



ANTIBACTERIAL ACTIVITY OF AQUEOUS EXTRACTS OF KENDU (*DIOSPYROS MELANOXYLON*) AND PUNARNAVA (*BOERHAVIA DIFFUSA*) LEAVES AGAINST URINARY TRACT INFECTIONS (UTI) CAUSING PATHOGENS.

Biotechnology

Kuldeep Prasad*

Research Scholar, Department of Microbiology, YBN University, Ranchi.
*Corresponding Author

Asha Mishra

Associate Professor, Department of Biotechnology, YBN University, Ranchi.

ABSTRACT

Urinary Tract Infection (UTI) is a common yet one of the serious health concerns afflicting women in India. It is caused by the pathogens that generally originate the digestive tract and get passed on to urethra and start to multiply. However, UTI can be caused by various other reasons, most prominent being lack of hygiene. The first line of treatment generally includes antibiotics that can sometimes prove to be ineffective due to cases of Antimicrobial Resistance (AMR) or Multi Drug Resistance (MDR). The situation creates a need to explore newer sources of drugs that can bring relief to affected population. The current study explores the antibacterial activity of Kendu (*Diospyros melanoxylon*) and Punarnava (*Boerhavia diffusa*) leaf extract against common UTI causing pathogens.

KEYWORDS

UTI, pathogens, antibacterial activity, antimicrobial resistance, etc.

INTRODUCTION:

Cases of Urinary Tract Infection (UTI) among women have seen a steady increase in India. Women are most affected by the UTI primarily because the length of urethra in women is shorter. Rate of infection among women is generally high due various reasons that include lack of hygiene, diabetes, accidental invasion of gut bacteria into urethra, and some more. UTI forms 25% of all the clinical bacterial infections that affect women. Reports suggest that up to 60% of women suffer from UTI once in their life time.

The front line treatment for UTI generally involves antibiotics like combination drug trimethoprim and sulfamethoxazole, fluoroquinolones, trimethoprim, β -lactams, nitrofurantoin, and fosfomycin tromethamine, ciprofloxacin, gentamicin (Jancel and Dudas, 2002, MacGowan and Albur, 2013). However, various independent studies have revealed that the efficacy of front line antibiotics has deteriorated due to Anti Microbial Resistance (AMR) among pathogens (Kothari and Sagar, 2008, Mohapatra *et al.*, 2022, Bhuiya *et al.*, 2025).

To counter the scenario of AMR UTI it is crucial to explore newer sources of antibacterial drugs. Certain plants have long been known to be medicinal with various health benefits. Current study aims to test the antibacterial activity of Kendu (*Diospyros melanoxylon Roxb*) and Punarnava (*Boerhavia diffusa*) leaf extract against common UTI causing bacteria like *E.coli*, *Klebsiella pneumonia*, and *Pseudomonas aeruginosa*.

MATERIALS AND METHODS:

Sample Collection And Processing: Kendu (*Diospyros melanoxylon Roxb*) and Punarnava (*Boerhavia diffusa*) leaf samples were collected from a location near Harli in Hazaribagh, Jharkhand (Geo-location: 23.824816, 85.274400 and 23.824030, 85.274726 respectively) and brought to laboratory. Punarnava and Kendu were chosen for this study due to their longstanding use in traditional medicine for treating a variety of health conditions, including infections. The leaves were cleaned with running tap water to remove dust and other impurities, followed by air drying indoor at room temperature (Kokate *et al.*, 2015). After drying, the leaves were finely ground into powder using a mortar and pestle. To ensure the integrity and efficacy of the powdered samples, they were stored in airtight containers to prevent moisture absorption and contamination.

Extract Preparation:

To prepare the hot water extracts, 10 grams of each powdered plant sample was meticulously dissolved in 100 ml of distilled water. The mixtures were carefully heated on a hot plate until they reached a vigorous boil, ensuring thorough extraction of the plant's bioactive compounds. Subsequently, the solutions were allowed to simmer gently for 30 minutes, facilitating the release and dissolution of phytochemicals into the solvent. After cooling to room temperature, the extracts underwent filtration using Whatman No. 1 filter paper to eliminate any solid particulates and obtain clear filtrates. To concentrate the extracted compounds, the filtrates were then subjected

to drying in a hot air oven set at 40°C. This drying process aimed to remove excess water while preserving the potency of the phytochemicals present in the extracts.

Extract was collected in a centrifuge tube and centrifuged at 8,000 rpm to remove the solid particles. The supernatant was collected in fresh centrifuge tube and concentrated using desiccator. The final yield of the extract was approximately 5.73% relative to the dry leaf powder.

Phytochemical Analysis:

The phytochemical composition of the Kendu and Punarnava aqueous leaf extracts were assessed separately. Standard qualitative methods were employed for the detection of alkaloids, flavonoids and phenolic phytochemicals (Harborne, 1998; Wagner and Bladt, 2001, Banu and Catherin, 2015): (a) alkaloid test: Wagner's Test - Kendu and Punarnava extracts were treated with a few drops of Wagner's Reagent to test the presence of alkaloid. (b) phenol test: Ferric Chloride Test - Kendu and Punarnava extracts were treated with a few drops of Ferric Chloride to test the presence of phenols, and, (c) flavonoids test: Alkaline Reagent test - Kendu and Punarnava extracts were treated with a few drops of Sodium Hydroxide and dilute Hydrochloric Acid to test the presence of flavonoids.

Sourcing Bacterial Isolates:

Urine samples were collected from female patients clinically diagnosed with urinary tract infections (UTIs) and processed to isolate the causative bacterial pathogens (Cheesbrough, 2006). Clinical samples were cultured on CLED agar and MacConky agar media. The plates were incubated at 37°C for 18-24 hours.

Biochemical Identification:

The isolated bacteria were identified using biochemical methods: Citrate test, Triple Sugar Iron (TSI) test, Urease test and Indole test (Godkar and Godkar, 2014).

Antibiotic Sensitivity Test:

Antibiotic sensitivity testing was performed to evaluate the susceptibility of the isolated UTI-causing bacteria (*E. coli*, *Klebsiella pneumoniae* and *Pseudomonas aeruginosa*) against the Kendu and Punarnava leaf extract. The Kirby-Bauer disc diffusion method was employed on Luria Bertani agar, a standard medium for antimicrobial susceptibility testing.

RESULTS:

Phytochemicals analysis of Kendu and Punarnava revealed significant presence of alkaloids, flavonoids and phenol group of phytochemicals (Table - 1).

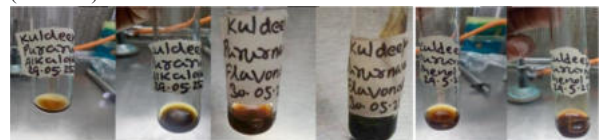


Figure 1 - Punarnava leaf extract phytochemical analysis – A:

Alkaloid test, B: Flavonoid test, C: Phenol test.



Figure 2 - Kendu leaf extract phytochemical analysis – A: Alkaloid test, B: Flavonoid test, C: Phenol test.

Phytochemical Compound	Kendu	Punarnava
	Presence (+/-)	
Alkaloids	+	+
Flavonoids	+	+
Phenols	+	+

On the basis of biochemical tests, the isolates were identified to be *E.coli* as the bacteria tested negative for Citrate and Urease tests and positive for Indole and TSI tests; *Klebsiella pneumoniae* was identified by positive Citrate and Urease tests, acidic pH and gas production in TSI test and negative for Indole test; and *Pseudomonas aeruginosa* was identified by characteristic positive tests for Citrate and TSI tests and negative for Urease and Indole tests. The identification criteria and test have been summarized in the Table – 2.

Bacteria	Citrate	TSI	Urease	Indole
<i>E. coli</i>	Negative	Positive/H ₂ S	Negative	Positive
<i>Klebsiella pneumoniae</i>	Positive	A/A, G	Positive	Negative
<i>Pseudomonas aeruginosa</i>	Positive	Positive	Negative	Negative



Figure 3 – Biochemical based identification of bacteria. A: *E. coli*, B: *Klebsiella pneumoniae*, C: *Pseudomonas aeruginosa*

Antibacterial Activity: Aqueous extract of Kendu and Punarnava leaf extracts significantly inhibited the growth of the three bacterial isolates (Table – 3).

Kendu		
Pathogen	Antibacterial activity	Inhibitory concentration (in mg)
<i>E. coli</i>	Positive	20
<i>Klebsiella pneumoniae</i>	Positive	20
<i>Pseudomonas aeruginosa</i>	Positive	20
Punarnava		
<i>E. coli</i>	Positive	0.1
<i>Klebsiella pneumoniae</i>	Positive	20
<i>Pseudomonas aeruginosa</i>	Positive	1.0



Figure 4 - Antibacterial activity of Punarnava leaf extracts against *E.coli* (A), *P. aeruginosa* (B) and *Klebsiella pneumoniae* (C).



Figure 5 - Antibacterial activity of Kendu leaf extracts against *E.coli* (A), *P. aeruginosa* (B) and *Klebsiella pneumoniae* (C).

DISCUSSION:

Kendu and Punarnava plants have long been known for various medicinal properties. However, their application as treatment options against Urinary Tract Infections (UTI) has been scantily explored. In this study aqueous extracts of Kendu and Punarnava leaves have shown significant antibacterial activity against UTI pathogens like *E. coli*, *Klebsiella pneumoniae* and *Pseudomonas aeruginosa* with Punarnava extract being more efficient than Kendu extract in inhibiting the growth of *E.coli* and *Pseudomonas aeruginosa*.

CONCLUSION:

The study clearly demonstrates the effective antibacterial activity of Kendu and Punarnava leaf extracts against common UTI causing pathogens *E. coli*, *Klebsiella pneumoniae* and *Pseudomonas aeruginosa*. However, more research needs to be done to determine the active bio-molecule responsible for antibacterial activity.

REFERENCES:

- Bhuiya, S., Kaushik, S., Logheeswaran, J., Karthika, P., Prathiviraj, R., Selvin, J., & Kiran, G. S. (2025). Emergence of Recurrent Urinary Tract Infection: Dissecting the mechanism of Antimicrobial Resistance, Host-Pathogen Interaction, and Hormonal Imbalance. *Microbial Pathogenesis*, 107698.
- Cheesbrough Monica. *District Laboratory Practice in Tropical Countries*. 2nd ed., Cambridge University Press, 2006.
- Jancel, T., and Dudas, V. (2002). Management of uncomplicated urinary tract infections. *The Western journal of medicine*, 176(1), 51–55. <https://doi.org/10.1136/ewjm.176.1.51>
- Kothari, A., and Sagar, V. (2008). Antibiotic resistance in pathogens causing community-acquired urinary tract infections in India: a multicenter study. *The Journal of Infection in Developing Countries*, 2(05), 354–358. <https://doi.org/10.3855/jidc.196>
- MacGowan, A., and Albur, M. (2013). Frontline antibiotic therapy. *Clinical medicine (London, England)*, 13(3), 263–268. <https://doi.org/10.7861/clinmedicine.13-3-263>
- Mohapatra, S., Panigrahy, R., Tak, V., J.V.S., K C, S., Chaudhuri, S., Pundir, S., Kocher, D., Gautam, H., Sood, S., Das, B. K., Kapil, A., Hari, P., Kumar, A., Kumari, R., Kalaivani, M., R, A., Salve, H. R., Malhotra, S., & Kant, S. (2022). Prevalence and resistance pattern of uropathogens from community settings of different regions: an experience from India. *Access microbiology*, 4(2), 000321. <https://doi.org/10.1099/acmi.0.000321>
- Godkar Praful B., and Darshan B. Godkar. *Textbook of Medical Laboratory Technology*. 2nd ed., Bhalani Publishing House, 2014.
- Harborne, A. J. (1998). *Phytochemical methods a guide to modern techniques of plant analysis*. Springer Science & Business Media.
- Wagner, H., & Bladt, S. (1996). *Plant drug analysis: a thin layer chromatography atlas*. Berlin, Heidelberg: Springer Berlin Heidelberg.
- Baru, K. S., & Cathrine, L. (2015). General techniques involved in phytochemical analysis. *International journal of advanced research in chemical science*, 2(4), 25-32.
- Kokate, C. K., Purohit, A. P., Gokhale, S. B. (2015). *Pharmacognosy*. 50th ed., Nirali Prakashan.