSIOLOGY, GOVERNMENT TD MEDICAL COLLEGE, VANDANAM, ALAPPUZHA, KERALA-688005

DR.DINESH ROY .D

CEO & SENIOR CYTOGENETICIST, GENETIKA, CENTRE FOR ADVANCED GENETIC STUDIES, TRIVANDRUM, KERALA – 695024

DR.GEETHA
DAMODARAN

PROFESSOR, DEPARTMENT OF BIOCHEMISTRY, GOVERNMENT MEDICAL COLLEGE, KUMMANNUR, KONNI, KERALA-689691

ABSTRACT

Background:- Down syndrome (DS) or Trisomy 21 is the most common genetic cause of mental retardation. The phenotypic abnormalities and the morbidity in DS are due to higher gene dosage of chromosome 21, increased oxidative stress and thyroid dysfunction. **Aims:-** The association of oxidative stress and thyroid function with the haematological profile in DS was assessed. **Methods and Material:-** 25 cases of Down syndrome were selected for the study after confirmation by karyotyping. 6ml of blood was taken and sent for haematological analysis and serum analysis of Malondialdehyde (MDA), Catalase, Glutathione peroxidase, Uric acid, and Thyroid function. **Statistical analysis used:-** The mean and standard deviation of each of the variables were calculated. The association between the variables was calculated using Pearson's correlation. **Results:** DS cases showed significant increase of MCV, significant decrease of WBC count, RBC count, Platelet count, MCHC and neutropenia. The haematological variables showed a significant association with oxidative stress parameters. **Conclusion:** The haematologic profile in DS showed significant macrocytosis, anemia, leucopenia, neutropenia and thrombocytopenia. The increased oxidant stress in DS of this study, significantly altered the haematology of DS. Hence measures to decrease the oxidative stress may have a role in correcting the altered haematology profile in DS.

INTRODUCTION

Down syndrome (DS) or Trisomy 21 occurs in about 1:800 live births¹. The amplified gene products of chromosome 21 includes cystathionine beta synthase² and cytoplasmic Cu-Zn-superoxide dismutase (SOD)³ among 300 other genes. The SOD overactivity leads to oxidative stress affecting every system in DS by damaging DNA, lipids and proteins⁴. The haematological abnormalities in this progeroid, prooxidant state of DS are neutropenia, thrombocytopenia, macrocytosis, transient abnormal myelopoiesis, myelodysplasia and myeloid leukemia⁵. This case control study was done to assess the association of haematological profile with oxidative stress and thyroid status.

SUBJECTS AND METHODS

Karyotypically confirmed 25 cases of Down syndrome were selected for the study after getting informed consent from the parents. 25 age and sex matched controls were selected from the neighbourhood. 6 ml of blood was taken from the cases and controls. Ethical clearance was obtained from Institutional ethics committee (No.B6/14038/2011/EC66/2014).

Karyotyping was done by peripheral blood lymphocyte micro culture as described by Moorhead et al⁶, GTG banding as described by Seabright⁷ and chromosome identification according to International System for Human Cytogenetic Nomenclature.

Haematological parameters were measured using automated analyser. Malondialdehyde estimation (MDA) was done by Valipasha and Sadasivudu method⁸. Catalase activity was measured by the Hugo Aebi method⁹. Glutathione peroxidase estimation was measured by the decrease in absorbance at 340 nm on mixing the blood sample with Drab-

kins reagent and cumene hydroperoxide. Quantitative determination of uric acid was done using Trinder reaction¹⁰. Thyroid function was assessed using ELISA.

Statistical analysis was done using the SPSS Software¹⁷. The results were expressed as mean + Standard deviation. p values < 0.05 was considered significant. Pearson correlation coefficient (r) was used to test the correlation of different variables.

RESULTS

Table1: Haematological Profile of Down syndrome and controls

Variable	Controls/ cases	Mean	SD	t	p value
Hb g/dl	DS	13.4	1.6	-0.305	0.762
	Control	13.5	1.1		
MCV (fl)	DS	96.8	6.9	12.169	0.000
	Control	79.2	2.3		
RBC ($\times 10^6 / \mu\text{l}$)	DS	4.7	0.5	-2.753	0.008
	Control	5.0	0.3		
MCHC g/dl	DS	29.8	2.2	-7.046	0.000
	Control	33.8	1.8		
RDW (%)	DS	12.7	0.7	-3.826	0.000
	Control	13.5	0.9		
WBC ($\times 10^3 / \mu\text{l}$)	DS	2.9	1.2	-13.583	0.000
	Control	6.8	0.9		
Neutrophil (%)	DS	17.4	7.2	-14.503	0.000
	Control	47.4	7.5		
Lymphocyte count ($\times 10^3 / \mu\text{l}$)	DS	2.4	1.0	-3.807	0.000
	Control	3.3	0.6		
Platelet count ($\times 10^3 / \mu\text{l}$)	DS	142.2	50.7	-4.818	0.000
	Control	204	39.2		

It is evident from Table 1 that Down syndrome cases have significant macrocytosis, anemia, leucopenia,

lymphocytosis, neutropenia and thrombocytopenia.

Table 2: Values of Oxidative stress parameters , Antioxidant enzymes and Thyroid status in DS and controls

Variable Mean ±SD	MDA (nmol/L)	GPX (u/l)	Catalase (Kat/gm Hb)	Uric acid (mg/dl)	TSH μIU/ml	F T3 pmol/L	F T4 pmol/L
DS	120.0 ± 28.6	945.4 ± 324.1	102.9 ± 22.1	5.7 ± 1.5	3.6 ± 4.2	8.0 ± 2.2	11.4 ± 2.1
Controls	60.2 ± 6.6	5501.0 ± 1145.9	77.4 ± 0.6	3.3 ± 0.7	3.9 ± 2.6	5.5 ± 2.6	19.2 ± 4.2

Table 3: Correlation of Hematological parameters with parameters of oxidant stress and antioxidant activity. p<0.05 considered significant.

Pearson Correlation	MDA (nmol/l)		GPX (u/l)		CAT (Kat/gmHb)		UA (mg/dl)	
	r	p	r	p	r	p	r	p
MCV(fl)	0.720	0.000	-0.823	0.000	0.499	0.000	0.713	0.000
RBC count (× 106 /μl)	-0.345	0.014	0.361	0.010	-0.105	0.468	-0.088	0.543
MCHC(g/dl)	-0.633	0.000	0.767	0.000	-0.577	0.000	-0.456	0.001
RDW(%)	-0.339	0.016	0.469	0.001	-0.490	0.000	-0.384	0.006
WBC count (× 103 /μl)	-0.779	0.000	0.833	0.000	-0.595	0.000	-0.662	0.000
Neutrophil %	-0.782	0.000	0.858	0.000	-0.620	0.000	-0.714	0.000
Lymphocyte count (× 103 /μl)	-0.483	0.000	0.472	0.001	-0.333	0.018	-0.388	0.005
Platelet count (× 103 /μl)	-0.356	0.011	0.610	0.000	-0.453	0.001	-0.589	0.000

Results in Table 2 and Table 3 shows the following findings in Down syndrome. The WBC count has a significant negative correlation with MDA , GPX,CAT and UA values. RBC count has significant correlation with MDA,GPX and CAT values. MCV has a significant correlation with MDA,GPX,CAT and UA levels. Platelet count shows significant negative correlation with MDA ,UA ,CAT values and positive correlation with GPX values. Neutropenia shows significant negative correlation with MDA values,CAT and UA values and a significant positive correlation with GPX values.

Table 4: Correlation of Hematological parameters with thyroid status.

Pearson Correlation	FT3(pmol/l)		FT4(pmol/l)		TSH(μIU/ml)	
	r	p	r	p	r	p
WBC count(× 103 /μl)	-0.548	0.000	0.673	0.000	-0.041	0.779
RBC count(× 106 /μl)	-0.183	0.203	0.265	0.063	0.281	0.056
MCV(fl)	0.472	0.001	-0.684	0.000	-0.105	0.466
MCHC (g/dl)	-0.442	0.001	0.496	0.000	-0.013	0.930
Platelet count(× 103 /μl)	-0.327	0.020	0.556	0.000	-0.117	0.420
Lymphocyte %	0.423	0.002	-0.723	0.000	-0.004	0.977
Neutrophil %	-0.418	0.003	0.751	0.000	0.017	0.909
Lymphocyte count(× 103 /μl)	-0.264	0.064	0.394	0.005	-0.055	0.706
RDW%	-0.188	0.191	0.450	0.001	-0.384	0.006

Results in Table 4 shows that the haematological variables correlated significantly with FT3 and FT4 values, but failed to show any association with TSH values.

DISCUSSION.

In the present case control study, DS cases shows significant macrocytosis . The macrocytosis and reduced blood cell survival in DS has been documented in previous studies 11-14. The correlation of macrocytosis with oxidative stress in DS has not been documented before. This study shows significant correlation of macrocytosis with MDA values. In this study, the increase in MCV also correlated significantly with CAT and uric acid values , which has not been documented before.

studied in alcoholism 18 .In alcoholism, acetaldehyde and MDA form stable aldehyde-hemoglobin condensates .The site of haemoglobin that is modified by aldehyde is located near the erythrocyte membrane . Antibodies to acetaldehyde modified erythrocyte membrane proteins in peripheral blood and bone marrow aspirates brings about the morphological changes such as macrocytosis in chronic alcoholism 18 .Similar reason may be producing macrocytosis in Down syndrome .Moreover , such aldehyde bound RBC bring wide spread toxic effects due to oxidative stress 18.

Previous studies has been done on macrocytosis in Down syndrome and its association with Vit B12 and Folic acid

Correlation of macrocytosis with MDA values has been

levels^{12,15}. David et al in his study on DS concluded that macrocytosis could be due to altered folate remethylation pathway secondary to cystathionine beta synthase overactivity in ch21 as they could not find a significant association of macrocytosis with RBC folate or Vit B12 values¹². Nancy et al also could not find any significant association of macrocytosis with folate concentration in DS¹⁵.

The MCV, in this study did not show any significant association with TSH values even though it showed correlation with FT3 and FT4 values. Autoimmune hyper and hypothyroidism is common in Down syndrome and macrocytosis does occur in hypothyroidism according to earlier studies^{19,20}. The cause for macrocytosis in DS was postulated to be due to alteration in red cell membrane lipid distribution^{21,22} and also to vit B12 and folic acid deficiency, secondary to hypothyroidism²³.

Even though the Hemoglobin levels was decreased in DS, it was not statistically significant. But the RBC count was significantly decreased in DS and showed significant correlation with MDA, UA, and catalase. The negative correlation with MDA could be due to reduced red cell survival due to increased oxidative stress, causing accelerated aging of RBC as suggested by Wachtel et al¹¹ or due to autoimmune destruction of aldehyde bound cells as suggested by Latvala et al²⁴.

The negative correlation of RBC count with uric acid could be due to increased destruction of RBC producing increase in uric acid level as it is the final product of purine metabolism in humans. RBC count in this study did not show any association with Thyroid status.

The RDW indicates the degree of anisocytosis and it was significantly increased in the DS in this case control study. The RDW showed positive correlation with MDA values²⁵, Uric acid²⁶ as in other previous studies.

The significant Leucopenia, Neutropenia in DS in this case control study is in accordance with previous reports^{12,14-16}. The WBC count, lymphocyte count and neutrophil count showed significant correlation with oxidative stress markers, but failed to show any association with thyroid status. The role of aldehyde protein adducts as a cause for leucopenia in Down syndrome need to be studied in the upcoming research. The decreased count and function of leucocytes makes the DS susceptible for infection due to low immunity²⁷.

Platelet count was significantly decreased in DS and showed significant negative correlation with oxidative stress markers. This has been suggested due to autoimmune destruction of platelets by increased oxidative stress^{15,16}.

CONCLUSION

Macrocytosis, neutropenia and thrombocytopenia is significantly associated with Down syndrome in this case control study. The haematologic variables showed significant association with oxidative stress markers but failed to show a significant association with thyroid status. Hence as observed from this study and in the light of previous studies on oxidative stress and thyroid status, it should be assumed that the haematological abnormalities in DS is primarily due to significant oxidative stress. Macrocytosis may have a role as an index of oxidative stress in DS as well as in other disorders caused by oxidative stress, where other causes of macrocytosis has been ruled out.

REFERENCES

- Nussbaum LR, Mc Innes RR, Willard FH. Thompson and Thompson Genetics in medicine 6th ed, WB Saunders Philadelphia 2004: 157-159.
- Chadefaux B, Rethore MO, Raoul O, Ceballos L, Poissonnier M, Gilgenkranz S, Allard D. Cystathionine beta synthase gene dosage effect in trisomy 21. *Biochem Biophys Res Commun*. 1985;128:40-44.
- Sinet P. Metabolism of oxygen derivatives in Down syndrome. *Ann NY Acad Sci*. 1982;396:83-94.
- Roizen N, Patterson D. Down's Syndrome. *Lancet*. 2003;361:1281-9.
- John K. Choi. Haematopoietic disorders in Down syndrome. *Int J Clin Exp Pathol*. 2008;1:387-395.
- Moorehead PS, Nowell PC, Mellman WJ, Battipati DM, Hungerford DA. Chromosome preparations of leucocytes cultured from human peripheral blood. *Exp Cell Res*. 1960;20:613-6.
- Seabright M.A. rapid banding technique for human chromosomes. *Lancet*; 1971;2:971-972.
- Pasha, K.V., Sadasivadu, B. (1984) Intracellular content of thiol compounds, thiobarbituric acid reactive substances and gamma-glutamyl transpeptidase in rat brain during anoxia. *Neuroscience Letter* 46:209-214.
- Aebi H. Catalase in vitro. *Methods Enzymol*. 1984;105:121-6.
- Prencipe L, Fossati P, Vanzetti G. Enzymatic determination of uric acid in serum with the trinder reaction. *Quad Scavo Diagn*. 1978 Sep;15(3):382-94.
- Wachtel TJ, Poeschel SM. Macrocytosis in Down syndrome. *Am J Ment Retard*. 1991;95:417-420.
- David O, Fiorucci GC, Tosi MT, Altare F, Valori A, Saracco P. Haematological studies in children with Down SYNDROME. *diatr Hemat Oncol*. 1996;13:271-5.
- Ariel Tenenbaum, Sarah Malkiel, Isaiah D. Wexler, Floris Levy-Khademi, Shoshana Revel-Vilk, and Polina Stepensky, "Anemia in Children with Down Syndrome," *International Journal of Pediatrics*, vol. 2011, Article ID 813541, 5 pages, 2011. doi:10.1155/2011/813541.
- Kathryn Akin. Macrocytosis and leukopenia in Down's syndrome. *JAMA*. 1988;259:842.
- Nancy J, Roizen MD, Anthony P, Amarose. Hematologic abnormalities in children with down syndrome. *AJMG*. 2005.
- Onorata David, Giancarlo Fiorucci, Maria Teresa Tosi, Franco Altare, Alessandro Valori, Paola Saracco, Paola Asinardi. Haematological studies in children with Down syndrome. 1996;13:271-275.
- Tsantes AE, Bonovas S, Travlou A, Sitaras NM. Redox imbalance, macrocytosis and RBC homeostasis. *Antioxid Redox Signal*. 2006;8:1205-16.
- Mashketo Setshedi, Jack R Wands, Suzanne M de la Monte. Acetaldehyde adducts in alcoholic liver disease. *Oxid Med Cell Longev*. 2010;3:178-185.
- Omar S, Hadj Taeib S, Kanoun F, Hammami MB, Kamoun S, Ben Romdhane N, Feki M, Slimane H, Kaabachi N. Erythrocyte abnormalities in thyroid dysfunction. *Tunis Med*. 2010;88:783-8.
- Geetha J P, Srikrishna R. Role of red blood cell distribution width (RDW) in thyroid dysfunction. *Int J Biol Med Res*. 2012; 3(2):1476-1478.
- Montagnana M, Lippi G, Targher G, Salvagno GL and Guidi GC. The red blood cell distribution width is associated with serum levels of thyroid stimulating hormone in the general population. *Int J Lab Hematol*. 2009;31:581-2.
- Davidson RJ, Cumming AM, Leel VH, How J, Bewsher PD, Khir AS. A search for the mechanism underlying the altered MCV in thyroid dysfunction: a study of serum and red cell membrane lipids. *Scand J Haematol*. 1984;32(1):19-24.
- Antonijević N, Nesović M, Trbojević B, Milosević R. Anemia in hypothyroidism. *Med Pregl*. 1999;52:136-40.
- Jaana Latvala, Seppo Parkkila, Jukka Melkko, Onni Niemelä. Acetaldehyde Adducts in Blood and Bone Marrow of Patients With Ethanol-Induced Erythrocyte Abnormalities. *Molecular Medicine*. 2001; 7(6): 401-405.
- Zhiqiang Zhao, Tong Liu, Jian Li, Wansong Yang, Enzhao Liu, Guangping Li. Elevated red cell distribution width level is associated with oxidative stress and inflammation in a canine model of rapid atrial pacing. *International journal of cardiology*. 2014; 189:174-176.
- Min Luo, Zhan-Zhan Li, Yan-Yan Li, Li-Zhang Chen, Shi-Peng Yan, Peng Chen, Ying-Yun Hu. Relationship between red cell distribution width and serum uric acid in patients with untreated essential hypertension. *Scientific Reports*. 2014; 4: 7291.1-5.
- G Ram, J Chinen. Infections and immunodeficiency in Down syndrome. *Clin Exp Immunol*. 2011; 164(1): 9-16.