



**ORIGINAL RESEARCH PAPER**

**Biological Science**

**HRLCMS ANALYSIS OF MORUS NIGRA L. AND CISSUS QUADRANGULARIS L. FOR IDENTIFICATION OF ANTIOXIDANT COMPOUNDS**

**KEY WORDS:** DPPH, Antioxidant, Morus nigra and Cissus quadrangularis, ascorbic acid.

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**ABSTRACT**

The antioxidant activity of leaves extracts of Morus nigra and Cissus quadrangularis was evaluated by free radical scavenging activity using 1, 1-diphenyl-2-picrylhydrazil (DPPH), reducing power assay. The results of the assay were then compared with a natural antioxidant ascorbic acid (vitamin C). The Methanolic extract of the Leaves of Morus nigra and Cissus quadrangularis is a potent source of compounds with antioxidant properties while the extract also exhibited significant free radical scavenging activity. The methanolic extract of Morus nigra and Cissus quadrangularis was subjected to preliminary phytochemical studies. The results indicate the presence of alkaloids, flavonoids, terpenoid proteins, and carbohydrates.

**INTRODUCTION**

Reactive oxygen species and free radicals play an important role in the initiation and evolution of numerous diseases. The use of compounds with antioxidant property is expected to be useful for the treatment of these diseases. Therefore, there has been a growing interest in finding novel antioxidants in order to meet the requirements of pharmaceutical industries. From classical period humankind has always been interested in naturally occurring components from plants and animals sources. Various extracts of different plant parts have been widely used in folk medicines and perfumes as well as in food flavor, preservatives and preparation and are more commonly utilized in chronic as well as common diseases. Plants contain several active compounds known as bioactive compounds such as alkaloids, steroids, tannins, glycosides, volatile oils, fixed oils, resins, phenols and flavonoids that are deposited in their different parts.

**MATERIAL AND METHODS**

**PLANT MATERIAL**

Disease free leaves were collected and identified by following the flora of Marathwada by V.N.Naik. The collected leaves were surface sterilized with 0.1% mercuric chloride & then washed with D/W 2-3 times separately & shade dried. Fine powder were made after complete drying and used for the experimental work.

**SOLVENT EXTRACTION OF LEAVES**

Extracts were made in 80% methanol at room temperature by simple extraction method (Deshpande et al). 10 gm dried powder of leaves mixed with 100ml solvent in 250 ml flask and were kept on shaker for 24 hrs. Then it was allowed to stand for the 30 min to stand the plant material. Thereafter it was filtered & centrifuged at 5000 rpm for 15 min .The supernatant was collected & solvent was evaporated at 45 OC in rotary evaporator to make the final volume 1/5 of the original volume.

**THIN LAYER CHROMATOGRAPHY-**

TLC is the most commonly used planar chromatographic method in natural product research. This is the easiest and cheapest technique and can be applied in the analysis, isolation and setting the parameters for column chromatography [9]. Usually, silica or alumina (more polar) is used as the stationary phase and organic solvents (less polar) are used as the mobile phase. This situation is categorized as normal phase chromatography. In contrast to this, reverse phase TLC is available, in which stationary phase is alkyl bonded silica or alumina (less polar) and mobile phase is polar solvent like water, alcohol etc.

**DPPH FREE RADICAL SCAVENGING ACTIVITY -**

The free radical scavenging activity was followed by the DPPH method. 0.1 mM solution of DPPH in methanol was prepared

and 1.0 ml of this solution was added to 3.0 ml of extract solution in methanol at different concentration (50-250 µg/ml). Thirty minutes later, the absorbance was measured at 517 nm. A blank was prepared without adding extract. Ascorbic acid at various concentrations (50 to 250 g/ml) was used as standard. Lower the absorbance of the reaction mixture indicates higher free radical scavenging activity. The capability to scavenge the DPPH radical was calculated using the following equation:

DPPH Scavenged (%) = [(A control - A test) / A control] × 100  
Where A control is the absorbance of the control reaction and A test is the absorbance in the presence of the sample of the extracts. The antioxidant activity of the methanolic leaves extract was expressed as IC50 and compared with standard. The IC50 value was defined as the concentration (in g/ml) of extracts .That scavenges the DPPH radicals by 50%.

**HRLC-MS ANALYSIS:** At SAIF (Sophisticated Analytical Instrument Facility), IIT Powai, Mumbai, equipment and conditions Identification of metabolites from an active sub-fraction of methanol extract was carried out. Samples were analyzed on a LC-ESI-Q-TOF-MS (Agilent Technologies 6550 i-Funnel) system equipped with a G4220B pump, G4226A auto sampler and G1316C, and a diode array detector (DAD). The elution solvent consisted of a gradient system of 0.1% formic acid in water (A) and acetonitrile (B) at a flow rate of 0.3 ml/min. The gradient system started with 95% A: 5% B reaching 5% A: 95% B in 50 min. then back to initial composition 95% A: 5% B in 10 min which was held at same composition for 5 min. The MS analysis was carried out by ESI positive ionization mode. HRLC-MS gives the presence of fatty acids, organic compounds, phenolics, alkaloids, phytoharmane, coenzyme, aminopyrimidines, dipeptide and tripeptides like important metabolites in leaves. For HRLC the Column used---zorbax eclipse c18, 2.1 x 150mm 5-micron.

**RESULTS -**

DPPH free radical has strong electron attracting ability from antioxidant. The graph showed that the conc. of methanolic extract is directly proportional to Absorbance. The Maximum scavenging activity is shown at the lowest conc. 200µl/ml which was 3.50 %. The order of five scavenging activity is 50>100>150>200<250 µl/ml. (Table 1.) The percentage of scavenging activity for methanolic extract of Morus nigra and Cissus quadrangularis L. leaves is inversely proportional to absorbance .Ascorbic acid was used as control. The TLC analyzed result of methanolic extract of Morus nigra and Cissus quadrangularis L with mobile phase Chloroform: Methanol: ethyl acetate (8.0:1.5:1.0). Mean of the Rf values of TLC analysis is 0.57. The HRLCMS results also revealed that the presence of antioxidant like Carnitine, Racepinephrine ,Swetenine, Apiin is a natural flavonoid, a diglycoside of the flavone apigenin .Vigabatrin also present in methanolic

extract and posses antioxiand and anti-epileptic property as gabapentin .The anti-epileptic drug vigabatrin was developed as an inhibitor of gamma-aminobutyric acid transaminase, and its ability to increase inhibition in the central nervous system led to its testing in an animal model. In animal models chronic use of vigabatrin is associated with irreversible myelin vacuolation. Antioxidant drugs change the antioxidant capacity of the body. Oxidative stress of the body increased when valproic acid and carbamazepine were used chronically (Mijiga n Cengiz et al , (2005). probenecid were highly effective at reducing oxidative stress detected by dichlorofluorescein fluorescence. Lina Du et al (2016).

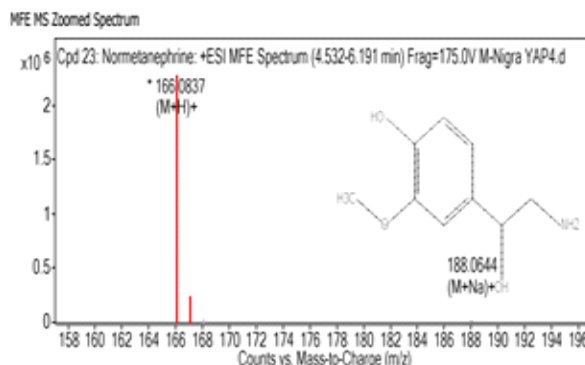
**DISCUSSION -**

Antioxidant assay revealed that the free radical scavenging activity showed by extract at different concentration. The maximum %scavenging activity at 250 µl/ml for both the plant extract was 72 % and 60 % and minimum activity at 50 µl/ml was 56 % and 48 % which was nearer to the standard activity of ascorbic acid. The reduction capability of DPPH radicals was determined by the decrease in its absorbance at 517 nm, which is induced by antioxidants. Table 1 shows the percentage of DPPH radical scavenged by ascorbic acid and methanolic extract of leaves at various concentrations ( g/ml). A substance may act as an antioxidant due to its ability to reduce reactive oxygen species by donating hydrogen atom.

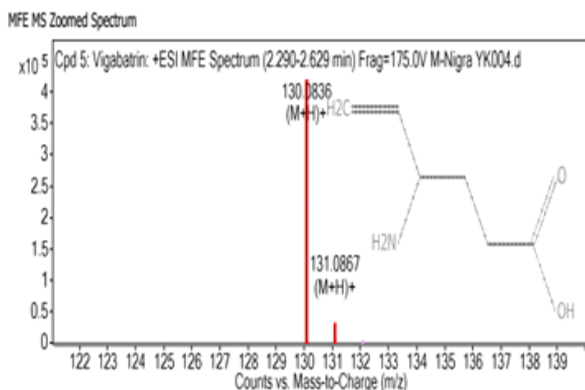
The reducing property of methanolic leaves extract of *Morus nigra* and *Cissus quadrangularis* L. implies that it is capable of donating hydrogen atom in a dose dependent manner. The high content of phenolic compounds in the extract may be a contributing factor towards antioxidant activity because the phenolic compounds are known to have direct antioxidant property due to the presence of hydroxyl groups, which can function as hydrogen donor. Research in the direction of partial isolation and characterization of the constituents of methanolic leaves extract of *Morus nigra* and *Cissus quadrangularis* L. in order to decipher the specific phytochemical constituent(s) responsible for the free radical scavenging activity of the plant. When this is done, extracts of *Morus nigra* and *Cissus quadrangularis* L, L. could find important application in phytotherapy. Qualitative

|   |     |     |     |     |
|---|-----|-----|-----|-----|
| 1 | 50  | 48% | 56% | 55% |
| 2 | 100 | 52% | 59% | 60% |
| 3 | 150 | 55% | 62% | 65% |
| 4 | 200 | 58% | 68% | 70% |
| 5 | 250 | 60% | 72% | 75% |

**Chromatograms of *Morus nigra* L. through HR-LCMS analysis.**



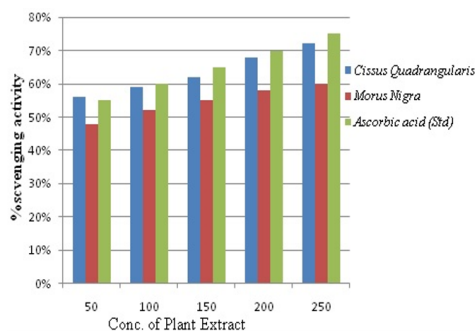
**MS Compound Spectrum With Structure**



**MS Compound Spectrum With Structure**

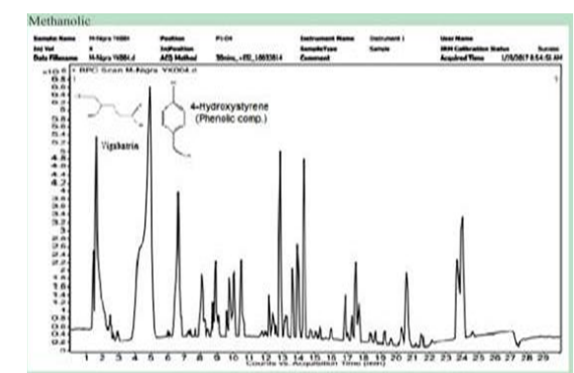
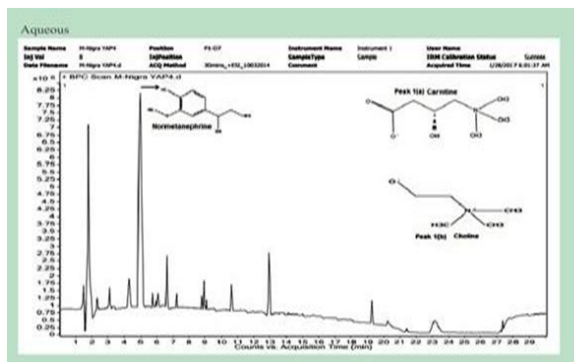
**Compound Report**

|                                       |                              |                      |                      |
|---------------------------------------|------------------------------|----------------------|----------------------|
| <b>Data File</b>                      | M-Nigra YAP4.d               | <b>Sample Name</b>   | M-Nigra YAP4         |
| <b>Sample Type</b>                    | Sample                       | <b>Position</b>      | P1-D7                |
| <b>Instrument Name</b>                | Instrument 1                 | <b>User Name</b>     |                      |
| <b>Acq Method</b>                     | 30mins_+ESI_10 032014_MSMS.m | <b>Acquired Time</b> | 1/28/2017 6:01:37 AM |
| <b>IRM Calibration Status Comment</b> | Success                      | <b>DA Method</b>     | default.m            |



**Table 1- Scavenging activity of MeoH leaf extracts**

| Sr.no. | Conc. of extract | Percent scavenging activity |                       |                     |
|--------|------------------|-----------------------------|-----------------------|---------------------|
|        |                  | Morus Nigra                 | Cissus Quadrangularis | Ascorbic acid (Std) |
| 1      | 50               | 48%                         | 56%                   | 55%                 |
| 2      | 100              | 52%                         | 59%                   | 60%                 |
| 3      | 150              | 55%                         | 62%                   | 65%                 |
| 4      | 200              | 58%                         | 68%                   | 70%                 |
| 5      | 250              | 60%                         | 72%                   | 75%                 |



| RT     | Mass     | Name                               | Formula       | DB Diff (ppm) |
|--------|----------|------------------------------------|---------------|---------------|
| 1.677  | 104.1092 | Choline                            | C5 H14 N O    | 16.35         |
| 1.776  | 133.0741 | 1,4-Dideoxy-1,4-Imino-D-Arabinitol | C5 H11 N O3   | 1.58          |
| 1.823  | 162.1108 | Carnitine                          | C7 H16 N O3   | 13.68         |
| 4.327  | 145.1101 | 2S-aminoheptanoic acid             | C7 H15 N O2   | 1.38          |
| 6.63   | 183.089  | Racpinephrine                      | C9 H13 N O3   | 2.84          |
| 9.095  | 186.1612 | 4-methyl-decanoic acid             | C11 H22 O2    | 4.4           |
| 12.959 | 563.3116 | dihydroergocornine                 | C31 H41 N5 O5 | 1.54          |
| 12.96  | 568.267  | Swietenine                         | C32 H40 O9    | 0.39          |
| 19.267 | 452.2932 | Dihydrocelastrol                   | C29 H40 O4    | 1.09          |

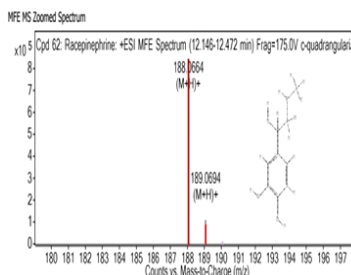
| RT     | Mass     | Name   | Formula         | DB Diff (ppm) |
|--------|----------|--|-----------------|---------------|
| 1.833  | 133.0728 | 1,4-Dideoxy-1,4-Imino-D-Arabinitol           | C5 H11 N O3     | 8.58          |
| 2.428  | 129.0763 | Vigabatrin                                   | C6 H11 N O2     | 10.46         |
| 2.668  | 285.1016 | Probenecid                                   | C13 H19 N O4 S  | 6.5           |
| 4.895  | 120.0585 | 4-Hydroxystyrene                             | C8 H8 O         | 8.03          |
| 6.628  | 183.0874 | Racpinephrine                                | C9 H13 N O3     | 11.67         |
| 6.633  | 113.0836 | epsilon-Caprolactam                          | C6 H11 N O      | 3.88          |
| 6.75   | 141.0778 | Ethosuximide                                 | C7 H11 N O2     | 8.31          |
| 10.149 | 513.2695 | Sulfolithocholylglycine                      | C26 H43 N O7 S  | 12.75         |
| 10.491 | 141.0778 | Ethosuximide                                 | C7 H11 N O2     | 8.17          |
| 10.492 | 113.0834 | epsilon-Caprolactam                          | C6 H11 N O      | 5.57          |
| 10.567 | 200.0771 | Barbituric acid, 5-ethyl-5-(2-hydroxyethyl)- | C8 H12 N2 O4    | 13.24         |
| 10.568 | 94.0076  | Dimethyl sulfone                             | C2 H6 O2 S      | 13.4          |
| 11.278 | 395.2027 | Spiperone                                    | C23 H26 F N3 O2 | 4.5           |
| 11.715 | 186.1599 | 3-methyl-decanoic acid                       | C11 H22 O2      | 11.37         |
| 12.874 | 563.3085 | dihydroergocornine                           | C31 H41 N5 O5   | 4.05          |
| 12.875 | 568.2636 | Swietenine                                   | C32 H40 O9      | 6.39          |
| 13.965 | 326.203  | Hydroquinine                                 | C20 H26 N2 O2   | 10.81         |
| 19.255 | 452.2904 | Dihydrocelastrol                             | C29 H40 O4      | 5.03          |
| 25.233 | 586.2709 | Khivorin                                     | C32 H42 O10     | 11.82         |

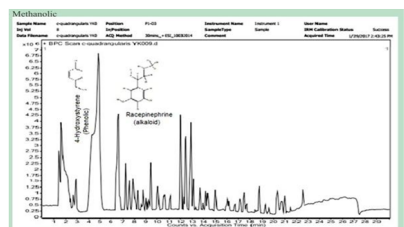
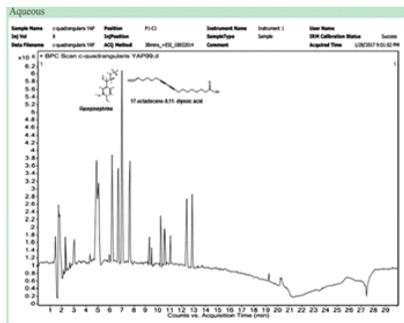
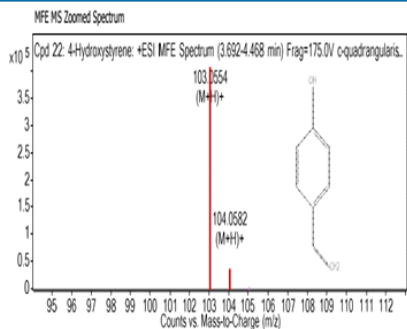
|        |          |                            |               |       |
|--------|----------|----------------------------|---------------|-------|
| 2.241  | 116.0581 | 1^--Diacetylhydrazine      | C4 H8 N2 O2   | 4.15  |
| 2.786  | 116.0832 | 2-methyl valeric acid      | C6 H12 O2     | 4.17  |
| 5.066  | 138.0891 | nn-Dimethylaniline-n-oxide | C8 H12 N O    | 20.48 |
| 6.701  | 183.0876 | Racpinephrine              | C9 H13 N O3   | 10.41 |
| 9.287  | 188.1162 | Acetyllysine               | C8 H16 N2 O3  | 0.47  |
| 10.265 | 159.1246 | DL-2-Aminooctanoic acid    | C8 H17 N O2   | 8.36  |
| 12.875 | 563.309  | Dihydroergocornine         | C31 H41 N5 O5 | 3.22  |

|        |          |                            |                |       |
|--------|----------|----------------------------|----------------|-------|
| 12.876 | 568.2643 | Swietenine                 | C32 H40 O9     | 5.1   |
| 19.267 | 452.2909 | Dihydrocelastrol           | C29 H40 O4     | 3.98  |
| 2.671  | 285.102  | Probenecid                 | C13 H19 N O4 S | 5.15  |
| 4.288  | 120.0586 | 4-Hydroxystyrene           | C8 H8 O        | 9.32  |
| 6.607  | 113.0838 | epsilon-Caprolactam        | C6 H11 N O     | 2.52  |
| 6.612  | 121.0881 | Phenylethylamine           | C8 H11 N       | 8.46  |
| 6.614  | 183.0878 | Racpinephrine              | C9 H13 N O3    | 9.63  |
| 6.734  | 141.0781 | Ethosuximide               | C7 H11 N O2    | 6.23  |
| 7.159  | 418.1765 | Nimodipine                 | C21 H26 N2 O7  | 5.86  |
| 7.576  | 121.0881 | Phenylethylamine           | C8 H11 N       | 9.02  |
| 7.665  | 164.0935 | Isonicotinamide, 2-propyl- | C9 H12 N2 O    | 8.66  |
| 7.914  | 564.1392 | Apiin                      | C26 H28 O14    | 15.4  |
| 8.279  | 188.116  | Acetyllysine               | C8 H16 N2 O3   | 0.62  |
| 8.28   | 128.1193 | Octanal                    | C8 H16 O       | 6.23  |
| 10.535 | 244.0799 | 5-azacytidine              | C8 H12 N4 O5   | 3.47  |
| 12.861 | 563.309  | dihydroergocornine         | C31 H41 N5 O5  | 3.16  |
| 12.862 | 568.264  | Swietenine                 | C32 H40 O9     | 5.69  |
| 15.946 | 408.2411 | Dextromoramide M2          | C25 H32 N2 O3  | 0.48  |
| 16.035 | 444.1975 | Mitoxantrone               | C22 H28 N4 O6  | 7.6   |
| 19.258 | 452.2908 | Dihydrocelastrol           | C29 H40 O4     | 4.13  |
| 20.71  | 286.2475 | Avocadene                  | C17 H34 O3     | 11.45 |

**CONCLUSION**

Oxidative stress can arise from overproduction of ROS by metabolic reactions that use oxygen and shift the balance between oxidant/antioxidant statuses in favor of the oxidants. ROS are produced by cellular metabolic activities and environmental factors, such as air pollutants or cigarette smoke. ROS are highly reactive molecules because of unpaired electrons in their structure and react with several biological macromolecules in cell, such as carbohydrates, nucleic acids, lipids, and proteins, and alter their functions. ROS also affects the expression of several genes by upregulation of redox-sensitive transcription factors and chromatin remodeling via alteration in histone acetylation/deacetylation. Regulation of redox state is critical for cell viability, activation, proliferation, and organ function. Experiments confirming these activities of the extract of *Morus nigra* and *Cissus quadrangularis* in an in vivo system would be necessary. However, herbal remedies often do not produce any side effects. Therefore, alternative medicine become popular remedy to various types of ailments. In conclusion, *Morus nigra* and *Cissus quadrangularis* extracts have revealed significant antibacterial activities against test organisms used for the study.





Chromatograms of *Cissampelos quadrangularis* L. through HR-LCMS analysis.

16. Mohammed Golam Rasul Extraction, Isolation and Characterization of Natural Products from Medicinal Plant International Journal of Basic Sciences and Applied Computing 2018; 2 (6) 1-6.
17. Morakinyo AO, Oludare GO, Aderinto OT, Tasdup A Biology and Medicine Antioxidant and free radical scavenging activities of aqueous and ethanol extracts of *Zingiber officinale* 2012; 3 (5): 25-30.
18. Sai Koteswara D. Sharma, Phytochemical and Antimicrobial Activity of Whole Plant of *Madhuca indica* International Journal of Research in Pharmacy and Chemistry 2013, 3(1): 15-19.
19. V. Bulugahapitiya, Plant Based Natural Products Extraction and Phytochemical analysis, self, 2013. <https://www.researchgate.net/publication/324136685>.

**REFERENCES**

1. Adhikarimayum haripyaree, Kshetrimayum guneshwor, Maibam damayanti Evaluation of Antioxidant Properties of Phenolics Extracted from *Ananas comosus* L. *Notulae Scientia Biologica*: 2010; 2 (2), 68-71.
2. Akash P. Dahake1 Antioxidant activity of methanolic extract of *Madhuca indica* bark, *Journal of Pharmacy Research* 3(8), 2010; 1709-1711.
3. Aliyul A. B, Ibrahim M. A, Musa A. M, Ibrahim H., Abdulkadir I. E and Oyewale A. O *Journal of Medicinal Plants Research* Evaluation of antioxidant activity of leave extract of *Bauhinia rufescens* Lam. (Caesalpinaceae) 2009; Vol. 3(8), 563-567.
4. Aruoma I.O. Antioxidant action of action of plant foods. Use of oxidative DNA damage , as a tool of Studying Antioxidant efficacy. *Free Radical Res.* 1999; 30: 419-427.
5. Aruoma I.O. and Cuppette S.L. *Antioxidant Methodology: In vivo and Vitro concepts* IL 1997; AOAS Press.
6. Climpoiu C; Analysis of Some Natural Antioxidants by Thin-Layer Chromatography and High Performance Thin-Layer Chromatography *Journal of Liquid Chromatography & Related Technologies*, 2006; (9) 7-8.
7. Hanspeter r. witschi and David g. doherthy Butylated Hydroxyanisole and Lung Tumor Development in A/J Mice *Toxicol. Sci.* 1984; 4 (5): 795-801
8. Ho-min kang and Mikal e. saltveit, Antioxidant Enzymes and DPPH-Radical Scavenging Activity in Chilled and Heat-Shocked Rice (*Oryza sativa* L.) Seedlings *Radicles, J. Agric. Food Chem.* 2002; 50, 513-518
9. Kalaivani.M and Jegadeesan.M Antimicrobial activity of alcoholic extract of leaves and flowers of *Madhuca indica* International Journal of Scientific and Research Publications, 2013; 3(5) 1-3.
10. Kaushik. Evaluation of antioxidant and antimicrobial activity of *madhuca indica* *pharmacology online* 2010; vol 2: 1-8.
11. M.S.F. Lie ken jie Novel halo-oxo-allenic fatty ester derivatives from epoxidized methyl santalbate *Science Direct Elsevier .chemistry and physics of lipids*, 2003; (125), 93-101.
12. Sheetal Anandjiwala, Honnegowda Srinivasa, Jyoti Kalola, Mandapati Rajani Free-radical scavenging activity of *Bergia suffruticosa* (Delile) Fenzl *Journal of Natural Medicines*, 2007; (61) 1, 59-62.
13. Suprava Sahoo, Goutam Ghosh and Sanghamitra Nayak *Journal of Medicinal Plants Research* Evaluation of in vitro antioxidant activity of leaf extract of *Alpinia malaccensis*, 2012; Vol. 6(23), 4032-4038.
14. Lina Du, Philip E Empey Jing Ji, Honglu Chao, Patrick M Kochanek, Hülya Bayır, Robert S B Clark Probenecid and N-Acetylcysteine Prevent Loss of Intracellular Glutathione and Inhibit Neuronal Death after Mechanical Stretch Injury *In Vitro J Neurotrauma* doi: 10.1089/neu.2015.4342. Epub 2016 Mar 22.. 2016 Oct 15; 33(20): 1913-1917.
15. Miijga n Cengiz, Adnan Yiksel, Ahmet Özyaydin, Anil Özkiliç, Ümran etinel and Mehmet The Effects of Vigabatrin on Rat Liver Antioxidant Status, *Seven* Vol. 21, No. 2, 2005 109-115.