PARIPEX - INDIAN JOURNAL OF RESEARCH | Volume-8 | Issue-12 | December - 2019 | PRINT ISSN No. 2250 - 1991 | DOI : 10.36106/paripex

| ournal or | |
|-----------|---|
| S S | - |
| | |
| | 9 |
| ARIPET | |

ORIGINAL RESEARCH PAPER

TISSUE CULTURE STUDIES CALLUS TREATMENT ON STEM NODE EXPLANTS OF CITRULLAS VALGARIS, L.

Botany

KEY WORDS:

 Mandaloju.
 Department Of Botany, Kakatiya University, Warangal – 506 009, Telangana

 Venkateshwarlu
 India.

The plant tissue culture methods also provide base for the improvement of crop to induce somaclonal variations, In Vitro mutations, genetic transformation of medicinally important genes and development of somatic hybrids plant regeneration protocol is required. Embryo genic callus induction and plant let proliferation of Solanum nigrum Venkateshwarlu M (2017). Shoot regeneration better than other cytokinins (Hussain et al 2007). Phyto chemical analysis of Solanum Surattense young leaves evalated for the presence of bioactive compounds using various polarity solvents petroleum (Venkateshwarlu et al 2018). The present study established reliable and reproducible protocol for rapid multiple shoot induction from node explants of Citrullas Valgaris, using different concentration and combination of cytokinins. Murashige and Skoog (1962) medium supplemented with 0.5 to NAA 2.0 mg/1 BAP was found to be optimum to induce shoots directly from the stem node explants. Since very scarce information is available about micro propagation of this important medicinal plant, an attempt was made to develop a reproducible protocol for multiple shoot induction form stem node explants of one the tissue culture. Several workers in past have micro propogated some of the important Asclepiadaceae members such as Cerogia bibosa (Britto et al., 2003), holostemma adakodien (Martin, 2002-2003). Significant increase in the number of shoots per explants was found ion M.S. medium supplemented with NAA, BAP and 14 mg/l adenine sulphate . All the tested combinations have little effect on increasing the number of shoots. Nodal explants derived shoot cultures were sub cultured to M.S. medium fortified with same concentration of hormone for shoot elongation. The percentage of explants exhibiting shoot induction was found to be between 50-60 i. Most of the concentrations of Benzyl amino purine tested except M.S medium supplemented with 0.5-2.0 mg/l benzyl amino purine. Stem segments are used as important explants for genetic transformation system, described in many plants species (Rastogia and Dwivedi, 2006)

INTRODUCTION:

ABSTRACT

The best results were obtained when stem node explants were initially cultured with (4.0mgl l BAP, 1.0mg/l NAA, 3.0mg/l) developed callus after 15-20 days of culture. In Vitro techniques using micro propagation and tissue culture offer a great possibility to overcome their problem. Micro Propagation using stem node explants. The auxin 2,4-D has been dermined as potent callus inducing phyto hormones in studies with many plant species. Various in the callus forming ability of different concentration of BAP, NAA and 24-D Table-I, Plate-I were placed in the medium to compare their growth responses. Among the tested NAA, Indeed the hight yield of callus followed by 2,4-D similarly. The callus so produced was green in colony and soft in texture presence of 2,4-D has been shown to be essential for callus formation. They observed that the maximum callus growth on MS medium containing 2,4-D in contrast to our present findings. The combination of Auxins and Cytokinins promote cellular differentiation and also organogenesis. In the medium concentration of BAP was increased up to 20-30 mg/l the multiple number of shoots Direct In Vitro plant propagation was achieved in pusa ruby cucutivar of Tamato (Solanum lycoperiscan L Mill)T. Ugender, Venkateshwarlu M (2018).

MATERIAL AND METHODS:

In the present investigation we present the result of our efforts to develop a protocol for plant regeneration through stem node explants in *Citrullus Valgaris L* a medicinally important plant. Stem node explants of different sizes were cultured with the induction medium consisting of MS Salts and Vitamins 6% Sucrose supplemented with BAP, 1.0mg/1 to 4.0mg/1 NAA (1.0mg1 I to 3.5mg/1) and 2,4-D (1.5mg1 I to 3.5mg/1). PH 5.7-5.8 The percentage of explants responding was evaluated after 4-6 weeks of cultures. The cultures were transferred to fresh medium after an internal of 6 weeks.

The nodal raised from control seeds could produce only callus on MS with different supplements which was regenerated into a single shoot. Well filled undamaged and uniform sized seeds were handpicked from the seed lot and equilibrated to the moisture content of 12 percent. For each dose of physical mutagen and random sample of 100 seeds were treated in *Citrillus Valgaris*. Explants were excised aseptically and were inoculated on the MS based medium supplemented by kinetin or BAP at concentrations ranging from 0.5 to 5 mg/l. Cultures were incubated under 10 h fluorescent light at $25 \pm 2^{\circ}$ C temperature

RESULTS AND DISCUSSION:

The maximum number of shoots on the explants were observed at 4.0 mg/l BAP or 5.0 mg/l kinetin, but at higher levels of BAP or kinetin, the formation of callus had taken place and the number of shoots per explant was reduced. The isolated *in vitro* raised shoots of 1-2 cm long, rooted profusely on MS medium with BAP (2 mg/l) + NAA (1 mg/l) within 15 days resulting in the formation of complete plantlets.

Table-1 . Tissue culture studies on stem node explants of *Citrillus Valgaris.*

| % of Growth regulators | Stem Node | |
|------------------------------------|---|---------------------------|
| (mg/l) | Mean number of shoots per shoot tip | % of callus production |
| MS+1.0BAP+1.0NAA+Kn2,4-D | 10.3±2.2 | 44 |
| MS+2.0BAP+1.5NAA+Kn2,4-D | 9.3±2.4 | 42 |
| MS+3.0 BAP+2.0 NAA+Kn2,4-D | 6.2 ± 1.6 | 30 |
| MS+4.0 BAP+3.0 NAA+Kn2,4-D | 1.4 ± 0.2 | 35 |
| MS+5.0BAP+3.5NAA+Kn2,4-D | 6.6 ± 1.3 | 25 |
| MS+2.0BAP+1.0L-glutamic acid | 12.4 ± 1.0 | 20 |
| MS+3.0BAP+1.0L-glutamic acid | 15.6 ± 0.5 | 18 |
| MS + 4.0 BAP + 1.0 L-glutamic acid | 14.4 ± 0.4 | 15 |

Plate 1. Tissue Culture studies on stem node explants of CitrullusValgaris L



www.worldwidejournals.com

PARIPEX - INDIAN JOURNAL OF RESEARCH | Volume-8 | Issue-12 | December - 2019 | PRINT ISSN No. 2250 - 1991 | DOI : 10.36106/paripex

CONCLUSION:

The stem node explant transferred to a fresh medium containing the some concentration of growth regulators, again resulted in the formation of multiple shoots.

REFERENCE:

- Ugender T, Odelu G, Ayodhya ramulu, Ch Anitha Devi U, Sammaiah D and 1. Venkatedhwarlu M(2016). Plant Regeneration via Callus induction from leaf explants of Solanum Surrattense Burm (F) a Medicinally important plant. European Journal of Biomedical and Pharmaceutical Sciences : Vol: 3 Issue : 3 161-166
- 2. Rastogi and Dwivedi, U.N. (2006) Down regulation of lignin biosynthesis in transgenic Leucaena leucocephala harbouring 0-methyl transferase gene. Biortechnol. Prog-22, 609-616.
- Venkateshwarlu Mandaloju(2017) Embryogenic Callus induction and plant 3. let proliferation of Soanum nigrum L. Through leaf explants European Journal of Biomedical and Pharmaceutical Sciences. Vol:4, issue:9, 582-588
- Venkateshwarlu M, Odelu G, Babitha kumara D, N Raju and Ugender T. (2018) 4. Studies in the Phytochemical analysis and biological activities of leaves of Solanum surattense Burm f A medicinally important plant Bioscience Discovery.9(1):1144-121 jan-2018.
- 5. T Ugender, M Venkateshwarlu, G Odelu, B Rajendra Prasad and Anitha Devi (2018) In Vitro plant let regeneration of tamato (Solanum Lycoperiscan Mill. CV. Pusa Ruby (from shoot tip explants using five cytokinins J. Indian bot.Soc Vol:97, (3 & 4): 138-145
- Ricci A Carra A, Torelli A, Maffiali CA, Vicini P, Zani F and Branca C (2001) 6. Cytokinin like activity of N-Substituted N-Phenylureas plant growth Regulated 34(2) 167-172
- 7. Hussain VS, Prakash DP and Asokan R (2007) effects of explants, preconditioning and light on the regeneration of transgenic tamato. Indian J. Hort 64(1):104-105
- 8. Murashige, J. (1974). Plant propagation through tissue culture Annu. Rev. Plant Physiol. 25: 135-166.
- 9. Bottino, P.J. (1975). The Potential of Genetic manipulation in plant cell cultures for plant breeding. Rad Bot. 15: 1-16.
- King, P.J. (1984). Mutagenesis in cultured cells. In cell culture and somatic 10. cell Genetics vol. 1. (ed) Vasil L.K., Laboratory Procedure and their application. Acad, Press Orlando. Pp. 547-551.
- Ahmed and Pande, (1988). Shoot multiplication was obtained from hypocotyls and cotyledon explants of Niger, Bionature 8:95. 11.
- 12. Gautam, A.S., Sood, K.C and Richaria, A.C. (1991). Mutagenic effectiveness and efficiency of gamma rays, EMS and their synergistic effects in black gram Vigna mungo L. Cytologia 57:85-89.
- Singh Rochica (2005). Studies on some larger fungi of faizabad with reference 13. to their eco-physiological characteristics. Ph.D. Thesis, Dr. R.M.L. Avadh. University, Faizabad. Pp. 114-115. Ma, J.F., Higashitani, A. Sato, K and Takeda, K (2002). Genotypic variation in
- 14.
- Silicon concentration of barley grains. Plant and Soil. 249(3):83-387.
 Rodrigues, F.A., Benhamou, N., Datnoff, L.E., Jones, J.B. and Belanger, R.R. (2003). Ultra structure and cytological aspects of silicon mediated rice blast 15. (2006). Only a structure and cytological aspects of shoot mediated rice blast resistance. Phytopathology, 93:535-546. Ahamed John, S. (1991). Mutation studies in black gram (Vigna Mungo (L)
- 16. Hepper, Ph.D., Thesis, Bharathidasan University, Thiruchirapalli.