



SCREENING OF SOME WEEDS AGAINST SEED-BORNE FUNGI OF PIGEON PEA [CAJANUS CAJAN (L.) MILLSP.]

Botany

Jalander, V Department of Botany, Telangana University, Dichpally, Nizamabad (T.S.)

M. Mamatha Department of Botany, Telangana University, Dichpally, Nizamabad (T.S.)

ABSTRACT

In the present study leaf extracts from 15 weed plants such as *Acalypha indica* L., *Achyranthes aspera* L., *Alternanthera sessilis* (L.) R. Br. Ex. DC., *Cassia tora* L., *Croton bonplandianum* Baill., *Datura stramonium* L., *Euphorbia hirta* L., *E. heterophylla* L., *Hyptis suaveolens* (L.), *Parthenium hysterophorus* L., *Sida cordifolia* L., *Solanum xanthocarpum* Schrad. & H. Wendl., *Tephrosia purpuria* (L.) Pers., *Tridax procumbens* L. and *Xanthium strumarium* L. were tested for their antifungal activity (*in-vitro*) against seed-borne fungi of pigeon pea [*Cajanus cajan* (L.) Millsp.]. All the alcoholic leaf extracts showed good inhibitory activity against seed borne fungi and increased percent of seed germination, root & shoot length and vigour index. Alcoholic leaf extracts prepared from *X. strumarium* and *D. stramonium* leaves were showed good inhibitory activity against the seed borne fungi. 98% of seed germination was observed in seeds treated with alcoholic leaf extract of *X. strumarium*. The highest vigour index was recorded after the application of alcoholic and aqueous extracts prepared from *X. strumarium* and *D. stramonium* while the lowest was observed in case of untreated seeds of pigeon pea (control).

KEYWORDS

Antifungal activity, weed plants, seed-borne fungi, pigeon pea

Introduction

Pigeon pea [*Cajanus cajan* (L.) Mill. Sp.] also known as red gram or arhar, is one of the major legume crop, grown in the semi-arid tropics and most important food legume of India. Seeds are good vectors to transport plant pathogenic fungi (Agrawal and Sinclair, 1996). Seed is a primary source of plant growth and 90% of the food crops propagated by seed (Neergaard, 1977); seed is the most important unit of the crop; which should be of good quality and free from pathogen. Pathogen free and healthy seeds are used for sowing to achieve preferred germination, emergence, healthy seedlings and plant population. Fungal diseases are known to cause great damages all over the world. Seed borne diseases affect growth and productivity of crop plants (Kubiak and Korbas, 1999; Weber et. al., 2001). Different species of *Alternaria*, *Aspergillus*, *Curvularia*, *Drechslera*, *Fusarium*, *Penicillium*, *Rhizoctonia*, *Rhizopus* and *Trichoderma* are most common associates of seeds of pigeon pea (Jalander and Gachande, 2011). Seed-borne pathogens causes pre-and post-infections and considerable quality losses viz. seed abortion, seed rot, seed necrosis, reduction or elimination of germination capacity, seedling damage and their nutritive value have been reported (Miller, 1995; Janardhana et. al., 1998; Kavitha et. al., 2005). In recent years we are using very harmful chemicals to control pathogenic fungi. Using of such type of chemicals are environmentally unsafe. Using of plant extracts against plant pathogenic fungi is however, a recent approach to plant disease control. It helps to avoid environmental pollution by chemicals. Seed treatment is the safest and the cheapest way of control of seed-borne fungal diseases and to prevent biodeterioration of grains (Chandler, 2005; Bagga and Sharma, 2006). In the present study, the effect of leaf extracts prepared from different weed plants on seed borne fungi of pigeon pea seeds, seed germination, root & shoot length and vigour index were investigated.

Materials and Methods:

Collection of Plant Material: Fresh and healthy leaves of weed plants such as *Acalypha indica* L., *Achyranthes aspera* L., *Alternanthera sessilis* (L.) R. Br. Ex. DC., *Cassia tora* L., *Croton bonplandianum* Baill., *Datura stramonium* L., *Euphorbia hirta* L., *E. heterophylla* L., *Hyptis suaveolens* (L.), *Parthenium hysterophorus* L., *Sida cordifolia* L., *Solanum xanthocarpum* Schrad. & H. Wendl., *Tephrosia purpuria* (L.) Pers., *Tridax procumbens* L. and *Xanthium strumarium* L. growing in agriculture field near by the campus area and washed with tap water; surface sterilized with 0.01% mercuric chloride (HgCl₂) for 1 to 2 min and washed thoroughly 2-3 times with sterile distilled water then shade dried. Dried leaves were pulverized in electric blander to obtain fine powder.

Preparation of Plant Extracts:

Aqueous Extracts: To prepare aqueous leaf extracts, 20 g of powdered leaves were boiled in 100 ml of sterile distilled water for 10 min. Then extracts were filtered through double layered muslin cloth

and finally through Whatman filter paper No.1, the volume of the extracts were adjusted to 100 ml with sterile distilled water and preserved at 4°C in pre-sterilized flasks until use.

Ethanolic Extracts: To prepare ethanolic leaf extracts, 20 g of powdered leaves were boiled in 100 ml of 80% ethanol for 10 min. Then extracts were filtered through double layered muslin cloth and finally through Whatman filter paper No.1. Then the volume was adjusted to 100 ml with sterile distilled water and preserved at 4°C in pre-sterilized flasks until use.

Seed Treatment and Isolation of fungi: Seed samples of Pigeon pea were collected from local market and stored in jute bags for six months of period at room temperature (28±2°C). Hundred seeds of tested samples were immersed in aqueous and ethanolic leaf extracts of above mentioned weed plants and kept for overnight. Control was maintained with distilled water. The control and treated seeds were dried in shade. Seeds were inoculated and fungi were collected by using blotter method as described by International Seed Testing Association (Anon., 1966). Ten seeds were placed per Petri dish containing moist blotter paper. Plates were incubated at 28±2°C for 7 days. On incubation fungi were identified (Barnett and Hunter, 1972; Subramanian, 1971), root length and shoot length of the seedlings was recorded and percentage of seed germination and vigour index was calculated by the formulae.

% of seed germination = $\frac{\text{No. of seeds germinated in control} - \text{No. of seeds germinated in test}}{\text{No. of seeds germinated in control}} \times 100$
 Vigour Index = $\frac{\text{Mean of Root Length (cm)} + \text{Mean of Shoot Length (cm)}}{\text{Percentage of Seed Germination}}$

Results and Discussion:

Total 11 fungal species belonging to 7 genera were isolated and identified from seed samples of pigeon pea, five species belongs to *Aspergillus* i.e. *A. flavus*, *A. niger*, *A. nidulans*, *Aspergillus* sp.1, *Aspergillus* sp.2 and *Cladosporium herbarum*, *Curvularia lunata*, *Drechslera tetramera*, *Fusarium* sp. *Penicillium* sp. *Rhizopus stolonifer*. All the alcoholic leaf extracts showed good inhibitory activity against seed borne fungi and increased percent of seed germination, root & shoot length and vigour index. Alcoholic leaf extracts prepared from *X. strumarium* and *D. stramonium* leaves were showed good inhibitory activity against the seed borne fungi. 98% of seed germination, increased root length (7.482 cm) and shoot length (5.820 cm) was observed in seeds treated with alcoholic leaf extract of *X. strumarium*. The highest vigour index was recorded after the application of alcoholic and aqueous extracts prepared from *X. strumarium* and *D. stramonium* while the lowest was observed in case of untreated seeds of pigeon pea (control). Next to these, the alcoholic leaf extracts prepared from *E. hirta* and *C. tora* showed good inhibitory activity against the seed-borne fungi of pigeon pea. Aqueous leaf

extracts from weed plants also showed antifungal activity (less than the alcoholic extracts) against seed-borne fungi of pigeon pea. Similar results were observed in case of different crop plants by different workers; Kandhare (2015) studied effect of petroleum ether extract of different plant parts on seed mycoflora and seed health of green gram and he was observed that increase in seed germination and root length and shoot length. Kandhare (2016) also studied the effect of petroleum ether extract of different plant parts on seed mycoflora and seed health of Pigeon pea on blotter (after ten days of incubation) and reported that maximum stimulation in root length was observed in plant extract of *Cyperus rotundus* and shoot length was reported maximum in *Muntingia calabura*. Umer et.al. (2014) studied antifungal activity of 20 plant species was assessed against the fungus *Macrophomina phaseolina* and they were reported all the test plants inhibited the growth of *M. phaseolina* significantly to varying levels. Satish et. al. (2017) were studied aqueous extracts of 52 plants from different families for their antifungal potential against eight important species of *Aspergillus* isolated from sorghum, maize and paddy seed samples. Among fifty-two plants tested, aqueous extract of *Acacia nilotica*, *Achras zapota*, *Datura stramonium*, *Embllica officinalis*, *Eucalyptus globules*, *Lawsonia inermis*, *Mimusops elengi*, *Peltophorum pterocarpum*, *Polyalthia longifolia*, *Prosopis juliflora*, *Punica granatum* and *Syngium cumini* have recorded significant antifungal activity against *Aspergillus* species. Manoorkar et. al. (2015) reported antifungal activity of aqueous and ethanolic extracts of leaf & latex of *Calatropis procera* against ten seed-borne fungi isolated from oil seeds. Zakaria et.al. (2015) reported that ethanolic extracts of *Datura stramonium*, *Mentha longifolia* and *Malva parviflora* were effective against *Alternaria alternata*, *Botrytis cinerea* and *Penicillium italicum*.

Hasan et. al. (2005) studied the antifungal effect of plant extracts on seed borne fungi of wheat seed regarding seed germination seedling health and vigour index. They have been reported that all the water and alcoholic extracts gave good results against seed borne fungi of wheat seeds.

Table 1: Antifungal effect of aqueous leaf extracts prepared from different weeds on seed-borne fungi of pigeon pea [*Cajanus cajan* (L.) Millsp.]

S. No.	Name of plant	% of seed germination	Average Root length (cm)	Average Shoot length (cm)	Vigour index
1	<i>Acalypha indica</i>	58.00	4.628	3.000	442.42
2	<i>Achyranthes aspera</i>	56.00	4.512	2.712	404.54
3	<i>Alternanthera sessilis</i>	55.00	3.880	2.426	346.83
4	<i>Cassia tora</i>	60.00	4.926	3.226	489.12
5	<i>Croton bonplandianum</i>	52.00	3.240	1.526	247.83
6	<i>Datura stramonium</i>	70.00	5.212	3.562	614.18
7	<i>Euphorbia hirta</i>	62.00	4.942	3.420	518.44
8	<i>E. heterophylla</i>	58.00	4.642	3.120	450.19
9	<i>Hyptis suaveolens</i>	56.00	4.212	2.612	382.14
10	<i>Parthenium hysterophorus</i>	53.00	3.820	2.120	314.82
11	<i>Sida cordifolia</i>	58.00	4.812	3.020	454.25
12	<i>Solanum xanthocarpum</i>	55.00	3.982	2.524	357.83
13	<i>Tephrosia purpurea</i>	50.00	2.828	1.320	207.40
14	<i>Tridax procumbens</i>	58.00	4.620	3.000	441.96
15	<i>Xanthium strumarium</i>	75.00	5.812	4.200	750.90
16	Control	45.00	2.230	1.170	153.00

Table 2: Antifungal effect of alcoholic leaf extracts prepared from different weeds on seed-borne fungi of pigeon pea [*Cajanus cajan* (L.) Millsp.]

S. No.	Name of plant	% of seed germination	Average Root length (cm)	Average Shoot length (cm)	Vigour index
1	<i>Acalypha indica</i>	69.00	6.000	4.012	690.82
2	<i>Achyranthes aspera</i>	62.00	5.524	3.520	560.72
3	<i>Alternanthera sessilis</i>	65.00	5.812	3.824	626.34
4	<i>Cassia tora</i>	77.00	6.540	4.522	851.77
5	<i>Croton bonplandianum</i>	60.00	5.024	3.100	487.44
6	<i>Datura stramonium</i>	90.00	7.042	5.520	1130.58
7	<i>Euphorbia hirta</i>	78.00	6.624	4.822	892.78
8	<i>E. heterophylla</i>	71.00	6.214	4.212	740.24
9	<i>Hyptis suaveolens</i>	65.00	5.800	3.820	625.30
10	<i>Parthenium hysterophorus</i>	60.00	5.122	3.120	494.52
11	<i>Sida cordifolia</i>	69.00	6.100	4.000	696.90
12	<i>Solanum xanthocarpum</i>	65.00	5.752	3.862	624.91
13	<i>Tephrosia purpurea</i>	61.00	5.424	3.240	528.50
14	<i>Tridax procumbens</i>	70.00	6.110	4.012	707.84
15	<i>Xanthium strumarium</i>	98.00	7.482	5.820	1303.59
16	Control	45.00	2.230	1.170	153.00

REFERENCES

- Agrawal, V. K. and J. B. Sinclair 1996. Principles of Pathology. 2nd Edn. CRC Press, Inc., Boca Taton, FL, 539pp.
- Anonymous (1966). Proc. Int. Seed Test association. 31:1
- Bagga, P.S. Sharma V.K. (2006). Evaluation of fungicides as seedling treatment for controlling bakanae/foot-rot (Fusarium moniliforme) disease in basmati rice. J. Mycol. Plant Pathol., 59: 305-308.
- Barnett, H. L. and B. B. Hunter (1992). An Illustrated genera of imperfect fungi. 3rd Edn. Burgess Pub. Co., Minneapolis, Minnesota. 241pp.
- Chandler, J. 2005. Cost reduction in SIT programmes using exosect auto-dissemination as part of area wide integrated pest management. Int. J. Pest Control, 42(2): 257-260.
- Hasan, M. M., S.P. Chowdhury, Shahidul Alam, B. Hossain and M.S. Alam (2005). Antifungal effect of plant extracts on seed borne fungi of wheat seeds regarding seed germination, seedling health and vigour index. Pak. J. Biological Sci. 8(9)1284-1289.
- Jalander, V. and B. D. Gachande (2011). Seed-borne mycoflora of different varieties of pigeon pea [*Cajanus cajan* (L.) Mill. Sp.]. Bioinfollet, 8(2): 167-168.
- Janardhana, G.R., K.A. Raveesha and H.S. Shetty (1998) Modified atmosphere storage to prevent mould-induced nutritional loss in maize. J. Sci. Food Agric. 76(4): 573-578.
- Kandhare, A.S. (2016). Observation of plants for their antimicrobial and pro-pigeon pea growth activity. World J. Pharm. and Pharmaceutical Sci., 5(4) 1170-1175
- Kandhare, A.S. (2015). Effect of petroleum ether extract of different plant parts on seed mycoflora and seed health (seed germination, shoot length and root length) of Green gram (*Vigna radiata* L.) by blotter method. Int. J. of Life Sciences, A5: 100-104
- Kavitha, R., Umeha S., Shetty H.S. 2005. Dose dependent impact of dominant seed-borne fungi on seed germination and seedling vigour of cotton seeds. Seed Res. 33(2): 187-194.
- Kubiak K. and M. Korbas. 1999. Occurrence of fungal diseases on selected winter wheat cultivars. Postępy w Ochronie Roslin, 39(2):801-804.
- Manoorkar VB, Mandge SV and Gachande BD (2015) Antifungal activity of leaf and latex extracts of *calatropis procera* (ait.) against dominant seedborne storage fungi of some oil seeds. Bioscience discovery, 6(1):22-26.
- Miller, J.D. (1995) Fungi and mycotoxins in grain implications for stored product. Research. J. Stored Product Res., 31(1): 1-16.
- Neergaard, P (1977). Seed Pathology Vol. I and II McMillan Press Ltd. London: pp 1187.
- Satish, S., Mohana, D.C., Ranhavendra, M.P. and Raveesha, K.A. (2007). Antifungal activity of some plant extracts against important seed borne pathogens of *Aspergillus* sp. Journal of Agricultural Technology 3(1): 109-119.
- Subramanian, C. V. (1971). "Hypomyces an account of Indian species, except *Cercosporae*" ICAR Pub. New Delhi, 930p.
- Umer Iqbal, Tariq Mukhtar, Sheikh Muhammad Iqbal (2014) In vitro and in vivo evaluation of antifungal activities of some antagonistic plants against charcoal rot causing fungus *Macrophomina phaseolina*. Pak. J. Agri. Sci., 51(3), 689-694.
- Weber, R., B. Hrynyczuk, B. Runowska-Hrynyczuk and W. Kita 2001 Effects of tillage methods on the occurrence of culm base disease in several winter wheat cultivars. Electronic J. Polish Agric. Univ., Agronomy, 4(2): 6.
- Zakaria AM Baka, Mamdoh S Serag and Mohamed I Abodobra (2015) Antifungal activity of medicinal plant extracts against the predominant fungi causing spoilage of some fruits collected from markets of Egypt. Life sciences leaflets, 69:1-10.