



Monitoring of foliar blight pathogens in wheat (*Triticum aestivum* L.)

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ABSTRACT

The wheat crop in eastern India, suffers from a number of fungal foliar diseases, namely, *Alternaria* leaf blight (*Alternaria tritricina*), *Curvularia* leaf spot (*Curvularia lunata*), Spot blotch (*Bipolaris sorokiniana*) and Tan spot (*Pyrenophora tritici-repentis*). Spot blotch covers over 70 per cent of the foliar blight incidence. Over the years the focus, as major foliar blight pathogen, is shifting to *Cochliobolus sativus* (Ito and Kurib.) Drech. Ex Dastur [anamorph: *Bipolaris sorokiniana* (Sacc.) Shoem. Thus, spot blotch caused by the pathogen is, logically, responsible for major losses attributed to foliar blights. Spot blotch are responsible for over two thirds of area coverage and much of the yield losses by these diseases. Foliar blight initiated by December end on foliage, January end on sheath and March beginning on neck and ear head of normal sown of wheat. Colony shape of foliar blight pathogens isolates from different parts of wheat were observed as uniformly circular with aerially produced mycelia.

KEYWORDS

Wheat, *Bipolaris sorokiniana*, *Alternaria tritricina*, *Curvularia lunata*, *Pyrenophora tritici-repentis*.

Wheat (*Triticum aestivum* L. emend. Fiori and Paol.) crop suffers from a number of devastating diseases caused by fungi, bacteria, viruses, mycoplasma, nematodes and environmental factors. Sixty-nine listing of fewer pathogens causing twenty-three diseases being more important than those caused by other groups of pathogens or factors. However, in eastern Uttar Pradesh only fungal diseases, such as black rust [*Puccinia graminis* f.sp. *tritici* (Pers) Eriks and Henn], brown rust [*Puccinia recondita* Rob. Ex. Desm), yellow rust (*Puccinia striiformis* West), foliar blights (*Helminthosporium stivum* Pamm. King & Bakke and *Alternaria tritricina* Prasada & Prabhu), loose smut [*Ustilago nuda* var. *tritici* (Jens) Schaf], karnal bunt [*Neovossia indica* (Mitra) Mundkur] and powdery mildew (*Erysiphe graminis* D C), are of major consequence. With the change in cropping system, cropping intensity, crop management and varietal spread, the foliar blights are causing major losses to wheat crop in eastern Uttar Pradesh. A number of pathogens causing leaf blight, blotches and spots on this crop have been reported. Foliar blights in wheat are caused by species of *Alternaria* and *Helminthosporium*. This disease caused by both of these pathogens is generally taken as a group because in most of cases it is rather difficult to distinguish one from the other field condition.

Wheat foliar blights have been studied extensively in India (Joshi *et al.*, 1986; Singh *et al.*, 1986 and Singh and Srivastava, 1997), but due to difficulty in distinguishing the symptoms at field level and isolation and identification of pathogens in laboratory (Goel *et al.*, 1999), many of the findings are suspect in nature and represent foliar blights in general, though attributed, most of the time, to a specific pathogen. Consequently work on epidemiology and on the components of integrated management of foliar blights, exclusively, has been scarcely attempted.

Material and method

Studies were conducted at the Student Instructional Farm (SIF), Main Experiment Station (MES), Net House, Glass House, Wheat Pathology Laboratory and other facilities at the Main Campus of N. D. University of Agriculture & Technology, Kumarganj, Faizabad (U.P.). It is located in the Indo-Gangetic plains of Eastern Uttar Pradesh at latitude 26.47°N, longitude 82.12° E and altitude 113 m above the sea level. Field trials were conducted during Rabi 2009-10 and 2010-11. Samples were closely examined using naked eyes and hand lens to selectively choose foliar blight affected one following the symptoms described by Misra (1973 a, b), Bazlur

Rashid *et al* (1987) and Anderson (1952). Sheath, leaf, neck and ear head showing typical symptoms were cut and arranged separately for isolating the fungi associated with the samples. The diseased leaf samples, showing distinct characteristics of foliar blights disease, were selected for isolation of the pathogen. The selected plant parts were washed with fresh sterilized water in order to remove the dust particles and surface contaminants. The washed diseased plant parts were cut into small bits, with some healthy portion. The cut plant parts were surface sterilized with 0.1 per cent mercuric chloride solution under aseptic condition inside a laminar flow and washed thoroughly 3 to 4 times with sterilized water to remove the traces of mercuric chloride. Excess moistures were removed by placing these in the fold of sterilized blotting papers. The pieces, thus sterilized, were transferred in petridishes previously sterilized at 160°C for two hours in an electric oven and poured with 2 per cent potato dextrose agar medium. The medium was previously autoclaved at 15 pound per square inch pressure for 20 minutes. Three to four pieces of diseased plant parts were placed per petridish properly marked with glass marking pencil indicating date of isolation and isolate number etc. The petridishes were then transferred at 30°C in an incubator.

Isolated and raised pure cultures of different fungi were identified on the basis of morphological and cultural characters (Gilman, 1967, 1975; Subramaniam, 1971; Ellis, 1971, 1976; Bernett and Barry, 1972; Malone and Musket, 1964 and Singh, 1983).

Result and Discussion

Field observations revealed presence of foliar blights in wheat crop by the end of December during both the years. Collection of samples start on 1st January and isolations made on 2 and 19 of January. In identification on 10 and 27 January produced *Bipolaris sorokiniana* (Sacc.) Shoem. and *Alternaria tritricina* Prasada and Prabhu (Table-1,2). Foliar blight pathogens were isolated from beginning (January, 01) to the end of the season (April, 02). February, 01 and 16 isolations yielded *Alternaria tritricina* and *Drechslera tritici-repentis* also, March, 03 isolations additionally, yielded *Alternaria alternata* also. *A. tritricina* and *A. alternata* too remained present till the end of season. Occurrence of one or more of these pathogens in the country and region, have been reported by Sokhi *et al.* (1972), Singh *et al* (2001a, b, c), Sharma *et al.* (2004a, b) and Singh *et al.* (2004c, e) also. Rare occurrence of *D. tritici-repentis* in the region is in conformity with its presence being

recorded by Mishra 1973a and Mishra (1973b) in Bihar, Singh *et al.* (2001b, c) in wider areas in very low proportion and absence by Mahto (2000a), Dubin and Van Ginkel (1991) and Nagarajan (2001) have also emphasized the fact that in South-Asia, the incidence of *B. sorokiniana* prevails over that of *D. tritici-repentis*.

Sheath showed apparent symptoms by the January end. February, 3rd to March, 3rd isolations, in both years, and March, 18 isolations only in 2009-10 yielded *B. sorokiniana* alone. Later isolations, March, 18 in 2010-11 and March, 18 and April, 02 in 2009-10 yielded *A. alternata* and *A. triticina* as well. This coincides with late season isolation of this fungus from leaves also. Neck and ear head symptoms appear in March beginning. April isolations, however,

yielded *Curvularia* spp. also. In addition April isolations yielded *A. alternata* as well. *B. sorokiniana* causing sheath, neck and ear head blight phase of spot blotch of wheat has been well documented in India (Mishra, 1973a and Mishra, 1973b) and abroad (Anderson, 1952). *B. sorokiniana* isolates obtained from wheat proved pathogenic to their host reported earlier also from this centre (Singh *et al.*, 1998b; Singh, 1999 and Singh *et al.*, 2001a,b,c). But the successful cross pathogenicity of the isolates from this region has never been reported earlier, such studies elsewhere in wheat isolation, showed the pathogen to appear as a continuum of isolates differing in aggressiveness as noted by Duveiller *et al.*, (2005).

Table-1 Fungi isolated from different infected parts (Sheath, Leaf, Neck and Ear head) of wheat plant during 2009-2010

Date of sampled collections	Date of isolation	Sheath	Leaf	Neck	Ear head
1 Jan.	3 Jan.	Healthy	<i>Bipolaris sorokiniana</i>	Healthy	Healthy
16 Jan.	19 Jan.	Healthy	<i>B. sorokiniana</i> <i>Alternaria triticina</i>	No heading	No heading
1 Feb.	3 Feb.	<i>Bipolaris sorokiniana</i>	<i>B. sorokiniana</i> <i>A. triticina</i> <i>A.alternata</i>	No heading	No heading
16 Feb.	18 Feb.	<i>B. sorokiniana</i> <i>A. triticina</i> <i>A.alternata</i>	<i>B. sorokiniana</i> <i>Alternaria tinussima</i> <i>D. tritici-repentis</i>	Infected sample not available	Infected sample not available
1 March	3 March	<i>B. sorokiniana</i> <i>A.alternata</i> <i>A. triticina</i>	<i>B. sorokiniana</i> <i>D. tritici-repentis</i> <i>A. triticina</i> <i>A. tinussima</i>	<i>B. sorokiniana</i>	<i>B. sorokiniana</i>
16 March	18 March	<i>B. sorokiniana</i> <i>A. triticina</i> <i>A.alternata</i> <i>Epicocum sp.</i>	<i>B. sorokiniana</i> <i>D.tritici-repentis</i> <i>A.alternata</i> <i>Epicocum sp.</i>	<i>B. sorokiniana</i> <i>A. triticina</i> <i>A.alternata</i>	<i>B. sorokiniana</i> <i>A. triticina</i> <i>Epicocum sp.</i>
1 April	3 April	<i>B. sorokiniana</i> <i>A.alternata</i> <i>Epicocum sp.</i> <i>A. triticina</i>	<i>B. sorokiniana</i> <i>D. tritici-repentis</i> <i>A. tinussima</i> <i>A. triticina</i>	<i>B. sorokiniana</i> <i>A. triticina</i> <i>C. lunata</i> <i>A.alternata</i>	<i>B. sorokiniana</i> <i>A. triticina</i> <i>C. lunata</i> <i>Epicocum sp.</i>

Table-2 Fungi isolated from different infected parts (Sheath, Leaf, Neck and Ear head) of wheat plant during 2010-2011

Date of sampled collections	Date of isolation	Sheath	Leaf	Neck	Ear head
1 Jan.	2 Jan.	Healthy	<i>Bipolaris sorokiniana</i>	Healthy	Healthy
16 Jan.	19 Jan.	Healthy	<i>B. sorokiniana</i> <i>A.alternata</i> + <i>A. triticina</i>	No heading	No heading
1 Feb.	3 Feb.	<i>Bipolaris sorokiniana</i>	<i>B. sorokiniana</i> <i>D. tritici-repentis</i> <i>A. triticina</i> + <i>A.alternata</i>	No heading	No heading
16 Feb.	18 Feb.	<i>Bipolaris sorokiniana</i>	<i>B. sorokiniana</i> <i>Alternaria tenuissima</i> <i>D. tritici-repentis</i>	Infected sample not available	<i>B. sorokiniana</i>
1 March	3 March	<i>B. sorokiniana</i> <i>A. triticina</i> <i>A.alternata</i>	<i>B. sorokiniana</i> <i>D.tritici-repentis</i> <i>A.alternata</i> + <i>A. triticina</i> , <i>A. tinussima</i>	<i>B. sorokiniana</i>	<i>B. sorokiniana</i> <i>A. triticina</i> <i>Epicocum sp.</i>
16 March	18 March	<i>B. sorokiniana</i> <i>A. triticina</i> , <i>A.alternata</i> <i>Epicocum sp.</i>	<i>B. sorokiniana</i> <i>D. tritici-repentis</i> <i>A.alternata</i> <i>Epicocum sp.</i>	<i>B. sorokiniana</i> <i>A. triticina</i> <i>A.alternata</i>	<i>B. sorokiniana</i> <i>A. triticina</i> <i>Epicocum sp.</i>
2April	2 April	<i>B. sorokiniana</i> <i>A.alternata</i> , <i>C. lunata</i> <i>Epicocum sp.</i>	<i>B. sorokiniana</i> <i>D. tritici-repentis</i> <i>A. triticina</i> , <i>A. tenuissima</i>	<i>B. sorokiniana</i> <i>A. triticina</i> <i>A.alternata</i> <i>C. lunata</i>	<i>B. sorokiniana</i> <i>A. triticina</i> <i>C. lunata</i> <i>Epicocum sp.</i>

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